

A Preliminary Investigation of the Jack-Bean Urease Inhibition by Randomly Selected Traditionally Used Herbal Medicine

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Abstract

Helicobacter pylori (*H. pylori*) infection leads to different clinical and pathological outcomes in humans, including chronic gastritis, peptic ulcer disease and gastric neoplasia and even gastric cancer and its eradication depends upon multi-drug therapy. The most effective therapy is still unknown and prompts people to make great efforts to find better and more modern natural or synthetic anti-*H. pylori* agents. In this report 21 randomly selected herbal methanolic extracts were evaluated for their effect on inhibition of Jack-bean urease using the indophenol method as described by Weatherburn. The inhibition potency was measured by UV spectroscopy technique at 630 nm which attributes to released ammonium. Among these extracts, five showed potent inhibitory activities with IC₅₀ ranges of 18-35 µg/mL. These plants are *Matricaria disciforme* (IC₅₀:35 µg/mL), *Nasturtium officinale* (IC₅₀:18 µg/mL), *Punica granatum* (IC₅₀:30 µg/mL), *Camelia sinensis* (IC₅₀:35 µg/mL), *Citrus aurantifolia* (IC₅₀:28 µg/mL).

Keywords: Herbal extract; Urease; Inhibitor; Indophenol method; Lead discovery.

Introduction

Many hundreds of plants worldwide are used in traditional medicine as treatments for different kinds of diseases including bacterial infections and gastrointestinal disorders. Among these bacteria, *H. pylori*, a Gram-negative pathogenic bacterium which specifically colonizes the human gastric mucosa, has been regarded as a primary causative agent of chronic gastritis and peptic ulcer diseases including mucosa-associated lymphoid tissue lymphoma (1). Conventional multiple drug therapy in management of *H. pylori* infection usually provide effective therapy but there is an increasing problem of

antibiotic resistance, side effects and significant cost of therapy which associate with these kinds of drugs (2-4).

While *H. pylori* is acid sensitive and only replicates at pH of 7-8, it survives in the stomach under highly acidic conditions (5-7) urease activity in bacteria is believed to be essential for the colonization of and survival of *H. pylori* at very acidic pH (8, 9). Thus virulence of *H. pylori* could be controlled using chemicals that inhibit urease activity.

Ureases (E.C 3.5.1.5), the first enzyme crystallized from Jack bean (*Canavalia ensiformis*) was shown to contain nickel ions (10) which rapidly catalyzes the hydrolysis of urea to form ammonia and carbon dioxide (11) have been shown to be an important virulence determinant in the pathogenesis of many clinical

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conditions, which is detrimental for human and animal health as well as for agriculture (12). The product, ammonia, of such decomposing reactions diffuses across the cytoplasmic membrane, buffering the periplasmic space and allows growth in the presence of extracellular gastric acid (13), and responsible for negative effects of urease activity in human health (14), such as causing peptic ulcers, stomach cancer, etc. Besides, in agriculture the efficiency of soil nitrogen fertilization with urea decreases due to ammonia volatilization and root damage caused by soil pH increase (15).

Then, it's interesting to control the activity of urease through the use of its inhibitors in order to counteract these negative effects in medicine, environmental and agronomic. Many urease inhibitors have been described in the past decades, such as phosphorodiamidates (16), α -hydroxyketones (17), Polyhalogenated benzo- and naphthoquinones (18) and imidazoles such as proton pump inhibitors of lansoprazole, rabeprazole and omeprazole (19).

Natural urease inhibitors from *Euphorbia decipiens* (20) and sulfated polysaccharide found mainly in various species of brown seaweed (fucoïdan compounds) had been reported previously (21). The use of some chemical or herbal compounds were banned in vivo or entering clinical trials because of their toxicity, chemical or physical instability or low bioavailability (22).

Thus, seeking novel and efficacious urease inhibitors with good bioavailability and low toxicity are significant especially in low income countries with high infection rate of *H. pylori* is desirable. This report is focused on seeking for novel natural urease inhibitors from herbal sources that can be used directly or as a lead compounds in management of *H. pylori* infection.

Experimental

Plant extraction and preparation of extracts

Twenty one medicinal plants which are listed in Table 1, are obtained randomly from local herbal market in September 2010, Tehran and identified by one of authors, Dr. F. Mojab and were evaluated against Jack-bean urease. Each

plant sample was individually powdered and 1 g was extracted by maceration method using aqueous methanol (10 mL; 50:50 v/v) as solvent for 24 h. Each extract was filtered, concentrated under reduced pressure to dryness and stored at 0°C until time of analysis. The percentage of inhibition at 1000 μ g/mL concentration of extracts, dissolved in same solvent was accurately defined.

Chemicals

All the chemicals used were of analytical grade from Merck Co., Germany. All aqueous solutions were prepared in MilliQ (Millipore, USA) water. Jack-bean urease was obtained from Merck (5 units/mg).

Urease inhibition activity assay

For urease inhibition assays after addition of 10 mL of phosphate buffer to accurately weight of enzyme, sonication was performed for 60s, followed by centrifugation and evaluating absorbance of upper solution in $\lambda = 280$ nm which is attributed to enzyme. By using the following equation $A = \epsilon bc$ where c is the concentration of solution (mol/L), b is the length of the UV cell and ϵ represents molar absorptivity in the specific wavelength, we can calculate the concentration of initially urease solution. After proper dilution, the concentration of enzyme solution adjusts at 2 mg/mL.

The assay mixture, containing 100 μ L (2 mg/mL) of Jack-bean urease and 100 μ L of the test compound with 0.2 mL of 100 mM phosphate buffer pH 6.8 containing 25 mM urea was pre-incubated for 30 min in water bath at 37°C. The urease reaction was stopped after 30 min incubation with 600 μ L of 4% H₂SO₄ acid. Enzyme inhibition activity performed by Berthelot alkaline phenol-hypochlorite method to examine the efficiency of adsorptive immobilization. This method is based on the released ammonia (NH₃) which reacts with hypochlorite (OCl⁻) to form a monochloramine (23). This product then reacts with phenol to form blue-colored indophenols whose absorbance is measured at 625 nm.

The liberated ammonia was estimated using 500 μ L of solution A (contained 5.0 g phenol and 25 mg of sodium nitroprusside) and 500 μ L of

Table 1. Name of the plants, part used in traditional medicine and percent of inhibition of urease enzyme in presence of 1 mg/mL of each herbal extract.

	Scientific name	Common name in English	Common name in Persian	Part used	Percent of inhibition (%)*
1	<i>Alhagi maurorum</i>	Camel thorn	Khar-e shotor	resin	32.0 ± 1.1
2	<i>Boswellia carterii</i>	Frankincense	Kondor	resin	11.3 ± 1.9
3	<i>Camelia sinensis</i>	Tea shrub	Chai	leaf	95.4 ± 2.40
4	<i>Cerasus avium</i>	Cherry tail	Dom-e gilās	tail	26.6 ± 2.90
5	<i>Citrus aurantifolia</i>	Basra lime	Limu Omani	fruit	97.6 ± 0.78
6	<i>Citrullus colocynthis</i>	Bitter apple	Hanzal	Fruit	70.1 ± 0.91
7	<i>Cotoneaster nummularia</i>	Pockspary manna	Shirkhesht	resin	10.3 ± 0.47
8	<i>Fraxinus velutina</i>	Velvet Ash	Zaban-e ghonjeshk	leaf	16.4 ± 0.55
9	<i>Laurus nobilis</i>	Grecian laurel	Bargh-e bu	leaf	68.8 ± 1.63
10	<i>Matricaria recutita</i>	Chamomile	Babun-e shirazi	flower	88.8 ± 0.99
11	<i>Nardostachys jatamansi</i>	Spikenard	Sonboletib	rhizomes	0
12	<i>Nasturtium officinale</i>	Watercresses	Bolagooti	leaf	99.1 ± 1.78
13	<i>Nepeta bracteata</i>	Catmint	Zufa	flower	21.4 ± 2.24
14	<i>Nepeta menthoids</i>	French lavender	Stoqodus	branch	26.6 ± 1.43
15	<i>Physalis alkekengi</i>	Winter cherry	Arusak-eposht-e pardeh	fruit	72.2 ± 1.88
16	<i>Polygonum aviculare</i>	Seresh	Alafe haft band	leaf	62.5 ± 3.10
17	<i>Punica granatum</i>	pomegranate	Golnar	flower	96.7 ± 2.55
18	<i>Salvia officinalis</i>	Sage	Maryam Goli	leaf	71.3 ± 0.22
19	<i>Sambucus nigra</i>	Black Elder	Agti	leaf	41.0 ± 1.28
20	<i>Trachyspermum copticum</i>	Ajwain	Zenyan	seed	38.8 ± 1.67
21	<i>Zea mays</i>	Corn Crest	Kakol-e dhorrat	noodle	22.2 ± 1.47

Values are expressed as mean ± SD of 3 experiments.

solution B (contained of 2.5 g sodium hydroxide and 4.2 mL of sodium hypochlorite in 500 mL of distilled water) at 37°C for 30 min and the absorbance was measured at 625 nm against the control. All reactions were performed in triplicate in a final volume of 1 mL. Percentage of inhibitions were calculated using the formula $(100 - (\text{OD sample} / \text{OD control}) \times 100)$.

The concentration of compounds that inhibited the hydrolysis of substrate by 50% (IC_{50}) was determined through monitoring the inhibition effect of various concentrations of extracts in the assay. The IC_{50} values were then calculated using above mentioned formula in the previous section.

Results and Discussion

As evidence in beneficial effects of medicinal plants traditionally used to manage different disorders, twenty one samples, are available

from local herbal and medicinal plants shop were examined against Jack-bean urease by Berthelot alkaline phenol-hypochlorite method and results revealed varied inhibitory activities (Table 1). Five extracts which showed maximum inhibitory effect ($\geq 90\%$ of enzyme inhibition) were selected and further studied for IC_{50} determination by UV-spectroscopy technique; the relevant data is presented in Table 2.

As shown in Figure 1, concentration-dependent activities against Jack-bean urease were observed between selected extracts and inhibitory effect increased together with increasing the concentration of each plant's extract in the range of (0-100 $\mu\text{g/mL}$).

As shown in Table 2, the inhibitory activities of five selected extracts were found to be the most potent inhibitors are: *Camelia sinensis* ($\text{IC}_{50} = 35 \mu\text{g/mL}$), *Citrus aurantifolia* ($\text{IC}_{50} = 28 \mu\text{g/mL}$), *Nasturtium officinale* ($\text{IC}_{50} = 18 \mu\text{g/mL}$), *Punica granatum* ($\text{IC}_{50} = 30 \mu\text{g/mL}$) and

Table 2. The IC₅₀ and the percent of enzyme inhibition in the presence of plants extract at concentration of 1 mg/mL.

Plant name	IC ₅₀ (µg/mL)	Percent of Inhibition (%)
<i>Camelia sinensis</i>	35 ± 1.9	95.38 ± 2.40
<i>Citrus aurantifolia</i>	28 ± 0.6	97.64 ± 0.78
<i>Matricaria recutita</i>	37 ± 1.6	88.88 ± 0.99
<i>Nasturtium officinale</i>	18 ± 1.4	99.13 ± 1.78
<i>Punica granatum</i>	30 ± 1.2	96.75 ± 2.55

Preincubation time was 0.5 h. IC₅₀ vales were calculated from urease inhibition rate of six doses, 10, 20, 40, 60 and 80 µg/mL in triplet.

Nasturtium officinale (IC₅₀ = 35 µg/mL).

Medicinal plants serve as a useful source of novel drugs (24). In developing countries, since the application of antibiotics is still under a poor management as a whole, there is a growing need for finding new medicinal plants especially anti-*H. pylori* agents that can help eradicate the invasion and presence of survived *H. pylori* strains to avoid relapse of gastric ulcer. In this regard, the literature has reported extracts of certain plants such as cashew apple (25), cinnamon (26), and Chinese tea (27) inhibit growth of *H. pylori* and some urease inhibitory activity.

According to reported investigations on *Camelia sinensis* (Green tea) its *H. pylori*'s urease inhibition has been proven (28). Achieving IC₅₀ equal to 35 µg/mL for *C. sinensis* in this study which is varied from previous researches (IC₅₀ = 13 µg/mL), is probably due to the diversity of growth, harvesting and extraction methods. Regarding the excess amount of flavon, flavol, catechin epigallocatechin gallate, gallic acid, gallic acid, gallic acid, and many other flavonoids in *C. sinensis* (28), we can conclude that urease inhibitory activity of black tea (this study) is attributed to the similar substances found in green tea extract in different amounts. The chemical constituents and functional groups of these compounds play an important role in inhibition of urease. The functional groups such as hydroxyl and ketones which are linked with aromatic rings can interact with Ni ions in active site of enzyme, resulting in its inhibition.

The Key lime (*Citrus aurantifolia*; Omani lime) is a citrus species with a globose fruit, 2.5-5 cm in diameter (1-2 in) that is yellow when ripe but usually picked green commercially. It is smaller, seedier, has a higher acidity, a stronger

aroma, and a thinner rind than that of the Persian lime (*Citrus x latifolia*). It is valued for its unique flavor compared to other limes, with the key lime usually having a more tart and bitter flavor. In Malaya, the juice is taken as a tonic and to relieve stomach ailments and it is given as a vermifuge in combination with oil. In India, the pickled fruit is eaten to relieve indigestion. It has been used as an antiseptic, tonic, an antiscorbutic, an astringent, and as a diuretic in liver ailments, a digestive stimulant, a remedy for intestinal hemorrhage and hemorrhoids, and as a disinfectant for all kinds of ulcers when applied in a poultice (29-33). In Iran dried fruits are usually consumed as vegetable and prevent indigestion.

The chemical composition of *C. aurantifolia* is well known and limonene, γ -terpinene, terpinolene and α -terpineol present in different amounts (32). Antibacterial activity of essential oil of lime is related to its composition (31) but none of those compounds could inhibit urease enzyme. However urease inhibitory of *C. aurantifolia* was found to be (IC₅₀ = 28 µg/mL). The methanolic extract of dried fruits may contain new substance which is not reported in literature and need more investigation on its isolation and identification.

Another plant which shows high inhibitory effect on urease is *Matricaria recutita* (*Chamomile*). Previously, it was shown that the *Chamomilla recutita* (*M. recutita*) oil extract is rich in fatty acids, coumarins, terpenes, spiroethers and flavonoids contributing to its medicinal properties (34, 35). Additionally, *Roman chamomile* sample showed high antimicrobial activity against all strains of tested microbes Gram-negative bacteria (*Escherichia coli*, *Pseudomonas aeruginosa*,

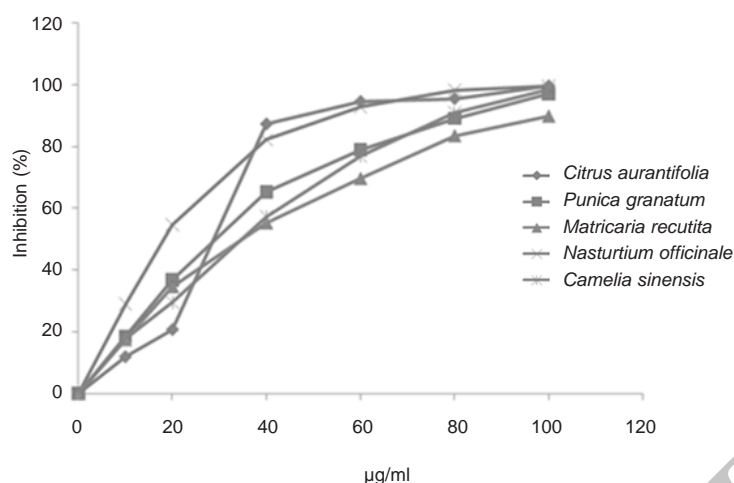


Figure 1. Inhibition profile of five selected extract against Jack-bean urease activity by indophenol method.

Proteus vulgaris, *Klebsiella pneumoniae* and *Salmonella sp.*) (36, 37).

The chamomile oil inhibited the *H. pylori*'s growth in very low concentrations of 0.0075% (v/v). The reported MIC₅₀ and MIC₉₀ of *Chamomilla recutita* (*M. recutita*) oil extract for *H. pylori* were 62.5 mg/mL and 125.0 mg/mL, respectively. In addition, it was found that urease production of *H. pylori* was inhibited by the *M. recutita* oil extract (35). This finding was of particular interest, since urease activity is critical for the survival of this microorganism in the stomach.

It was shown that the *M. recutita* oil extract influenced the morphological and fermentative properties of *H. pylori*. Thus, it is possible that these compounds (fatty acids, coumarins, terpenes, spiroethers and flavonoids) could be responsible for the morphological and fermentative changes and subsequent anti-*H. pylori* activity of the *M. recutita* oil extract. However, a 50 % aqueous methanol extract from *M. recutita* flowers was also found to be inhibitory against urease, which corroborates previously reported results (35). Both *M. recutita* oil extract or its methanolic extract may be useful as additional remedy in the complex treatment of stomach ulcers and duodenal intestinal diseases which are subjected to *H. pylori*, especially for patients with allergic responses to antibacterial drugs (35).

Watercresses (*Nasturtium officinale*) are

fast-growing, aquatic or semi-aquatic, perennial plants native from Europe to central Asia, and one of the oldest known leaf vegetables consumed by human beings. These plants are members of the family Brassicaceae or cabbage family, botanically related to garden cress and mustard with a peppery, tangy flavor due to high contents of phenethyl isothiocyanate (38). Many benefits from eating watercress are claimed, such as that it acts as a stimulant, a source of phytochemicals and antioxidants, an anti-microbial, a diuretic, an expectorant, and a digestive aid (38-40). The phenethyl isothiocyanate content of watercress inhibits hypoxia-inducible factors which can inhibit angiogenesis in lung cancer (41). The strong urease activity (IC₅₀=18 µg/mL) reported in this study among all other extracts, which reported for the first time, is concurrent with its daily usage as digestive aid in people whom suffering gastric upset. Phenethyl isothiocyanate may interact with urease Ni ions and inhibit its catalytic activity. More investigations are needed to confirm this hypothesis.

Another plant which shows positive effect on urease inhibition is *Punica granatum* (Pomegranate). Previous studies on pomegranate demonstrated its medicinal usage against *salmonella* (42). Anti-*helicobacter pylori* activity of aqueous and ethanolic extract of *punica granatum* pericarps are recently reported (43). The minimal inhibitory concentration (MIC) value was reported to be 0.78-6.25 mg/

mL, but the mechanism of this inhibition was not clearly investigated.

The medicinal plant possesses a high amount of tannin (25%). The antimicrobial properties of this substance were well established. Polar fraction of *P. granatum* was reported to contain ellagitannin and punicalagin (43). The strong urease inhibitory activity of *P. granatum* (IC_{50} 30 μ g/mL) reported here revealed that antibacterial properties of *P. granatum* against different strands of *H. pylori* could be at least partially due to inhibition of urease by interaction of ellagitannin and punicalagin with active site of enzyme or modulating its activity by aggregation properties of tannin.

Medicinal plants, traditional medicinal and other natural sources are still good source for lead discovery. The results of this study revealed that random screening of medicinal plants could lead to introducing new candidate for further studies which, in the end, can help and enhance human health.

Acknowledgment

This study was supported by a grant from the Research Council of Tehran University of Medical Sciences.

References

- Wotherspoon AC, Ortiz-Hidalgo C, Falzon MR and Isaacson PG. *Helicobacter pylori*-associated gastritis and primary B-cell gastric lymphoma. *Lancet* (1991) 338: 1175-1176.
- Jenks P and Edwards DI. Metronidazole resistance in *Helicobacter pylori*. *Int. J. Antimicrobial Agents* (2002) 19: 1-7.
- Kim IH, Lee DG, Lee SH, Ha JM, Ha BJ, Kim SK and Lee JH. Antibacterial activity of *Ulva lactuca* against methicillin-resistant *Staphylococcus aureus*. *Biotechnol. Bioprocess Eng.* (2007) 12: 579-582.
- Karen WM and Edzard E. Herbal medicines for treatment of bacterial infections: a review of controlled clinical trials. *J. Antimicrob. Chemother.* (2003) 51: 241-246.
- Stingl K, Altendorf K and Bakker EP. Acid survival of *Helicobacter pylori*: how does urease activity trigger cytoplasmic pH homeostasis? *Trend Microbiol.* (2002) 10: 70-74.
- Stingl K and De Reuse H. Staying alive overdosed: how does *Helicobacter pylori* control urease activity? *Int. J. Med. Microbiol.* (2005) 295: 307-315.
- Zhang L, Mulrooney SB, Leung AFK, Zeng Y, Ko BBC, Hausinger RP and Sun, H. Inhibition of urease by bismuth(III): Implications for the mechanism of action of bismuth drugs. *Biometals* (2006) 19: 503-511.
- Mobley HL, Hu LT and Foxal PA. *Helicobacter pylori* urease: properties and role in pathogenesis. *Scand. J. Gastroenterol.* (1991) 187: 39-46.
- Bauerfeind P, Garner R, Dunn BE and Mobley HL. Synthesis and activity of *Helicobacter pylori* urease and catalase at low pH. *Gut* (1997) 40: 25-30.
- Michetti K. Pathogenesis of *Helicobacter pylori* infection. *Curr. Opin. Gastroenterol.* (1998) 14: 57-63.
- Dixon NE, Gazzola C, Blakeley RL and Zerner B. Jack bean urease (EC 3.5.1.5), A metalloenzyme, A simple biological role for nickel? *J. Am. Chem. Soc.* (1975) 97: 4131-413.
- Karplus PA, Pearson MA and Hausinger RP. 70 Years of crystalline urease: what have we learned? *Acc. Chem. Res.* (1997) 30: 330-337.
- Sachs G, Scott D, and Weeks D. The compartment buffered by the urease of *Helicobacter pylori*: cytoplasm or periplasm. *Trends Microbiol.* (2002) 10: 217-218.
- Mobley HLT, Island MD and Hausinger RP. Molecular biology of microbial ureases. *Microbiol. Rev.* (1995) 59: 451-480.
- Zheng Q, Van Cleemput O, Demeyer P and Baert L. Nitrous oxide and methane emissions during rice growth and through rice plants: Effect of dicyandiamide and hydroquinone. *Biol. Fertil. Soils.* (1991) 11: 41-47.
- Pedrazzini F, Tarsitano R and Nannipieri P. The effect of phenyl phosphorodiamidate on urease activity and ammonia volatilization in flooded rice. *Biol. Fertil. Soils* (1987) 3: 183-188.
- Tanaka T, Kawase M and Tani S. α -Hydroxyketones as inhibitors of urease. *Bioorg. Med. Chem.* (2004) 12: 501-505.
- Ashiralieva A and Kleiner D. Polyhalogenated benzo- and naphthoquinones are potent inhibitors of plant and bacterial ureases. *FEBS Lett.* (2003) 367-370.
- Tanaka T, Kawase M and Tani S. Urease inhibitory activity of simple α,β -unsaturated ketones. *Life Sci.* (2003) 73: 2985-2990.
- Ahmad V, Hussain J, Hussain H, Jassbi AR, Ullah F, Lodhi MA, Yasin A and Iqbal M. First natural urease inhibitor from *Euphorbia decipiens*. *Chem. Pharm. Bull.* (2003) 51: 719-723.
- Limuro M and Wakabayashi K. Preventive effects of *Cladosiphon Fucoïdan* against *Helicobacter pylori* infection in mongolian gerbils. *Helicobacter* (2003) 8: 59-65.
- Goshima H and Kimura N. Acid dependent adherence of *Helicobacter pylori* urease to diverse polysaccharides. *Gastroenterology* (2000) 119: 358-367.
- Weatherburn MW. Phenol-hypochlorite reaction for determination of ammonia. *Anal. Chem.* (1967) 39: 971-974.
- Principe PP. The economic significance of plants and their constituents as drugs. In: Wagner H, Hiroshi Hikino Farnsworth, N.R. (eds). *Economic and*

- Medicinal Plant Research*. Vol. 3, Academic Press, London (1985) 17.
- (25) Kubo J, Lee R and Kubo I. Anti-*Helicobacter pylori* agents from the Cashew apple. *Agric. Food Chem.* (1999) 47: 533-537.
- (26) Tabak M, Artmon R and Neeman I. Cinnamon extracts' inhibitory effect on *Helicobacter pylori*. *J. Ethnopharmacol.* (1999) 67: 269-277.
- (27) Yee YK, Koo MWL and Szeto ML. Chinese tea consumption and lower risk of *Helicobacter* infection. *J. Gastroenterol. Hepatol.* (2002) 17: 552-555.
- (28) Matsubara S, Shibata H, Ishikawa F, Yokokura T, Takahashi M and Sugimura T. Suppression of *Helicobacter pylori* induced gastritis by green tea extract in Mongolian gerbils. *Biochem. Biophys. Res. Comm.* (2003) 310: 715-719.
- (29) George-Okafor UO and Anosike EE. The potentials of lime (*Citrus aurantifolia*) for improving traditional corn fermentation for probiotic lactic acid bacterial proliferation. *Pak. J. Nutr.* (2010) 9: 1207-1213.
- (30) Srinivasan D, Ramasamy S and Sengottuvelu S. Protective effect of polyherbal formulation on experimentally induced ulcer in rats. *Pharmacologyonline* (2008) 1: 331-350.
- (31) Ndukwe KC, Lamikanra A and Okeke IN. Antibacterial activity in plants used as chewing sticks in Africa. *Drugs Future* (2004) 29: 1221-1233.
- (32) Pino JA and Rosado A. Comparative investigation of the distilled lime oils (*Citrus aurantifolia* Swingle and *Citrus latifolia* Tanaka) from Cuba. *J. Essent. Oil Res.* (2001) 13: 179-180.
- (33) Adeniyi BA, Onwubuche BC, Anyiam FM, Ekundayo O and Mahady GB. Anti- *Helicobacter pylori* activities of *Eucalyptus grandis*: Effects on susceptibility, urease activity and cell surface hydrophobicity. *Pharm. Biol.* (2009) 47: 13-17.
- (34) Srivastava JK and Shankar E. Chamomile: an herbal medicine of the past with bright future. *Mol. Med. Report* (2010) 3: 895-901.
- (35) Denkova Z, Slavchev A, Albena S, Schmidt E and Geissler M. Antimicrobial activities of roman chamomile oil from France and its main compounds. *J. Essent. Oil Res.* (2009) 21: 283-286.
- (36) Weseler A, Geiss HK and Saller R. A novel colorimetric broth microdilution method to determine the minimum inhibitory concentration (MIC) of antibiotics and essential oils against *Helicobacter pylori*. *Pharmazie* (2005) 60: 498-502.
- (37) Owlia P, Saderi H, Rasooli I and Sefidkon F. Antimicrobial characteristics of some herbal Oils on *Pseudomonas aeruginosa* with special reference to their chemical compositions. *Iranian J. Pharm. Res.* (2009) 8: 107-114.
- (38) Özen T. Investigation of antioxidant properties of *Nasturtium officinale* (watercress) leaf extracts. *Acta Pol. Pharm.* (2009) 66: 187-193.
- (39) Tevefik Z. Investigation of antioxidant properties of *Nastratium officinale* (Watercress) leaf extracts. *Acta Poloniae* (2009) 66: 187-193.
- (40) Hecht SS. Chemoprevention of lung cancer by isothiocyanates. *Adv. Exp. Med. Biol.* (1996) 401: 1-11.
- (41) Kwon DY and Choi JG. *In-vitro* and *in-vivo* antibacterial activity of *Punica granatum* peel ethanol extract against *Salmonella*. *Evidence-based Compl. Alter. Med.* (2011) Art. No. 690518.
- (42) Voravuthikunchai SP and Mitchell H. Inhibitory and killing activities of medicinal plants against multiple antibiotic-resistant *Helicobacter pylori*. *J. Health Sci.* (2008) 54: 81-88.
- (43) Voravuthikunchai SP, Limsuwan S and Mitchell H. Effects of *Punica granatum* pericarps and *Quercus infectoria* nutgalls on cell surface hydrophobicity and cell survival of *Helicobacter. I. J. Health Sci.* (2006) 52: 154-159.

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