

Novel Oxadiazole Thioglycosides as Potential Anti-*Acinetobacter* Agents

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Abstract

The glycosylation of 1,3,4-oxadiazole-2-thiones has been performed with peracetylated β -pyranosyl bromide in the presence of potassium carbonate. Deprotection of acetylated thioglycosides was necessary for increasing their antibacterial effects. The structures of nucleosides were confirmed by ¹H NMR, ¹³C NMR and HRMS. The anomeric protons of nucleosides c₁₋₄ were assigned to the doublet, confirming the β -configuration. The synthesized compounds were tested for their antimicrobial activity against *Acinetobacter calcoaceticus* (Gram-negative) strain *in-vitro* in comparison with Ampicillin as a reference drug which is normally used for treating such infections. The synthetic compounds showed different inhibition zones against tested bacterial strain. Thioglycoside derivatives of 1,3,4-oxadiazole-2-thiones (c set) were more active against *Acinetobacter calcoaceticus* ATCC 23055 than "parent" 1,3,4-oxadiazole-2-thiones (a set), confirming the relation between glyco-conjugation and increasing of antiproliferative activity of antibiotic agents. The best result belonged to nucleoside bearing 2-furyl moiety in its heterocyclic nucleus (c₄). The existence of *m*-PhNO₂ group as Ar in structures of a set and their corresponding sugar derivatives decreased the antibacterial activity of them in comparison with the rest of synthetic compounds.

Keywords: 1,3,4-Oxadiazole-2-thione; β -Pyranosyl; Thioglycoside; Nucleoside; *in-vitro*; *Acinetobacter calcoaceticus*; Antiproliferative activity.

Introduction

The resistance of infective bacteria to present antibiotics demands research assigned to the discovery of new drugs in the antibacterial drug field. The majority of carbohydrates found in nature or biological systems exist as glycoconjugates in which the monosaccharide units are joined via O-, N-, or S-glycosidic bonds. Thioglycosides have received considerable attention, because they are widely employed as biological

inhibitors, inducers and ligands for affinity chromatography of carbohydrate-processing enzymes and proteins (1-8). They have excellent chemoselectivity in glycosylation processes as both donors and acceptors particularly via reaction processes that involve active and latent glycosylation protocols (9). The thioglycosyl heterocycles are sufficiently stable under a variety of reaction conditions and have the ability to be readily converted into a variety of other functionalities (10, 11). Multivalent display of carbohydrates is frequently used as a method to increase affinities in various contexts such as the binding of bacteria, bacterial toxins, galectins and other lectins (12-24). These properties may

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affect medicinal effect of antibiotic agents.

On the other hand, oxadiazole derivatives, which belong to an important group of heterocyclic compounds, have been the subject of extensive study in the recent past. Numerous reports have highlighted their chemistry and use. Diverse biological activities, such as anti-inflammatory, antitumor, antimicrobial and anticonvulsant, have been found to be associated with oxadiazole derivatives (25-28). Moreover, sulfur-containing heterocycles represent an important group of sulfur compounds that are promising for use in practical applications.

Therefore, it is interesting to report the synthesis of a new series of compounds in which the glycosyl moieties have been used as carriers for the heterocycles having the oxadiazole ring.

In our previous work (29), we reported the synthesis of antibacterial properties of new series of thioglycoside derivatives of 1,2,4-triazole-5-thiones, whereas in the present work, we report the synthesis of new groups of anti-*Acinetobacter calcoaceticus* agents in which 1,3,4-oxadiazole-2-thiones moiety is coupled to monosaccharide unit.

Experimental

^1H and ^{13}C NMR spectra were recorded on a Bruker AVANCE-300 spectrometer at 300 and 75 MHz, respectively in CDCl_3 using TMS as the internal standard. High-resolution mass spectra were obtained with a HPLC-Q-TOF system equipped with Q-TOF micromass spectrometer (dual ESI). Melting points were measured on a Philip Harris C4954718 apparatus without calibration. Optically active samples were analyzed by EHARTNACK apparatus (Paris, France) at 20 °C in dichloromethane. Thin layer chromatography (TLC) analyses were carried out on silica gel plates. All chemicals were purchased from Merck and used as received.

5-phenyl-1,3,4-oxadiazole-2(3H)-thione (a₁)

Yield: 74%; mp 158-160 °C; ^1H NMR spectrum, (300 MHz, CDCl_3), δ , ppm, (J, Hz): 7.52 (m, 3H, ArH), 7.96 (d, J = 6.9, 2H, ArH), 10.75 (bs, 1H, N-H); ^{13}C NMR spectrum, (75 MHz, CDCl_3), δ , ppm: 122.90, 126.47, 129.85, 132.67, 160.90 (Ar), 177.88 (C=S).

5-(2-hydroxyphenyl)-1,3,4-oxadiazole-2(3H)-thione (a₂)

Yield: 84%; mp 162-164 °C; ^1H NMR spectrum, (300 MHz, CDCl_3), δ , ppm, (J, Hz): 7.10 (m, 2H, ArH), 7.49 (t, 1H, J = 7.1, ArH), 7.72 (d, 1H, J = 8.1, ArH), 8.39 (bs, 1H, OH), 11.34 (bs, 1H, N-H); ^{13}C NMR spectrum, (75 MHz, CDCl_3), δ , ppm: 109.79, 111.17, 119.84, 129.50, 133.87, 156.78, 160.27 (Ar), 177.47 (C=S).

5-(3-nitrophenyl)-1,3,4-oxadiazole-2(3H)-thione (a₃)

Yield: 78%; mp 167-168 °C; ^1H NMR spectrum, (300 MHz, CDCl_3), δ , ppm, (J, Hz): 7.40-7.60 (m, 3H, ArH), 7.94 (d, J = 7.8, 1H, ArH), 11.10 (bs, 1H, OH); ^{13}C NMR spectrum, (75 MHz, CDCl_3), δ , ppm: 120.93, 124.43, 126.85, 131.76, 132.44, 148.53, 159.15 (Ar), 178.04 (C=S).

5-(2-furyl)-1,3,4-oxadiazole-2(3H)-thione (a₄)

Yield: 69%; mp 152-155 °C; ^1H NMR spectrum, (300 MHz, CDCl_3), δ , ppm, (J, Hz): 6.62 (bs, 1H, Furyl), 7.18 (bs, 1H, Furyl), 7.66 (bs, 1H, Furyl), 11.38 (bs, 1H, N-H); ^{13}C NMR spectrum, (75 MHz, CDCl_3), δ , ppm: 113.05, 115.37, 137.99, 147.61, 153.90 (Ar), 177.10 (C=S).

2-phenyl-5-(2,3,4,6-tetra-O-acetyl- β -D-1-thio-glucopyranose)-1,3,4-oxadiazole (b1)

Yield: 64%; mp 102-105 °C; ^1H NMR spectrum, (300 MHz, CDCl_3), δ , ppm, (J, Hz): 1.95 (s, 3H, OAc), 2.04 (s, 3H, OAc), 2.09 (s, 3H, OAc), 2.18 (s, 3H, OAc), 3.88-3.99 (m, 1H, H-6a), 4.12-4.20 (m, 1H, H-6b), 4.27-4.30 (m, 1H, H-5), 5.14-5.71 (m, 3H, H-2, -3, -4), 5.96 (d, 1H, $J_{1,2} = 9.3$, H-1), 7.52 (m, 3H, ArH), 7.94-8.04 (m, 2H, ArH); ^{13}C NMR spectrum, (75 MHz, CDCl_3), δ , ppm: 20.56 (2C), 20.72 (2C) ($4 \times \text{OCOCH}_3$), 61.55 (C-6), 67.56 (C-4), 69.76 (C-2), 73.52 (C-3), 74.71 (C-5), 83.42 (C-1), 121.93, 126.80, 129.18, 132.03, 132.79 (Ar), 166.52 (C-S), 168.93, 168.37, 170.06, 170.58 ($4 \times \text{OCOCH}_3$).

2-(2-hydroxyphenyl)-5-(2,3,4,6-tetra-O-acetyl- β -D-1-thio-glucopyranose)-1,3,4-

oxadiazole (b2)

Yield: 66%; mp 99-101 °C; ¹H NMR spectrum, (300 MHz, CDCl₃), δ, ppm, (J, Hz): 2.03 (s, 3H, OAc), 2.04 (s, 3H, OAc), 2.06 (s, 3H, OAc), 2.10 (s, 3H, OAc), 3.88-3.92 (m, 1H, H-6a), 4.13-4.17 (m, 1H, H-6b), 4.27-4.32 (m, 1H, H-5), 5.14-5.37 (m, 3H, H-2, -3, -4), 5.50 (d, 1H, J_{1,2} = 9.6, H-1), 7.02 (t, 1H, J = 7.8, ArH), 7.13 (d, 1H, J = 8.4, ArH), 7.47 (t, 1H, J = 6.9, ArH), 7.72 (d, 1H, J = 7.8, ArH), 9.87 (bs, 1H, OH); ¹³C NMR spectrum, (75 MHz, CDCl₃), δ, ppm: 20.54 (2C), 20.60 (2C) (4 × OCOCH₃), 61.56 (C-6), 67.72 (C-4), 69.74 (C-2), 73.46 (C-3), 76.62 (C-5), 83.33 (C-1), 107.58, 117.66, 120.11, 126.54, 134.04, 157.34, 160.02 (Ar), 165.92 (C-S), 169.34, 169.43, 169.966, 170.54 (4 × OCOCH₃).

2-(3-nitrophenyl)-5-(2,3,4,6-tetra-O-acetyl-β-D-1-thio-glucopyranose)-1,3,4-oxadiazole (b3)

Yield: 58%; mp 112-113 °C; ¹H NMR spectrum, (300 MHz, CDCl₃), δ, ppm, (J, Hz): 2.04 (s, 3H, OAc), 2.06 (s, 3H, OAc), 2.09 (s, 3H, OAc), 2.10 (s, 3H, OAc), 3.88-4.05 (m, 1H, H-6a), 4.14-4.21 (m, 1H, H-6b), 4.28-4.33 (m, 1H, H-5), 5.16-5.67 (m, 3H, H-2, -3, -4), 5.97 (d, 1H, J_{1,2} = 9.3, H-1), 7.75 (t, J = 7.8, 1H, ArH), 8.27-8.43 (m, 2H, ArH), 8.84 (d, J = 8.1, 1H, ArH); ¹³C NMR spectrum, (75 MHz, CDCl₃), δ, ppm: 20.53 (2C), 20.65 (2C) (4 × OCOCH₃), 61.50 (C-6), 67.70 (C-4), 69.71 (C-2), 73.49 (C-3), 74.83 (C-5), 83.32 (C-1), 121.80, 124.93, 126.36, 127.03, 130.52, 132.21, 148.67 (Ar), 164.50 (C-S), 169.33, 169.44, 169.97, 170.53 (4 × OCOCH₃).

2-(2-furyl)-5-(2,3,4,6-tetra-O-acetyl-β-D-1-thio-glucopyranose)-1,3,4-oxadiazole (b4)

Yield: 72%; mp 107-109 °C; ¹H NMR spectrum, (300 MHz, CDCl₃), δ, ppm, (J, Hz): 1.97 (s, 3H, OAc), 2.04 (s, 3H, OAc), 2.07 (s, 3H, OAc), 2.10 (s, 3H, OAc), 3.85-4.00 (m, 1H, H-6a), 4.13-4.21 (m, 1H, H-6b), 4.27-4.32 (m, 1H, H-5), 5.24 (t, 1H, J_{1,2} = J_{2,3} = 9.3, H-2), 5.42 (t, 1H, J_{2,3} = J_{3,4} = 9.3, H-4), 5.61 (t, 1H, J_{2,3} = J_{3,4} = 9.3, H-3), 6.34 (d, 1H, J_{1,2} = 9.3, H-1), 6.61 (bs, 1H, Furyl), 7.17 (bs, 1H, Furyl), 7.66 (bs, 1H, Furyl); ¹³C NMR spectrum, (75 MHz, CDCl₃), δ, ppm: 20.49 (2C), 20.70 (2C) (4 × OCOCH₃),

61.54 (C-6), 67.51 (C-4), 69.40 (C-2), 73.04 (C-3), 74.75 (C-5), 83.17 (C-1), 112.36, 116.17, 137.41, 146.80, 152.37 (Ar), 159.98 (C-S), 168.97, 169.28, 170.04, 170.56 (4 × OCOCH₃).

2-phenyl-5-(β-D-1-thio-glucopyranose)-1,3,4-oxadiazole (c1)

Yield: 33%; mp 108-110 °C; [α]_D²⁰ = 3° (c = 1.0, CH₂Cl₂). ¹H NMR spectrum, (300 MHz, CDCl₃), δ, ppm, (J, Hz): 3.92-4.03 (m, 1H, H-6a), 4.12-4.22 (m, 1H, H-6b), 4.27-4.30 (m, 1H, H-5), 4.35-4.85 (m, 4H, OH), 5.12-5.70 (m, 3H, H-2, -3, -4), 5.99 (d, 1H, J_{1,2} = 9.3, H-1), 7.50 (m, 3H, ArH), 7.94-8.02 (m, 2H, ArH); ¹³C NMR spectrum, (75 MHz, CDCl₃), δ, ppm: 61.76 (C-6), 67.64 (C-4), 69.26 (C-2), 73.59 (C-3), 74.78 (C-5), 83.37 (C-1), 122.23, 126.88, 129.40, 132.09, 132.99 (Ar), 166.45 (C-S); HRMS spectrum (ESI), *m/z*: Calculated, 340.0729. C₁₄H₁₆N₂O₆S [M+H]⁺. Found, 341.0817.

2-(2-hydroxyphenyl)-5-(β-D-1-thio-glucopyranose)-1,3,4-oxadiazole (c2)

Yield: 46%; mp 144-145 °C; [α]_D²⁰ = 4° (c = 1.0, CH₂Cl₂). ¹H NMR spectrum, (300 MHz, CDCl₃), δ, ppm, (J, Hz): 3.86-3.97 (m, 1H, H-6a), 4.11-4.16 (m, 1H, H-6b), 4.26-4.32 (m, 1H, H-5), 4.37-4.83 (m, 4H, OH), 5.13-5.37 (m, 3H, H-2, -3, -4), 5.57 (d, 1H, J_{1,2} = 9.6, H-1), 7.03 (t, 1H, J = 7.8, ArH), 7.12 (d, 1H, J = 8.4, ArH), 7.49 (t, 1H, J = 6.9, ArH), 7.74 (d, 1H, J = 7.8, ArH), 9.97 (bs, 1H, OH); ¹³C NMR spectrum, (75 MHz, CDCl₃), δ, ppm: 62.46 (C-6), 68.02 (C-4), 69.86 (C-2), 73.65 (C-3), 76.69 (C-5), 83.04 (C-1), 109.18, 117.36, 121.01, 126.94, 133.84, 157.86, 160.23 (Ar), 165.57 (C-S); HRMS spectrum (ESI), *m/z*: Calculated, 356.0678. C₁₄H₁₆N₂O₇S [M+H]⁺. Found, 357.0762.

2-(3-nitrophenyl)-5-(β-D-1-thio-glucopyranose)-1,3,4-oxadiazole (c3)

Yield: 39%; mp 122-124 °C; [α]_D²⁰ = -2° (c = 1.0, CH₂Cl₂). ¹H NMR spectrum, (300 MHz, CDCl₃), δ, ppm, (J, Hz): 3.84-4.01 (m, 1H, H-6a), 4.17-4.25 (m, 1H, H-6b), 4.24-4.33 (m, 1H, H-5), 4.36-4.85 (m, 4H, OH), 5.16-5.68 (m, 3H, H-2, -3, -4), 6.07 (d, 1H, J_{1,2} = 9.3, H-1), 7.78 (t, J = 7.8, 1H, ArH), 8.27-8.46 (m, 2H, ArH), 8.81 (d, J = 8.4, 1H, ArH); ¹³C NMR spectrum, (75 MHz, CDCl₃), δ, ppm: 62.20 (C-6), 67.74 (C-

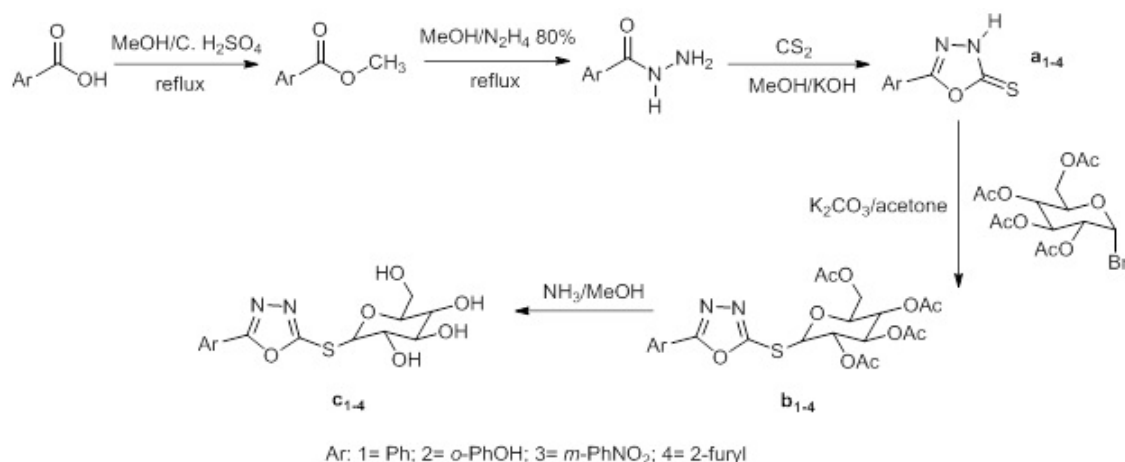


Figure 1. General synthetic pathway for the synthesis of thioglycosyl oxadiazoles.

4), 69.79 (C-2), 73.67 (C-3), 74.35 (C-5), 83.44 (C-1), 121.89, 123.90, 126.86, 127.43, 131.82, 132.28, 149.63 (Ar), 165.22 (C-S); HRMS spectrum (ESI), m/z : Calculated, 385.0580. C₁₄H₁₅N₃O₈S [M+H]⁺. Found, 386.0665.

2-(2-furyl)-5-(β-D-1-thio-glucopyranose)-1,3,4-oxadiazole (c4)

Yield: 52%; mp 111-113 °C; [α]_D²⁰ = -4° (*c* = 1.0, CH₂Cl₂). ¹H NMR spectrum, (300 MHz, CDCl₃), δ, ppm, (J, Hz): 3.82-4.08 (m, 1H, H-6a), 4.17-4.23 (m, 1H, H-6b), 4.24-4.39 (m, 1H, H-5), 4.35-4.89 (m, 4H, OH), 5.27 (t, 1H, J_{1,2}=J_{2,3} = 9.3, H-2), 5.45 (t, 1H, J_{2,3}=J_{3,4} = 9.3, H-4), 5.69 (t, 1H, J_{2,3}=J_{3,4} = 9.3, H-3), 6.31 (d, 1H, J_{1,2} = 9.3, H-1), 6.66 (bs, 1H, Furyl), 7.17 (bs, 1H, Furyl), 7.58 (bs, 1H, Furyl); ¹³C NMR spectrum, (75 MHz, CDCl₃), δ, ppm: 60.94 (C-6), 66.98 (C-4), 69.87 (C-2), 73.54 (C-3), 74.85 (C-5), 83.63 (C-1), 112.30, 117.20, 137.83, 146.88, 152.52 (Ar), 162.07 (C-S); HRMS spectrum (ESI), m/z : Calculated, 330.0522. C₁₂H₁₄N₂O₇S [M+H]⁺. Found, 331.0609.

Bacterial Strain

The antibacterial activity of compounds was assayed with our previous published method (30). The antibacterial activity of the compounds was tested against Gram-negative strain of *Acinetobacter calcoaceticus* ATCC 23055.

Results and Discussion

Thioglycosilation was performed according our previous published method (29). 1-bromide sugar and 1,3,4-oxadiazole-2-thione nuclei a₁₋₄ were synthesized to the literature procedures (29,31). Deprotection of acetylated nucleosides was performed to the literature procedure (32). The synthesis of the final nucleosides is depicted in Figure 1.

The structure of thioglycosides was confirmed by appropriate spectroscopic methods such as ¹H NMR, ¹³C NMR, and high resolution mass spectroscopy (HRMS). The anomeric protons of nucleosides c₁₋₄ were assigned to the doublet at 5.57–6.31 ppm with J_{1,2}=9.3–9.6 Hz, confirming the β-configuration.

In our recent published research (29), there was not any significant change in the antibacterial effect of the acetylated final thioglycosides in comparison with the parent heterocyclic nuclei. Therefore, in the present work, the deacetylated nucleosides c₁₋₄ have been compared to the parent heterocyclic nuclei a₁₋₄ against *Acinetobacter calcoaceticus*.

The *in vitro* antibacterial activity of the synthesized compounds in DMSO against *Acinetobacter calcoaceticus* shown in Tables 1. Diameter of inhibition zone for ampicillin as reference drug is 18 mm for 10 μg/μL concentration of drug in the same test conditions.

Table 1. *In-vitro* antibacterial activity of synthetic compounds against *Acinetobacter Calcoaceticus* ATCC 23055 (concentration = 10 µg/µL).

Compound	Diameter of inhib. zone (mm)	Compound	Diameter of inhib. zone (mm)
a ₁	19	c ₁	28
a ₂	18	c ₂	29
a ₃	16	c ₃	25
a ₄	22	c ₄	31

As shown in the Table 1. these compounds showed higher antibacterial effects in comparison with Ampicillin (18 mm) which is normally used for treating such infections.

In general, compounds from **c** set (nucleosides) showed more antimicrobial activity than the other set. Thioglycoside derivatives of 1,3,4-oxadiazole-2-thiones (**c** set) were more active against *Acinetobacter calcoaceticus* ATCC 23055 than “parent” 1,3,4-oxadiazole-2-thiones (**a** set), confirming the relation between glyco-conjugation and increasing of antiproliferative activity of antibiotic agents. The best results in the tables belonged to **c₄** that showing high activity against *A. calcoaceticus* (31 mm).

Going over the structure of these synthetic compounds confirmed that the existence of 2-furyl instead of *m*-PhNO₂ group as Ar increased their antibacterial activity against *A. calcoaceticus*. The existence of *m*-PhNO₂ group as Ar in structures of **a** set and their corresponding sugar derivatives decreased the antibacterial activity of them in comparison with the rest of synthetic compounds.

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