# QTL mapping of heading date and plant height in Barley cross "Azumamugi"×"Kanto Nakate Gold"

Fahimeh Shahinnia¹, Abdolmajid Rezai¹, Badraldin Ebrahim Sayed-Tabatabaei²\*, Takao Komatsuda³, Sayed Abolghasem Mohammadi⁴

<sup>1</sup>Department of Agronomy and Plant Breeding, <sup>2</sup>Department of Agrobiotechnology, College of Agriculture, Isfahan University of Technology, P.O. Box 84154-83111, Isfahan, I.R. Iran <sup>3</sup> Plant Genome Research Unit, National Institute of Agrobiological Sciences, Kannondai 2-1-2, Tsukuba, P.O. Box 305-8602, Ibaraki, Japan <sup>4</sup>Department of Agronomy and Plant Breeding, College of Agriculture, Tabriz University, P.O. Box 51666-16471, Tabriz I.R. Iran

#### **Abstract**

To identify quantitative trait loci (QTLs) controlling heading date and plant height, ninety nine F<sub>13</sub> recombinant inbred lines (RILs) derived from barley cultivars Azumamugi x Kanto Nakate Gold cross were evaluated. The field trails were conducted at randomized complete block design with two and three replications in 2004 and 2005, respectively. Significant differences and transgrassive segregates were observed among lines for heading date and plant height. Composite interval mapping (CIM) was done based on linkage map constructed using 99 RILs and 100 markers including isozyme, morphological, STS and AFLP markers. A strong QTL controlling 26% phenotypic variation of heading date on chromosome 5HL was located near to the e07m25.3-e12m199.1 markers. The QTL had the same interval of the Sgh<sub>2</sub> locus. Allele inherited from Azumamugi parent in this locus decreased heading date. The QTL for heading date in the map interval of ABC261-ABG055 markers on chromosome 1HL could be identical with the eam8 locus and accounted for 11% of the phenotypic variation. New QTL for plant height was detected near to uzu1 locus on chromosome 3HL, explaining 52% of the phenotypic variation. The effect of allele transmitted from Azumamugi parent in this locus decreased plant height. The QTLs identified on chromosomes 1HL, 3HL and 7HS in relation to eam8, uzu1 and dsp1 genes showed pleiotropic effects on controlling heading date and plant height.

**Keywords:** Composite interval mapping; Hordeum vulgare L.; Major genes; Pleiotropic effects; QTLs combination.

# INTRODUCTION

The relationship between heading date and plant height is particularly important in the grass family

\*Correspondence to: **Badraldin E. Sayed-Tabatabaei**, Ph.D. Tel: +98 311 3913344; Fax: +98 311 3912254 E-mail: sayedt@cc.iut.ac.ir

including barley, in which apical growth is terminated by flowering (Lin *et al.*, 1995). Increased plant height is often correlated with late flowering. Heading date is an important factor in adapting cereal varieties to environment and in maximizing yield potential. The control of plant height can be used to reduce yield loss arising from lodging and to increase harvest index (Bezant *et al.*, 1996). These are determined by the interaction of three genetic factors, vernalization response (sensitivity to temperature), photoperiodic response (sensitivity to short days) and earliness in the narrow sense (sensitivity to long days) (Takahashi and Yasuda, 1970; Hackett *et al.*, 1992).

Heading date and plant height show continuous variation and they are controlled by quantitative trait loci (QTLs). The inheritance of these characters is complex and usually assumed to involve numerous genetic factors that interact with environmental conditions (Kjær et al., 1995). The advent and development of molecular markers in quantitative genetics greatly facilitates the study of complex quantitatively inherited traits by the construction of high density genome linkage maps for crops such as barley and has made it possible to dissect the poly genes for such traits into individual Mendelian factors (Xiao et al., 1996). Using molecular linkage maps and QTL mapping methods, it is possible to estimate the number of loci controlling genetic variation in a segregating population and to characterize these with regard to their map position in the genome, gene action, phenotypic effects, pleiotropic effects and epistatic interactions with other QTLs (Xiao et al., 1996; Zhu et al., 1999).

Several statistical methods are available to detect and locate QTL. The most popular analytical method is of flanking marker mapping, either by the log-likelihood approach of interval mapping or by multiple regressions (Lander and Botestein, 1989; Haley and Knott, 1992; Martinez and Curnow, 1992). Combining interval mapping with multiple regressions has been proposed to attempt to separate the effects of possible multiple linked QTL (Jenson, 1992; Zeng, 1994). Cofactors are used to reduce the effect of back ground noise or variation by taking into account the variation of these selected markers with the QTLs (Bezant *et al.*, 1996).

The objectives of this study were to (1) identify QTLs controlling heading date and plant height (2) investigate correspondence between major genes and QTLs affecting the traits in a recombinant inbred lines population derived from the cross between two Japanese cultivars of barley.

### MATERIALS AND METHODS

Plant materials and trait evaluations: We used ninety nine F<sub>13</sub> recombinant inbred lines of barley derived via single seed descent method from F<sub>2</sub> plants of Azumamugi (AZ) × Kanto Nakate Gold (KNG) cross (T. Komatsuda, unpublished data), in field condition at National Institute of Agrobiological Sciences, Tsukuba, Japan. AZ is six-rowed Japanese, winter barley and KNG is two-rowed Japanese, spring cultivar. The RILs used in this study were produced to identify the QTLs for shoot differentiation in barley (Komatsuda et al., 1989; Mano and Komatsuda, 2002). Also the population showed variation for important agronomic traits including heading date and plant height. AZ parental line showed lower value for plant height, whereas KNG line had higher value for heading date (Shahinnia et al., 2005). The field trails were conducted at randomized complete block design with two replications in 2004 and three replications in 2005 at Isfahan University of Technology Research Station, Isfahan, Iran (32° 32′ N, 51° 32′ E). The experimental plots consisted of four row plots, 2m × 0.8 m with 4 cm within row spacing. Two-hundred kg/ha ammonium phosphate as fertilizer before planting and 250 kg/ha urea at tillering stage were used.

Heading date was recorded when an estimated 50% of the spikes in each plot had completely emerged from the boot. Plant height was measured from the base of the culms to the tip of the spike (awn excluded). Data were recorded for 10 randomly plants in the center rows of each plot with considering 0.5 m border to omit bordering effect.

**Statistical analysis**: Analysis of variance was performed for data obtained from 2-year experiments using the general linear model (GLM) procedure of the SAS software (SAS Institute, 2000). Heritability ( $h^2$ ) was calculated from variance components according to Kearsey and Pooni (1996).

Molecular linkage map and QTL analysis: For the QTL analysis the genetic base map covering 925.6-cM containing 100 markers including 2 isozyme, 2 morphological, 34 STS and 62 AFLP markers (Mano *et al.*, 2001; Mano and Komatsuda, 2002) was used to obtain a density of 5 to 10-cM intervals without the clustering markers. Map distances were estimated based on Kosambi's map function (Kosambi, 1944) and the map was constructed using MAPMAKER 3.0 (Lander *et al.*, 1987; Lincoln *et al.*, 1992).

QTL analysis was performed using phenotypic data for the mean values of the traits evaluated in 2004 and 2005. Initially, single marker analysis (SM) was performed for each trait to identify markers associated with variation for traits using MAPMAKER/QTL ver.1.1b (Lander and Botestein, 1989). Further evaluation was carried out by composite interval mapping (CIM) with a 15-cM window and a maximum of 15 marker cofactors per model using Windows QTL Cartographer version 2.0 (Wang et al., 2004). Tests were performed at 1-cM intervals, and cofactors were selected by forward-backward stepwise regression (Model 6). Genome wide, trait specific, threshold values ( $\alpha$ = 0.05) of the likelihood ratio (LR) test statistic for declaring the presence of a QTL was estimated from 2000 permutation test by random sampling of phenotypic data (Churchill and Doerge, 1994; Doerge and Churchill, 1996). The phenotypic variation explained by a OTL  $(R^2)$  conditioned by the CIM cofactors included in the model was calculated at the most likely OTL position. The additive effect of an allelic substitution at each QTL was also obtained. The LOD peak of each significant QTL was considered as the QTL location on the linkage map.

## **RESULTS**

Phenotypic variations and heritability of traits: Pooled analysis of variance across 2 years (Table 1) confirmed the presence of highly significant differences among lines for heading date and plant height. The environmental effect was small, but the genotypic variance was large for both traits; thus, high heritability estimated for heading date and plant height (93%\*\* and 90%\*\* respectively). According to the frequency

# Archive of SID

distribution of phenotypes (Fig. 1), transgressive segregates were observed for heading date and plant height. AZ parental line had the lower value for heading date and plant height compared to KNG line. Also, the climatic condition of 2004 growing season was warmer and dryer than 2005. This may explain the earlier heading date and shorter plant height in 2004.

**Table 1.** Pooled analysis of variance across 2 years for heading date and plant height evaluated in the RILs derived from Azumamugi × Kanto Nakate Gold.

| Source       | df - | Mean square  |              |  |  |
|--------------|------|--------------|--------------|--|--|
|              |      | Heading date | Plant height |  |  |
| Year         | 1    | 9493.63**    | 70527.85**   |  |  |
| Block (Year) | 3    | 115.58       | 4519.78      |  |  |
| Line         | 98   | 250.18**     | 577.09**     |  |  |
| Line × Year  | 98   | 28.98        | 80.45        |  |  |

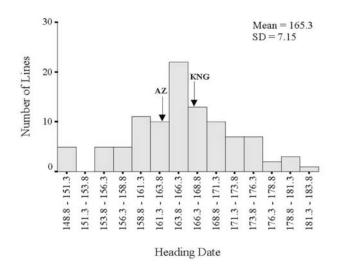
\*\* Significant at P<0.01

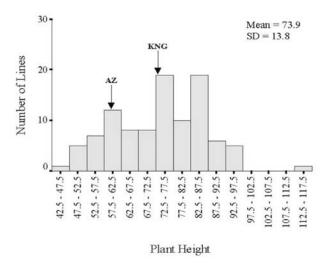
**Mapping QTLs:** For each trait, the linkage of QTLs to molecular markers mapped in the RILs population was assessed by single marker analysis. Although single marker analysis can be useful for a quick look at the data, especially in case of having missing observations that could affect later analysis, however QTL mapping methods such as CIM are more robust and powerful than SM (Basten *et al.*, 1999; Wang *et al.*, 2004). Therfore, only the results from CIM (Table 2) are presented here.

**Mapping QTLs for heading date:** The CIM analysis in  $F_{13}$  generation detected seven QTLs on chromosomes 1H, 2H, 3H, 5H and 7H (Fig. 2A). A strong QTL on chromosome 5HL near to the e07m25.3 mark-

er explained the highest phenotypic variance (26%) with the largest LOD score (Table 2). The effect of AZ allele in this locus reduced heading date. The QTL was located on chromosome 1HL near to the ABC261 marker had the highest additive effect on increasing heading date and explained 11 percent of total phenotypic variance with the considerable LOD score. The second QTL on chromosome 1HL with explaining about 10% phenotypic variance of heading date was located in interval of e02m18.4-e06m18.10 markers. The QTLs were detected on chromosome 2H in the interval of e11m19.3-ABG613 and e13m31.7.1e15m19.8.1 markers could totally explain 15% of heading date phenotypic variance. Both QTLs with allele from AZ parental line reduced heading date. The minor OTLs controlling heading date in interval of e15m19.7-ABA001 markers on chromosome 3HS and cMWG704-e11m1710.2 markers on chromosome 7HS showed the additive effect of AZ allele on increasing heading date.

Mapping QTLs for plant height: Results of CIM analysis indicated four QTLs controlling plant height (Fig. 2B). Of these, a major QTL located in the interval of *uzu-e06m30.10.1* markers on chromosome 3HL explained 52 percent of plant height phenotypic variance (Table 2). For this QTL allele inherited from AZ parent decreased plant height. Also three minor QTLs were identified near to the *ABC261*, *cMWG704* and *e12m2210.2* markers on chromosomes 1HL, 7HS and 7HL, respectively, could totally explained 19% of phenotypic variance. Allele of AZ parental line in the interval near to *ABC261* and *e12m2210.2* markers had positive effect on increasing plant height.





**Figure 1.** Frequency distribution of phenotypes for heading date and plant height in the RILs of Azumamugi × Kanto Nakate Gold cross based on means over years. Arrows indicate phenotypic values of AZ and KNG.

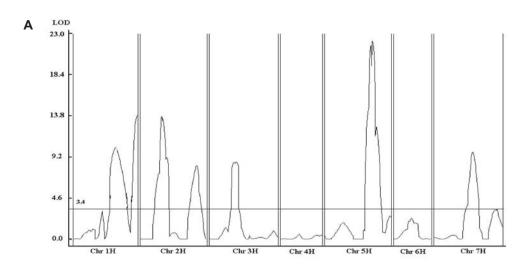
90

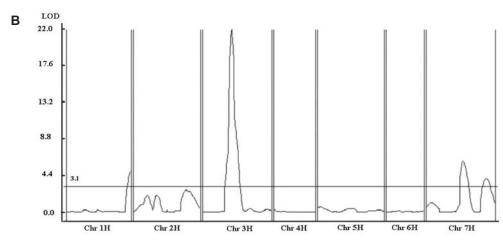
**Table 2**. The Chromosomal location, map interval, position of peak LOD, LOD score, additive effect and percent of explained variance by QTLs detected for heading date and plant height in the RILs derived from Azumamugi × Kanto Nakate Gold.

| Trait        | Chromosome | Interval              | Position <sup>a</sup><br>(cM) | LOD <sup>b</sup><br>Score | Additive <sup>c</sup><br>Effect | R <sup>2 d</sup> |
|--------------|------------|-----------------------|-------------------------------|---------------------------|---------------------------------|------------------|
| Heading date | 1HL        | e02m18.4-e06m18.10    | 99.21                         | 10.27                     | 2.45                            | 0.09             |
|              | 1HL        | ABC261-ABG055         | 143.81                        | 13.81                     | 2.56                            | 0.11             |
|              | 2HS        | e13m31.7.1-e15m19.8.1 | 60.61                         | 9.14                      | -2.21                           | 0.09             |
|              | 2HL        | e11m19.3-ABG613       | 125.31                        | 8.24                      | -2.06                           | 0.05             |
|              | 3HS        | e15m19.7-ABA001       | 49.91                         | 8.38                      | 1.69                            | 0.05             |
|              | 5HL        | e07m25.3-e12m199.1    | 107.21                        | 22.11                     | -3.93                           | 0.26             |
|              | 7HS        | cMWG704-e11m1710.2    | 87.81                         | 9.74                      | 2.22                            | 0.07             |
| Plant height | IHL        | ABC261-ABG055         | 144.81                        | 3.92                      | 2.52                            | 0.03             |
|              | 3HL        | uzu-e06m30.10.1       | 65.61                         | 30.08                     | -10.07                          | 0.52             |
|              | 7HS        | cMWG704-e11m1710.2    | 86.81                         | 7.87                      | -3.75                           | 0.08             |
|              | 7HL        | e12m2210.2-ABG608     | 135.61                        | 6.31                      | 3.95                            | 0.08             |

<sup>&</sup>lt;sup>a</sup> Distance of peak LOD score position from the left side marker.

<sup>&</sup>lt;sup>d</sup> Proportion of the phenotypic variance explained.





**Figure 2**. The Logarithm of odds (LOD) score plots of QTLs detected for (A) heading date and (B) plant height calculated by composite interval mapping in the RILs derived from Azumamugi × Kanto Nakate Gold based on means over years. LOD score profiles of chromosomes are shown in the order of 1H, 2H, 3H, 4H, 5H, 6H and 7H, oriented from short arm.

<sup>&</sup>lt;sup>b</sup> Peak LOD score of significant marker interval.

<sup>&</sup>lt;sup>c</sup>Additive effect of Azumamugi allele.

Correspondence between QTLs affecting heading date and plant height: We found a highly significant phenotypic correlation (r= 0.75) between heading date and plant height in this study. Some QTLs affecting heading date were associated with plant height QTLs (Table 2). Specifically, the two QTLs, close to the *ABC261* and *cMWG704* markers on chromosomes 1HL and 7HS, respectively, were overlapped. Also, by decreasing 1.0 LOD score from each side of the maximum value (Lander and Botestein, 1989), the QTLs controlling heading date and plant height in the interval of *e15m19.7-ABA001* and *uzu-e06m30.10.1* markers on chromosome 3H were mapped within overlapping 90% confidence intervals of each QTL LOD.

#### DISCUSSION

**Heading date:** In barley chromosome 1HL, the presence of the photoperiodic sensitive gene Ppd-H<sub>2</sub> was reported at the position being 20.6 cM proximal to the cMWG733 marker (Laurie et al., 1995). According to the barley consensus map in GrainGenes (http://www.graingenes.org), the cMWG733 marker located approximately at a position 10 cM proximal to that of the ABC261 marker and the OTL detected for heading date on chromosome 1HL near to the e02m18.4 marker in this study was located at a position 35.59 cM proximal to ABC261 marker. By decreasing 1.0 LOD score from each side of the maximum value (Lander and Botestein, 1989), the 90% confidence interval obtained for this QTL included the same position with the Ppd-H2 locus. Another QTL near to ABC261 marker on this chromosome was identical with the QTL detected by Marqueez-Cedillo et al., 2001. This QTL could be identical with the early maturity gene eam8 (Franckowiak, 1996) and detected by Takahashi and Yasuda (1970). Since the eam8 locus is present in Japanese two-rowed barley, it is possible that Kanto Nakate Gold (two-rowed) harbors an allele, which confers sensitivity to long days (Sameri and Komatsuda, 2004).

According to integrated molecular and morphological/physiological marker maps (Kleinhofs, 2002 and 2004) in barley, the QTL located on chromosome 2HS near to *e13m31.7.1* marker had the same interval including the *Eam1* (Early maturity 1) or *Ppd-H1* locus. This QTL has been reported before by Hayes *et al.*, 1993; Lauri *et al.*, 1995 and Baum *et al.*, 2003.

The QTL on chromosome 7HS close to cMWG704 marker might correspond to the dsp1 (dense spike 1) locus. For QTL in the interval of

e15m19.7-ABA001 near to uzu1 gene, no QTL have been identified in other mapping populations. It was known that AZ carries the uzu1 (semi brachytic) gene (Mano et al., 2001), so the intervals controlling heading date near to uzu1 locus can be investigated in this population and AZ parent causes delay in heading date.

Plant height: A strong QTL for plant height was detected on chromosome 3HL near to the uzu1 locus on which allele inherited from AZ parent causes plant height reduction. Since none of the parents carried the uzul gene in the former studies, no QTL in this locus have been reported before. The uzul gene has pleiotropic effects on several agronomic characters (Takahashi and Yamamoto, 1951). Even, a strong OTL for shoot differentiation has been found very close to the uzul locus, in tissue culture (Mano and Komatsuda, 2002). Kuraishi (1974) reported the pleiotropic effect of the uzul gene in reducing auxin hormone production. This may explain the effect of QTLs near to the *uzu1* gene on decreasing plant height. Marquez-Cedillo et al. (2001) and Teulat et al. (2001) reported two QTLs near the uzul locus, but the uzul gene coming from the Japanese cultivars (Takahashi and Yamamoto, 1951), so former QTLs may not be allelic with uzu1. The OTL detected for plant height at centromeric region on chromosome 7H, near to cMWG704 marker could be identical with QTL reported by Hayes et al. (1993) and Tinker et al. (1996). The location of cMWG704 marker was very close to the dsp1 locus on chromosome 7H, the locus with decreasing effect on stem internodes (Franckowiak and Konishi, 1997). The QTLs near to ABC261 marker on chromosome 1HL and e12m22.10 marker on chromosome 7HL might be new loci for controlling plant height detected in this population.

Sameri and Komatsuda (2004) and Sameri et al. (2006) using the recombinant inbred lines of the cross between AZ and KNG cultivars and a field experiment conducted in Japan, reported three strong QTLs for heading date located on chromosomes 1HL, 2HS and 5HL near to the *ABC261*, *e13m31.7.1* and *e07m25.3* markers, respectively and a major QTL near to uzu marker for plant height. The results of present study confirmed the stable position of these OTLs identified across two environments and are more confident for marker assisted selection in this population. Three QTLs for heading date on chromosomes 2HL, 3HS and 7HS near to e11m19.3, e15m19.7 and cMWG704 markers, respectively and a QTL for plant height on chromosomes 1HL near to ABC261 marker expressed in this study which were not detected in previous

report. Although the genomic position of a QTL is constant but the effect of a QTL may vary by environmental interaction and sampling variance (Tinker *et al.*, 1996; Utz *et al.*, 2000).

The results of present study revealed the correspondence between the QTLs affecting heading date and plant height in relation to *eam8*, *uzu1* and *dsp1* genes on chromosomes 1H, 3H and 7H, respectively. Marqueez-Cedillo *et al.* (2001) reported heading date QTLs on chromosome 3H and 5H, which had common flanking markers with plant height QTLs. Baum *et al.* (2003) also reported common flanking markers for plant height and heading date on chromosome 2H.

Major genes for earliness in barley have been shown to have pleiotropic effects on plant height (Kjær et al., 1991). However, it has been shown that major genes for height (i.e., the denso gene) have pleiotropic effects on heading date (Barua et al., 1993; Kjær et al., 1991). Comparative data support the possibility that independent closely linked genes, rather than a single gene, account for increased height and short day flowering, respectively (Lin et al., 1995).

It is important to determine to what extent QTL in one cross maybe equivalent to major genes in others, as this would allow close flanking markers for the major genes to be used to manipulate QTL in other crosses. This would increase the efficiency of marker assisted selection of QTL and the accuracy with which specific combination of QTL could be put together (Bezant *et al.*, 1996; Pillen *et al.*, 2003).

## Acknowledgments

Financial support provided by the Isfahan University of Technology, Isfahan, Iran is gratefully acknowledged.

#### References

- Barua UM, Chalmers KJ, Thomas WTB, Hackett CA, Lea V, Jack P, Forster BP, Waugh R, Powell W (1993). Molecular mapping of genes determining height, time to heading and growth habit in barley (*Hordeum vulgare*). *Genome* 36: 1080-1087.
- Basten CJ, Weir BS, Zeng ZB (1999). *QTL Cartographer*. Version 1.13, North Carolina State University, Raleigh, NC.
- Baum M, Grando S, Backes G, Jahoor A, Sabbagh A, Cecarelli S (2003). QTLs for agronomic traits in the Mediterranean environment identified in recombinant inbred lines of the cross "Arta" × H. spontaneum 41-1. Theor Appl Genet. 107: 1215-1225.
- Bezant J, Laurie D, Pratchett N, Chojeckls J, Kearsey M (1996). Marker regression mapping of QTL controlling flowering time and plant height in a spring barley (*Hordeum vulgare* L.) cross. *Heredity* 77: 64-73.
- Churchill GA, Doerge RW (1994). Empirical threshold values for quantitative trait mapping. *Genetics* 138: 963-971
- Doerge RW, Churchill GA (1996). Permutation tests for multiple

- loci affecting a quantitative character. *Genetics* 142: 285-294. Franckowiak JD (1996). Revised linkage maps for morphological markers in barley, *Hordeum vulgare*. *Barley Genet Newsl*. 26: 9-21
- Franckowiak JD, Konishi T (1997). BGS 9, dsp1. Barley Genet Newsl. 26:53.
- Hackett CA, Ellis RP, Forster BP, Mc-Nicol JW, Macualay M (1992). Statistical analysis of a linkage experiment in barley involving quantitative trait loci for height and ear-emergence time and two genetic markers on chromosome 4. *Theor Appl Genet*. 85: 120-126.
- Haley CS, Knott SA (1992). A simple regression method for mapping quantitative trait loci in line crosses using flanking markers. *Heredity* 69: 315-324.
- Hayes PM, Liu BH, Knapp SJ, Chen F, Jones B, Blake T, Franckowiak J, Rasmusson D, Sorrells M, Ullrich SE, Wesenberg D, Kleinhofs A (1993). Quantitative trait locus effects and environmental interaction in a sample of North American barley germplasm. *Theor Appl Genet.* 87: 392-401.
- Jenson RC (1992). A general mixture model for mapping quantitative trait loci by using molecular markers. *Theor Appl Genet*. 85: 252-260.
- Kearsey MJ, Pooni HS (1996). *The Genetical Analysis of Quantitative Traits*. Chapman and Hall, London, UK.
- Kjær B, Haahr V, Jensen J (1991). Association between 23 quantitative traits and 10 genetic markers in a barley cross. *Plant Breed*. 106: 261-274.
- Kjær B, Jensen J, Giess H (1995). Quantitative trait loci for heading date and straw characters in barley. Genome 38: 1098-1104.
- Kleinhofs A (2002). Integrating molecular and morphological /physiological marker maps. *Barley Genet Newsl.* 32: 152-159.
- Kleinhofs A (2004). Integrating molecular and morphological /physiological marker maps. *Barley Genet Newsl*. 34: 111-122.
- Komatsuda T, Enomoto S, Nakajima K (1989). Genetics of callus proliferation and shoot differentiation in barley. *J Hered*. 80: 345-350.
- Kosambi DD (1944). The estimation of map distances from recombination values. *Ann Eugen.* 12: 172-175.
- Kuraishi S (1974). Biogenesis of auxin in the coleoptiles of a semibrachytic barley, *uzu. Plant Cell Physiol.* 15: 295-306.
- Lander E, Botestein D (1989). Mapping Mendelian factors underlying quantitative traits using RFLP linkage maps. *Genetics* 121: 185-199.
- Lander ES, Green P, Abrahamson J, Barlow A, Daly MJ, Lincoln SE, Newburg L (1987). MAPMAKER: an interactive computer package for constructing primary genetic linkage maps of experimental and natural populations. *Genomics* 1: 174-181.
- Laurie DA, Pratchett N, Bezant JH, Snap JW (1995). RFLP mapping of five major genes and eight quantitative trait loci controlling flowering time in a winter × spring barley (*Hordeum vulgare* L.) cross. *Genome* 38: 575-585.
- Lin YR, Schertz KF, Paterson AH, 1995. Comparative analysis of QTLs affecting plant height and maturity across the Poaceae, in reference to an interspecific sorghum population. *Genetics* 141:391-411.
- Lincoln S, Daly M, Lander E (1992). *Constructing genetic maps with MAPMAKER/EXP 3.0.* 3<sup>rd</sup> ed, Whitehead Institute Technical Report.
- Mano Y, Kawasaki S, Takaiwa F, Komatsuda T (2001). Construction of a genetic map of barley (*Hordeum vulgare L.*)

- cross "Azumamugi" × "Kanto Nakate Gold" using a simple and efficient amplified fragment length polymorphism system. *Genome* 44: 284-292.
- Mano Y, Komatsuda T (2002). Identification of QTLs controlling tissue-culture traits in barley (*Hordeum vulgare L.*). Theor Appl Genet. 105: 708-715.
- Marquez-Cedillo LA, Hayes PM, Kleinhofs A, Legge WG, Rossnagel BG, Sato K, Ullrich SE, Wesenberg DM (2001). QTL analysis of agronomic traits in barley based on the doubled haploid progeny of two elite North American varieties representing different germplasm groups. *Theor Appl Genet*. 103: 625-637.
- Martinez O, Curnow RN (1992). Estimating the locations and the sizes of the effects of quantitative trait loci using flanking markers. *Theor Appl Genet.* 85: 480-488.
- Pillen K, Zacharias A, Leon J (2003). Advanced backcross QTL analysis in barley (*Hordeum vulgare L.*). Theor Appl Genet. 107: 340-352.
- Sameri M, Komatsuda T (2004). Identification of quantitative trait loci (QTLs) controlling heading time in the population generated from a cross between oriental and occidental barley cultivars (*Hordeum vulgare* L.). *Breeding Sci.* 54: 327-332.
- Sameri M, Takeda K, Komatsuda T (2006). Quantitative trait loci controlling agronomic traits in recombinant inbred lines from a cross of oriental- and occidental-type barley cultivars. *Breeding Sci.* 56: 243-252.
- SAS Institute (2000). *The SAS System for Windows*. Release 8.01, SAS Inst. Inc., Cary, NC.
- Shahinnia F, Rezai AM, Sayed-Tabatabaei BE (2005). Variation and path coefficient analysis of important agronomic traits in two- and six-rowed recombinant inbred lines of barley (Hordeum vulgare L.). Czech J Genet Plant Breed. 41: 246-250.
- Takahashi R, Yamamoto J (1951). Studies on the classification and geographical distribution of the Japanese barley varieties. III.

- On the linkage relation and the origin of the "uzu" or semibrachytic character in barley. Ber Ohara Inst Landw Forsch Kurashiki. 9: 399-410.
- Takahashi R, Yasuda S (1970). Genetics of earliness and growth habit in barley. *Barley Genetics II*. 388-408.
- Teulat B, Merah O, Souyris I, This D (2001). QTLs for agronomic traits from a Mediterranean barley progeny grown in several environments. *Theor Appl Genet*. 103: 774-787.
- Tinker NA, Mather DE, Rossnagle BG, Kasha KJ, Kleinhofs A, Hayes PM, Falk DE, Ferguson T, Shugar LP, Legge WG, Irvine RB, Choo TM, Briggs KG, Ullrich SE, Franckowiak J, Blake TK, Graf RJ, Doffing SM, Saghai-Maroof MA, Scoles GJ, Hoffman D, Dahleen LS, Kilian A, Chen F, Biyashev RM, Kudrna DA, Steffenson BJ (1996). Regions of the genome that affect agronomic performance in two-row barley. *Crop Sci.* 36: 1053-1062.
- Utz HF, Melchinger AE, Schon CC (2000). Bias and sampling error of the estimated proportion of genotypic variance explained by quantitative trait loci determined from experimental data in maize using cross validation and validation with independent samples. *Genetics*. 154: 1839-1849.
- Wang Sh, Basten CJ, Gaffney P, Zeng ZB (2004). Windowes QTL Cartographer 2.0 User Manual. Bioinformatics Research Center, NC. State University, USA.
- Xiao J, Li J, Yuan L, Tanksley SD (1996). Identification of QTLs affecting traits of agronomic importance in a recombinant inbred population derived from a sub specific rice cross. Theor Appl Genet. 92: 230-244.
- Zeng ZB (1994). Precision mapping of quantitative trait loci. *Genetics* 136: 1457-1468.
- Zhu H, Bricero G, Dovel R, Hayes PM, Liu BH, Ullrich SE (1999).
  Molecular breeding for grain yield in barley: an evaluation of QTL effects in a spring barley cross. *Theor Appl Genet*. 98: 772-779