Short Communication

Bactericidal effects of essential oils from clove, lavender and geranium on multi-drug resistant isolates of *Pseudomonas aeruginosa*

Mohaddeseh Mahboobi¹, Fereshteh Shahcheraghi², Mohammad Mehdi Feizabadi^{3*}

¹Laboratory for Microbiology, Barij Essence Pharmaceutical Industry, P.O. BOX 1178, Kashan, I.R. Iran ²Department of Microbiology, Pasteur Institute of Iran, P.O. BOX 13164, Tehran, I.R. Iran ³Department of Microbiology, School of Medicine, Tehran University of Medical Science, P.O. BOX 14155-5983 Tehran, I.R. Iran

Abstract

The inhibitory effects of essential oils including clove, lavender and geranium extracted from Eugenia caryophyllata, Lavandula officinalis and Pelargonium graveolens on multidrug resistant isolates of Pseudomonas aeruginosa were investigated. The main constituents of clove, lavander and geranium oil were eugenol (80-90%), 1,8-cineol (13%) and citronellol (45%) respectively. Clove had the most effective essential oil against P. aeruginosa. A combination consisting of clove, lavender and geranium oils at a ratio of 3:1:1 showed the most inhibitory effect (32-64 µg/ml) and strong synergy with gentamicin. The essential oils from clove, lavender and geranium exhibited bactericidal activity against multi-drug resistant strains of P. aeruginosa and may be alternatives compounds against these strains in the future. Keywords: Essential oils: Clove: Lavender: Geranium: Pseudomonas aeruginosa; Antimicrobial agents.

Pseudombiagaerfugibba is one of the major causes of serious infections in burn patients, resulting in mortality as high as 50% (Cohen, 1992; Lari *et al.*, 2000; Shahcheraghi *et al.*, 2003). This bacterium also causes opportunistic infections in different organs leading to diseases such as, bacteremia, and endocarditis. Resistance to different antibiotics is a major therapeutic problem in treatment of infections by this organism. In the last decade, resistance to the new generation of antibiotics within the population of *P. aeruginosa* has increased. The emergence of multi-drug resistant bac-

*Correspondence to: Mohammad Mehdi Feizabadi, Ph.D. Tel: +98 21 64432347; Fax: +98 21 88985810 E-mail: mfeizabadi@tums.ac.ir teria including *P. aeruginosa* strains has raised the needs for new antimicrobial drugs (Santos *et al.*, 1995).

Essential oils are the concentrated, hydrophobic liquids containing volatile aromatic compounds from plants. They possess a wide spectrum of pharmacological activities. The antimicrobial effects of essential oils have been documented and used in herbal medicine in many countries (Schilcher, 1998; Cowan, 1999; Schilcher, 2002; Longbottom *et al.*, 2004; Sonboli *et al.*, 2005).

Antibacterial effects of hydrous, methanolic and ethanolic extracts of clove, cinnamon, sage, thyme and rosmarinus on Gram-positive and Gram-negative bacteria had previously been investigated (Cowan, 1999). The results showed that all of these plants had antibacterial action on methicillin-resistant Staphylococcus aureus (MRSA) and Bacillus subtilis (Cowan, 1999), but they were weakly active against Gram-negative bacteria such as P. aeruginosa and enteropathogenic Escherichia coli (Shanab et al., 2004). With the exception of P. aeruginosa, the essential oil of Grammosciadium platycarpum with its major components, linalool (70%) and limonene (10%), displayed strong to intermediate antibacterial activity against Gram positive and Gram negative bacteria (Sonboli et al., 2005). Similarly, the minimum inhibitory concentration (MIC) for the essential oil of tea tree oil against *P. aeruginosa* was found to be much higher (%2 \leq MIC < 8%) than that against other bacteria.

Gram-negative bacteria are more resistant to antibiotics than the Gram-positive bacteria due to the permeability barrier provided by the cell wall or to the mem-

Strains	Inhibitory effects of essentials oils and their combinations										Synergy	
	С	G	L	CL 1:1	CLG 2:2:1	CLG 1:1:1	CLG 1:2:2	CLG 3:1:1	CLG 2:1:1	CLG 1:3:1	Gen	Gen+ CLG 3:1:1
ATCC 9027	18	13.6	16	14	14.3	15.3	15	18	16.3	14.3	24	24
513	13	12	12.5	12.5	14.5	12	13.5	13.5	13	15	-	20
514	12.5	10.5	12	13.5	15	13.5	12.5	16	14	15	-	16
515	14.5	10.5	11	13.5	13.5	14.5	13.5	14.5	12	13	22	22
516	11.6	11	11.3	15	11.6	13	13.6	14.3	13	12.3	20	22
589	13.3	10.6	12	13.3	12.3	13.6	12.3	13	12.3	13	-	14
502	12	12	12	13	11.5	15.5	11.5	13	13	12.5	-	16
594	13	11	11.5	13.7	14.7	12.7	11.5	14.2	14	11	-	13
584	12	9.3	11.6	13	13	12.3	11.6	14.6	13	13.5	-	15
581	13	10.5	13.5	13	12.5	13	11.5	14.5	11	13	-	16
586	14.3	12	11.6	14.3	14	14.3	13	13.5	14	14.6	-	14

Table 1. Anntibacterial activity and synergy of clove, geranium and lavender oils with gentamicin against different strains of *P. aeruginosa* determined by the disk diffusion method.

C: Clove oil, G: Geranium oil, L: Lavander oli, Gen: Gentamicin.

brane accumulation mechanism (Shanab *et al.*, 2004). Geranium, clove and lavender possess antimicrobial activities against a wide range of microorganisms (Santos *et al.*, 1995; Nascimento *et al.*, 2000; Shanab *et al.*, 2004). However, their bactericidal effects on *P. aeruginosa* strains have not yet been recognized.

To investigate the *in vitro* bactericidal effect of geranium, clove and lavender oils on *P. aeruginosa*, 10 isolates (isolates number 513-516, 502, 581, 584, 586 and 594) that were cultured from burn patients were used in this study. *P. aeruginosa* type strain ATCC 9027 was used as control in all experiments. The clinical isolates were multi resistant to gentamicin, ciprofloxacin and ceftazidime. The phenotypic characteristics of the clinical isolates have been reported elsewhere (Shahcheraghi *et al.*, 2003).

Archive essential oils were extracted from the appropriate plants by steam distillation (Guenther, 1982). Lavender oil (L) extracted from the flower of Lavandula (Lavandula officinalis) is yellowish or yellow-green in colour with a pleasant aroma and contains 1,8-cineol (13%). Geranium oil (G) was extracted from *Pelargonium graveolens*. It is a colorless or green-blue liquid with a floral aroma. The major fractions of this oil are citronellol (45%) and geranial (10-12%) (Guenther, 1982). Clove oil (C) was extracted from the dried flower buds of *Eugenia caryophyllata*. It has a warm, strong, spicy smell and the oil is colorless to pale yellow with a medium to watery viscosity.

Antibacterial activities of the essential oils against *P. aeruginosa* were determined by the disk diffusion and macrobroth dilution assays as recommended by National Committee for Clinical Laboratory Standards

(NCCLS, 2000; NCCLS, 2001). The isolates were inoculated onto Mueller-Hinton agar (MHA) and the disks impregnated with the essential oils were placed on the inoculated plates. C, L and G were tested separately and in combinations at different ratios (1:2, 2:1, 1:1, 2:2:1, 1:1:1, 1:2:2, 3:1:1, 2:1:1, 1:3:1). In brief, the surfaces of plates containing the MHA were swabbed with suspension of isolate adjusted to 0.5 McFarland (approximately 108 CFU/ml). The McFarland turbidity standard was prepared by adding 0.5 ml of 0.048 mol/l barium chloride (1.173 g BaCl₂. 2H₂O; 1.175% w/v) to 99.5 ml of 0.18 mol/l (0.36 N; 1% v/v) H₂SO₄ (NCCLS, 2001). The blank disks, 6 mm in diameter (Pad-Tan Teb, Tehran, Iran) were impregnated with 20 µl of each essential oil at a concentration of 100 mg/ml. Negative control was prepared using the same solvents (DMSO) employed to dissolve the essential oils. Dimethyl sulfoxide (DMSO) had no antimicrobial effect (Kontoyiannis et al., 2003, Shahidi Bonjar 2004, Baris et al., 2006). All the plates were incubated at 37°C for 24 h. Antibacterial activity was assessed by measuring the inhibitory zones around the disks (Table 1). The MICs were determined by the macrobroth dilution method using 2-fold dilutions of essential oils ranging from 4 to 512 µg/ml (NCCLS, 2001). The sizes of the inocula were adjusted to 105 CFU/ml and inoculated tubes were incubated at 37°C for 18 h (NCCLS, 2001). The MBCs of the oils were determined by plating 100 µl from each tube used for determining MIC and observed for any growth after 2 days of incubation.

The inhibitory zones of the 10 clinical strains were smaller than those of the reference strain. Shanab and Table 2. Determination of MIC and MBC of different combination of essential oils on the reference strain (*Pseudomonas aeruginosa* ATCC 9027) by the macrobroth dilution method.

Concentration (µg/ml)	Mixtures of different essential oils at different ratios									
	С	CG	CL	CLG	CLG	CLG	CLG	CLG	CLG	
		1:1	1:1	1:1:1	2:1:1	2:2:1	1:3:1	3:1:1	1:2:2	
MIC	93.75-	93.75-	93.75-	62.5-	62.5-	31.25-	31.25-	31.25-	62.5-	
	187.5	187.5	187.5	125	125	62.5	62.5	62.5	125	
MBC	187.5-	187.5-	187.5-	125-	125-	62.5-	62.5-	62.5-	125-	
	375	375	375	250	250	125	125	125	250	

C: Clove oil, G: Geranium oil, L: Lavander oli, Gen: Gentamicin, MIC: minimum inhibitory concentration, MBC: Minimum bactericidal concentration.

Table 3. Determination of the MIC and MBC of a mixture containing of Clove, Lavander and Geranium (3:1:1) on clinical isolates by the macrobroth dilution method.

Concentrations	Clinical Strains										
(µg/mi)	513	502	514	515	581	586	589	584	594	516	
MIC	32-64	32-64	32-64	32-64	32-64	32-64	16-32	32-64	16-32	32-64	
MDC	64-	64-	64-	64-	64-	64-	64-	64-	64-	64-	
MBC	128	128	128	128	128	128	128	128	128	128	

MIC: minimum inhibitory concentration, MBC: Minimum bactericidal concentration.

colleagues (2004) reported that the ethanolic extract of clove inhibited the growth of *P. aeruginosa* (MIC 781 μ g/ml). This was also confirmed by the results of this study. However, in our study the clove oil inhibited the growth of *P. aeruginosa* at concentrations between 93.75 to 187.5 μ g/ml. This study also demonstrated that clove oil possessed bactericidal activity at concentrations between 187.5 to 375 μ g/ml. Therefore, the clove essential oil extracted by steam distillation showed more activity than ethanolic extraction and may have therapeutic value.

Combination of clove, lavender and geranium at a ratio of 3:1:1 showed the best inhibitory effect on both the reference strain and clinical isolates of *P. aeruginosa* (MIC=32-64 μ g/ml). However, differences between the MICs and minimal bactericidal concentrations (MBCs) of different oils were found when they were tested on the reference strain separately (Table 2).

To determine the synergistic effect with gentamicin, the essentials oils were added to the disks containing this antibiotic (gentamicin $10 \mu g/disk$, Himedia Laboratories Pvt Limited, Mumbai, India) (Muroi and Kubo, 1996). The combination of clove, lavender and geranium oils at a ratio of 3:1:1 showed the most inhibitory effect (32-64 $\mu g/ml$) and strong synergy with gentamicin (Table 1).

The MBCs of the essential oils (3:1:1) for the reference strain did not differ from those of the clinical isolates and varied from 64-128 μ g/ml (Tables 2 and 3). Lavender oil is more effective against Gram-negative than Gram-positive bacteria. So, the use of lavender and clove oil as herbal drugs may be an alternative choice for the treatment of infections caused by Gramnegative bacteria such as the multi-drug resistant *P. aeruginosa.* Experimental studies with animals are needed to confirm our findings *in vivo*.

Acknowledgements

We thank N. Kazempour, M. Hosseini and Z. Haghighizadeh for their technical assistance, and are very grateful to Mr. Hejazi for his financial support of the project.

References

- Baris O, Güllüce M, Shahin F, özer H, Kilic H, özkan H, Sökmen M, özbek T (2006). Biological activities of the essential oil and methanol extract of *Achillea biebersteinii afan*. *Turk J Biol*. 30:65-73.
- Cohen L (1992). Epidemiology of drug resistance: implication for a post of antimicrobial era. *Science* 257: 1050-55.
- Cowan MM (1999). Plant products as antimicrobial agents. Clin Microbiol Rev. 12:564-582.
- Shahcheraghi F, Feizabadi MM, Yamin V, Abiri R, Abedian Z (2003). Serovar determination, drug resistance patterns and plasmid profiles of *Pseudomonas aeruginosa* isolated from burn patients at two hospitals of Tehran (IRAN). *Burns*. 29: 547-51.
- Guenther E (1982). The Essential oils. Robert E Krieger Publishing company Malabar Florida, Edithion 5, Volume IV, p. 671.
- Kontoyiannis DP, Lewis ER, Osherov N, Albert NA May GS WWW.SID.IF

(2003), Combination of caspofungin with inhibitors of calcineurin pathway attenuates growth *in vitro* in Aspergillus species. *J Antimicrob Chemoth.* 51:313-316.

- Lari AR, Alaghehbandan R, Nikui R (2000). Epidemiological study of 3341 burns patients during three years in Tehran, Iran. *Burns* 26: 49-53.
- Longbottom CJ, Carson CF, Hammer KA, Mee BJ, Riley TV (2004). Tolerance of Pseudomonas aeruginosa to Melaleuca alternifolia (tea tree) oil is associated with the outer membrane and energy-dependent cellular processes. *J Antimicrob Chemother*. 54: 386-92.
- Muroi H, Kubo I (1996). Antibacterial activity of anacardic acid and totarol, alone and in combination with methicillin, against methicillin-resistant *Staphylococcus aureus*. J Appl Bacteriol. 80: 387-94.
- Nascimento GFL, Locatell J, Freitas CP, Silva LG (2000). Antibacterial activity of plant extract and phytochemical on antibiotic resistant bacteria, *Braz J Microbiol.* 31: 347-251.
- National Committee for Clinical Laboratory Standards (2001). Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria That Grow Aerobically; *Approved Standard*, 5th ed: M7-A5 NCCLS, Wayne, PA.

National Committee for Clinical Laboratory Standards (2000).

Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically. *Approved standard*, 5th ed. NCCLS document M7-A5. NCCLS, Wayne, Pa.

- Santos PRV, Oliveira ACX, Tomassini TCB (1995). Controle Microbiogico de productous fitoterpicos. *Rev Farm Bioquim*. 31:35-38.
- Schilcher H (2002). Sind pflanzliche arzneimittel ttel bzwist die 'Naturmedizine' eine gefahr fur den anwender. *Arztezeitzchrift fur Naturheilver Fahren.* 43: 253-254.
- Schilcher H (1998). Pharmacologie and toxicologie atherischer. Ole Therapiewoche. 36: 1100-1112.
- Shanab AB, Ghaleb A, D'ahood AS, Naser J, Kanel A (2004). Antibacterial activity of some plant extract and utilized in popular medicine in Palestine. *Turk J Biol.* 28: 99-102.
- Shahidi Bonjar GH (2004). Evaluation of antibacterial properties of Iranian medicinal-plants against *Micrococcus luteus*, *Serratia marscescens*, *Klebsiella pneumonia* and *Bordetella bronchoseptica*. *Asian J Plant Sci.* 3:82-6.
- Sonboli A, Eftekhar F, Yousefzadi M, Kanani MR (2005). Antibacterial activity and chemical composition of the essential oil of Grammosciadium platycarpum Boiss. from Iran. Z Naturforsch [C] 60: 30-4.