

The Inhibitory Effect of Some Iranian Plants Extracts on the Alpha Glucosidase

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Abstract

Objective

Diabetes mellitus is manifested by hyperglycaemia. Different treatments such as diets and drugs are recommended for diabetes control. For various reasons in recent years traditional plant (herbal) therapies as prescribed by indigenous systems of medicine with different mechanisms have commonly been used. The digestive enzymes such as alpha glucosidase are among these herbal remedies.

Materials and Methods

One hundred species of plants were collected or purchased from the Medicinal Herbal Markets and botanically identified. Methanolic and aqueous extracts were prepared by the maceration method. The enzymatic activities of alpha glucosidase were determined colorimetrically by monitoring the release of p-nitrophenol from the appropriate p-nitrophenol glycoside substrate, after 30 mins incubation at 37 °C in the phosphate buffer (pH= 6.8).

Results

Among 200 prepared extracts, *Verbascum kermanensis*, *Rosa damascene*, *Rosmarinus officinalis*, *Levisticum officinale*, *Zataria multiflora*, *Sanguisorba minor*, *Alhagi camelorum*, *Pistacia vera*, *Vaccinium arctostaphylyus*, *Zhumeria majdae*, *Alpinia officinarum*, *Salvadora persica*, and *Thymus serpyllum* showed more than 50% inhibitory effect on the alpha glucosidase.

Conclusion

These active plants have no records in the literature for their anti diabetic effect and might be the new agents for diabetes control. This needs further *in vitro* and *in vivo* studies, some of which are under investigation.

Keywords: Alpha glucosidase, Diabetes, Herbals, Hyperglycemia, Inhibitors, Plant

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Introduction

Glycoside trimming enzymes are crucially important in metabolic pathways, such as glycoprotein and glycolipids processing and carbohydrate digestion in the intestinal tract. Amongst the vast variety of enzymes, glucosidases are postulated to be a powerful therapeutic target since they catalyze the cleavage of glycosidic bonds releasing glucose from the non-reducing end of an oligo- or polysaccharide chains (1). Glucosidase inhibitors are currently of interest owing to their promising therapeutic potential in the treatment of metabolic disorders such as diabetes (1).

Diabetes mellitus (DM) is a chronic disorder of metabolism caused by an absolute or relative lack of or resistance to insulin. It is characterized by exaggerated glycaemic response in postprandial and/or fasting state (2).

At the present time it is estimated that 150 million people worldwide have diabetes and this number will increase to 220 million by 2010 and 300 million by 2025. Globally, the percentage of type 2 diabetes is greater than 90% (3). Control of postprandial plasma glucose levels is critical in the early treatment of DM and in reducing chronic vascular complications (4).

One of the therapeutic approaches for reducing postprandial hyperglycemia is to prevent absorption of carbohydrates after food intake since only monosaccharides can be absorbed from the intestinal lumen and transported into the blood circulation. Complex polysaccharides must be digested by the enteric digestive enzymes including alpha glucosidase. Therefore, alpha glucosidase inhibitors such as acarbose, voglibose, and miglitol are widely used either alone, or in combination with other antidiabetic medications or insulin in patient with type 2 diabetes (5).

Screening of alpha glucosidase inhibitors from plants and synthetic sources are increasing. Inhibitors of these enzymes have recently been found from natural sources. In Mexico, 821 plant species have been registered from traditional medicine, and 51 of

them have been employed as anti-diabetic agent (4).

In addition glucosidase inhibitors have therapeutic potential in the treatment of disorders such as human immunodeficiency virus (HIV) infection, metastatic cancer, and lysosomal storage diseases (1).

Therefore, search for alpha glucosidase inhibitor, especially in plants give rise to a reliable, cheap and safe medicine in the management and control of diabetes and other diseases.

Materials and Methods

Plants

Different parts of plants, such as flowers, fruits, leaves, aerial parts, roots or seeds with known or unknown antidiabetic effect were collected from various provinces throughout Iran or purchased from the medicinal herbal markets in Kerman city and all of them were botanically identified. A voucher specimen was deposited at the herbarium of the Herbal Medicines Research Center Faculty of Pharmacy, Kerman University of Medical Sciences, Iran (Table 1).

Methanolic and aqueous extracts were prepared from 20 g of air-dried tissue of each plant, pulverized by maceration in 200 ml methanol or distilled water at room temperature for 24 hrs. After filtration, extracted methanolic substrates were air dried and the aqueous extracts dried at 40 °C in incubator. The dried materials either powdered or waxy shaped compound were kept in the dark vials at -20 °C.

Enzyme assay

P-Nitrophenyl- α -D-Glucopyranoside (PNPG) and Bakers yeast alpha glucosidase were purchased (Sigma, USA). Yeast alpha glucosidase has frequently been used to investigate the inhibitory activity of traditional medicinal plants (6).

The enzymatic activities of alpha glucosidase were determined colorimetrically by monitoring the release of p-nitrophenol from the appropriate p-nitrophenol glycoside substrate (3). The assay mixture for these experiments contained 5 μ mol PNPG, enzyme

solution (0.1 U), in 900 μ L of sodium phosphate buffer (50 mM), pH 6.8 in the final volume of 1 ml. Each extract 100 μ g was dissolved in 20 μ L of distilled water and added to the test mixture before adding the substrate. Blank sample contained whole test mixture and the extract without enzyme solution. Distilled water added to the control sample (20 μ L), and in the positive control 20 μ L acarbose (100 μ g) was enhanced.

The mixture incubated at 37 °C for 30 mins, the reaction terminated by adding 3 volumes of NH₄OH solution (0.05 M). The absorbance at 405 nm was determined by NOVA spectrophotometer (LKB, Sweden).

The inhibitory activity calculated using following formula (7):

$$\text{Inhibitory activity (\%)} = (\text{OD}_{\text{control}} - \text{OD}_{\text{test}}) / \text{OD}_{\text{control}} \times 100$$

Each test performed 3 times and the mean value was used for the inhibitory activity of plants extracts.

Results

Positive control

Acarbose, a known drug for alpha glucosidase inhibition, at the concentration used in this study showed 51% inhibitory effect on the alpha glucosidase. The results are summarized in Table1.

Table 1: Characteristics of inhibitory effects of plants extracts on alpha glycosidase (Positive control: Acarbose (51±3)).

Plants name	Family	Used parts	Methanolic (%)	Aqueous (%)
<i>Acantholepis orientalis</i>	Asteraceae	Aerial parts	1±0.1	1±0.2
<i>Achillea eriophora</i>	Asteraceae	Aerial parts	4±1	4±1
<i>Achillea wilhelmsii</i>	Asteraceae	Aerial parts	4±0.2	3±0.3
<i>Acroptilon repens</i>	Asteraceae	Aerial parts	1±0.3	1±0.1
<i>Alhagi camelorum</i>	Fabaceae	Aerial parts	92±3	2±0.3
<i>Alpinia officinarum</i>	Zingiberaceae	Rhizomes	48±3	16±4
<i>Arctium lappa</i>	Asteraceae	Roots	2±0.1	80±3
<i>Artemisia santolina</i>	Asteraceae	Aerial parts	3±0.3	3±0.1
<i>Berberis integrimma</i>	Berberidaceae	Aerial parts	1±0.2	1±0.2
<i>Berberis integrimma</i>	Berberidaceae	Roots	2±0.4	3±0.4
<i>Biebersteinia multifida</i>	Gerariaceae	Aerial parts & fruits	1±0.1	3±0.2
<i>Brassica nigra</i>	Brassicaceae	Seeds	2±0.1	0±0
<i>Bryonia aspera</i>	Cucurbitaceae	Aerial parts	1±0.1	0±0
<i>Bunium persicum</i>	Apiaceae	Seeds	7±2	0±0
<i>Camellia sinensis</i>	Theaceae	Leaves	95±3	85±1
<i>Cannabis sativa</i>	Cannabaceae	Seeds	2±0.6	0±0
<i>Cardaria draba</i>	Brassicaceae	Aerial parts & flowers	2±3	3±0
<i>Carthamus oxyacantha</i>	Asteraceae	Aerial parts	0±0	3±0.2
<i>Chaerophyllum khorassanicum</i>	Apiaceae	Aerial parts	1±0.2	8±2
<i>Cichorium intybus</i>	Asteraceae	Roots	5±0.4	4±0.3
<i>Cinnamomum zeylanicum</i>	Lauraceae	Derm	100±3	98±2
<i>Citrus aurantium</i>	Rutaceae	Flowers	0±0	58±7
<i>Citrus sinensis</i>	Rutaceae	Fruits hull	0±0	5±0
<i>Convolvulus pilosellaefolius</i>	Convolvulaceae	Aerial parts	3±0.2	2±0.3
<i>Cordia mixa</i>	Boraginaceae	Fruits	3±0.3	1±0.3
<i>Crocus sativa</i>	Iridaceae	Leaves	1±0.5	0±0
<i>Cuminum cyminum</i>	Apiaceae	Seads	0±0	1±0.2
<i>Ducrosia assadii</i>	Apiaceae	Aerial parts	1±0.2	1±0

Table 1 contd.

Plants	Family	Used parts	Methanolic (%)	Aqueous (%)
<i>Echium amoenum</i>	Boraginaceae	Flowers	0±0	4±0.4
<i>Equisetum arvense</i>	Equisetaceae	Whole the plant	4±1	2±0.2
<i>Eremostachys laciniata</i>	Lamiaceae	Whole the plant	2±0.2	7±0.5
<i>Eremurus persicus</i>	Liliaceae	Aerial parts	2±0.3	3±1
<i>Eremurus persicus</i>	Liliaceae	Flowers	1±0.2	0±2
<i>Eremurus persicus</i>	Liliaceae	Fruits	1±0.3	2±0.9
<i>Euphorbia hebecarpa</i>	Euphorbiaceae	Aerial parts & flowers	9±1	4±1
<i>Ferula Assa-foetida</i>	Apiaceae	Aerial parts & flowers	2±1	0±0
<i>Ferula oopoda</i>	Apiaceae	Aerial parts	1±0.1	0±0
<i>Ferulago angulata</i>	Apiaceae	Aerial parts	11±1	4±2
<i>Ficus carica</i>	Moraceae	Leaves	3±0.3	4±0.5
<i>Foeniculum vulgare</i>	Apiaceae	Fruits	0±0	1±0.3
<i>Francoeuria undulata</i>	Asteraceae	Aerial parts	3±0.3	6±0.4
<i>Fumaria parviflora</i>	Fumariaceae	Aerial parts	3±0.2	3±0.3
<i>Glycyrrhiza glabra</i>	Fabaceae	Aerial parts	41±3	5±2
<i>Gundelia tournefortii</i>	Asteraceae	Aerial parts	1±0	0±0
<i>Heracleum persicum</i>	Apiaceae	Fruits	4±0.4	1±0.2
<i>Hibiscus gossypifolius</i>	Malvaceae	Flowers	0±0	1±0
<i>Hyoscyamus senecionis</i>	Solanaceae	Aerial parts & flowers	3±0.2	4±0.4
<i>Hypecoum pendulum</i>	Fumariaceae	Aerial parts	10±0.5	1±0
<i>Juglans regia</i>	Juglandaceae	Fruits hull	0±0	2±0.2
<i>Juglans regia</i>	Juglandaceae	Leaves	0±0	14±4
<i>Laurus nobilis</i>	Lauraceae	Leaves	10±0.9	4±0.4
<i>Lawsonia inermis</i>	Lythraceae	Leaves	64±4	6±2
<i>Levisticum officinale</i>	Apiaceae	Roots	98±2	89±3
<i>Linum usitatissimum</i>	Liliaceae	Seeds	0±0	0±0
<i>Malva sylvestris</i>	Malvaceae	Flowers	4±0.4	6±2
<i>Marrubium anisodon</i>	Lamiaceae	Aerial parts	3±0.2	6±0.5
<i>Mentha longifolia</i>	Lamiaceae	Aerial parts	3±1	30±4
<i>Mentha piperita</i>	Lamiaceae	Leaves	0±0	90±2
<i>Myrtus communis</i>	Myrtaceae	Leaves	97±3	99±3
<i>Nepeta crispa</i>	Lamiaceae	Aerial parts	0±0	0±0
<i>Nepeta saccharata</i>	Lamiaceae	Whole the plant	0±0	0±0
<i>Nigella sativa</i>	Ranunculaceae	Seeds	0±0	2±0.3
<i>Onobrychis viciifolia</i>	Fabaceae	Aerial parts	0±0	1±0.1
<i>Otostegia persica</i>	Lamiaceae	Aerial parts	0±0	2±0.1
<i>Outreya carduiiformis</i>	Asteraceae	Aerial parts	0±0	0±0
<i>Peganum harmala</i>	Zygophyllaceae	Aerial parts	0±0	0±0
<i>Peucedanum Aucheri</i>	Apiaceae	Roots	0±0	2±0.2
<i>Pimpinella anisum</i>	Apiaceae	Seeds	0±0	2±0.1
<i>Piper nigrum</i>	Piperaceae	Fruit	3±0.3	5±0.3
<i>Pistacia vera</i>	Anacardiaceae	Fruits hull	90±4	54±7

Table 1 contd.

Plants	Family	Used parts	Methanolic (%)	Aqueous (%)
<i>Punica granatum</i>	Lythraceae	Fruits hull	93±1	83±4
<i>Quercus infectoria</i>	Fagaceae	Galls	98±2	96±4
<i>Rosa damascena</i>	Rosaceae	Floret	98±1	81±3
<i>Rosmarinus officinalis</i>	Lamiaceae	Aerial parts	65±2	81±5
<i>Rubia tinctorium</i>	Rubiaceae	Roots	6±2	1±0.1
<i>Salvadora persica</i>	Salvadoraceae	Wood	53±5	0±0
<i>Salvia rhytidea</i>	Lamiaceae	Whole the plant	0±0	0±0
<i>Scrophularia frigida</i>	Scrophulariaceae	Aerial parts	6±2	5±0.3
<i>Sanguisorba minor.</i>	Rosaceae	Aerial parts	92±3	64±8
<i>Scrophularia striata</i>	Scrophulariaceae	Aerial parts	1±0	0±0
<i>Solanum dulcamara</i>	Solanaceae	Fruits	0±0	0±0
<i>Sonchus asper</i>	Asteraceae	Aerial parts	3±0.2	2±0.2
<i>Sophora alopecuroides</i>	Fabaceae	Aerial parts	3±0.4	0±0
<i>Stachys inflata</i>	Lamiaceae	Aerial parts	2±0.3	7±0.3
<i>Stachys lavandulifolia</i>	Lamiaceae	Aerial parts	6±0.4	3±0.4
<i>Terminalia chebulla</i>	Combretaceae	Fruits	90±3	94±2
<i>Teucrium polium</i>	Lamiaceae	Aerial parts	7±0.3	1±0
<i>Teucrium scordium</i>	Lamiaceae	Aerial parts	4±0.9	1±0
<i>Thymus serpyllum</i>	Lamiaceae	Aerial parts	0±0	67±3
<i>Trigonella foenum graecum</i>	Fabaceae	Seeds	3±0.3	0±0.5
<i>Urtica dioica</i>	Urticaceae	Aerial parts	2±0.2	4±0.3
<i>Urtica urens</i>	Urticaceae	Aerial parts	2±0.3	11±2
<i>Vaccinium arcto-staphylus</i>	Ericaceae	Fruits	98±3	49±4
<i>Valeriana hispida</i>	Valerinaceae	Rhizomes	1±0.9	1±0.4
<i>Verbascum kermanensis</i>	Scrophulariaceae	Leaves	52±3	72±9
<i>Verbascum songaricum</i>	Scrophulariaceae	Aerial parts	0±0	1±0.9
<i>Zataria multiflora</i>	Lamiaceae	Aerial parts	0±0	89±3
<i>Zhumeria majdae</i>	Lamiaceae	Leaves	63±4	29±4
<i>Zingiber officinale</i>	Zingiberaceae	Rhizomes	10±3	5±1
<i>Ziziphus spina-christi</i>	Rhamnaceae	Leaves	72±4	50±3

Extracts with 75-100% inhibitory effect on the alpha glucosidase (high potency)

We found that both methanolic and aqueous extract of *Levisticum officinale*, *Myrtus communis*, *Rosa damascene*, *Terminalia chebulla*, *Punica granatum*, *Quercus infectoria*, *Camellia sinensis*, *Cinnamomum zeylanicum*, had more than 75% inhibitory effect on alpha glucosidase. In addition, the aqueous extract of *Arctium lappa*, *Mentha piperita*, *Rosmarinus officinalis*, *Zataria multiflora* and methanolic extract of *Alhagi camelorum*, *Pistacia vera*, *Sanguisorba minor* *Vaccinium arcto-staphylus* had shown more

than 75% inhibitory activity on alpha glucosidase.

Extracts with 50-75% inhibitory effect on the alpha glucosidase (moderate potency)

In our study the aqueous and methanolic extracts of *Verbascum kermanensis* and *Ziziphus spina-christi* and the aqueous extract of *Sanguisorba minor*, *Thymus serpyllum*, *Citrus aurantium* and methanolic extract of *Salvadora persica*, *Rosmarinus officinalis*, *Zhumeria majdae*, *Lawsonia inermis* showed 50%-75% inhibitory effect on the alpha glucosidase.

Extracts with 25-50% inhibitory effect on the alpha glucosidase (low potency)

The aqueous extract of *Zhumeria majdae*, *Verbascum kermanensis* and methanolic extracts of *Glycyrrhiza glabra* and *Alpinia officinarum*, had 25%-50% inhibitory activity on the alpha glucosidase.

Extracts with less than 25% or no inhibitory effect on the alpha glucosidase

Aqueous extract of *Juglans regia*, *Urtica urens* and methanolic extracts of *Laurus nobilis*, *Hypocoum pendulum* demonstrated 14%, 11%, 10% and 10% inhibitory activity respectively and the rest of plants extracts showed <10% or no inhibitory activity in this study. No extract was found to enhance the enzyme activity.

Discussion

Postprandial hyperglycemia could induce the non-enzymatic glycosylation of various proteins; resulting in the development of chronic complications. Therefore, control of postprandial plasma glucose levels is critical in the early treatment of DM and in reducing chronic vascular complications (4). Inhibition of enzymes involved in the metabolism of carbohydrates including alpha-glucosidase is one of the therapeutic approaches for reducing postprandial hyperglycemia (5).

Screening of plants for enzyme inhibitors such as angiotensin converting enzyme (ACE) and alpha glucosidase despite their natural constituents, have long been performed (8, 9). Although these plants are rich in tannin contents which may have an effect on the enzyme activity, no attempt have been made to relate these activities to their tannin content. Still many compounds have shown to be responsible for their functional activities. In this research there was no intention to correlate the activity of the crude extract to any active compound. Instead there were found many plants with high content of tannin which showed no or less inhibitory activity of glucosidase, showing no direct correlation with their tannin content as found by Cope *et al* (10). Beside that the inhibition was almost specific to this enzyme and when

β -glucosidase, β -galactosidase and alpha mannosidase were examined, no inhibitory effect was found against these enzymes (results not shown).

In most cases the *in vitro* results of active plants showed a very good correlation with that of *in vivo* studies (paper submitted). Lineweaver-burk plot analysis also showed a non competitive inhibition mode (results not shown) that is another phenomena for unique inhibitory action of the extract regardless of tannin content.

In this study we found 38 plants extracts with more than 50% inhibitory effect on the alpha glucosidase which might be effective in the diabetes control, Also this study manifested the dose dependent inhibitory effect of *Levisticum Officinale* on the postprandial glucose level.

Myrtus communis L. leaves as well as the volatile oil obtained from the leaves have been used to lower the blood glucose level in type-2 diabetic patients in Turkish folk medicine (11) and methanolic extract of *Terminalia chebula* are being used extensively in Indian herbal medicine. Oral administration of the extracts reduced the blood sugar level significantly in normal and in alloxan diabetic rats (12).

A prospective epidemiological study which was done in Japan found that men and women who drink 6 or more cups of green tea (*Camellia sinensis*) per day had one-third lower incidence of type 2 diabetes mellitus over 5 years (13). In another study, administration of green tea suppressed the elevation of serum glucose level significantly in non-diabetic rats (14).

In our study, aqueous and methanolic extract of *Myrtus communis*, *Terminalia chebula* and *Camellia sinensis* had a strong inhibitory effect on the alpha glucosidase (more than 90%). This may indicate that at least one reason for the reduction of blood glucose level by these plants. It could be consequence of a decrease in glucose absorption by the inhibiting this enzyme.

The extract of *Pistacia atlantica* have shown significant alpha amylase inhibitory activity (15). In our study the methanolic extract from fruits hull of *Pistacia vera* has

strong inhibitory effect (97%) on the alpha glucosidase, whereas its aqueous extract had moderate potency (61%). The difference between two methods of extraction could be due to different constituents of the fractions as seen for antioxidant properties of *Myrtus communis* L. extracts (16).

Glycyrrhiza glabra reduced the hyperphagia and polydipsia but did not alter the hyperglycaemia or hypoinsulinaemia (17). In contrast, Kimura reported that the water extract of *Glycyrrhiza glabra* markedly lowered blood glucose levels in diabetic animals (18).

We showed that methanolic extract of *Glycyrrhiza glabra* had low inhibitory activity (40%) for alpha glucosidase whereas its aqueous extract showed no inhibitory effect. *Zizyphus spina-christi* is one of the plants commonly used in Egyptian folk medicine for the treatment of different diseases (19). It has been shown that treatment of normal rats produced insignificant changes in some enzymes involved in carbohydrate metabolism. However, in diabetic rats, treatments with this herb significantly reduced the serum glucose level, liver phosphorylase and glucose-6-phosphatase activities, and significantly increased the serum pyruvate level and liver glycogen content (19).

In this study the aqueous and methanolic extract of *Zizyphus spina-christi* showed 53% and 76% inhibitory effect on the alpha glucosidase, respectively. So this plant might have other mechanism for reduction of blood glucose.

Aqueous extracts of *Cinnamomum zeylanicum* can reduce blood glucose concentration in diabetic rats and it seems that the cinnamon effect is probably linked to the potentiating action of cinnamon on insulin (20). Whereas part of this phenomenon could be related to a very strong inhibitory effect of this plant on the alpha glucosidase, showed in this study.

In Ayurvedic medicine almost all parts of *Punica granatum* (PG) have been used for the treatment of numerous disorders. Only PG flowering part (PGF), however have been recommended in Unani literature as a remedy

for diabetes. PGF extract demonstrated a potent inhibitory effect on the alpha-glucosidase activity. The inhibition was dependent on the concentration of enzyme and substrate, as well as on the pretreatment length of the enzyme (3) which are similar to our findings that showed by fruits derm of *Punica granatum* and this part of palnts have strong inhibitory effect on the alpha glucosidase.

Arctium lappa (21) and *Mentha piperita* (22) demonstrated anti diabetic effect. We showed that the aqueous extract of these have strong inhibitory effect on the alpha glucosidase and seems reasonable that at least one mechanism for their antidiabetic effect is an inhibitory action.

Trigonella foenum greacum, have been reported to be beneficial for treating type 2 diabetes. The stimulating or regenerating effect on beta cells or extra pancreatic effects were proposed for the hypoglycemic action of this herb (23). In contrast, *Trigonella foenum greacum* had no inhibitory effect on the alpha glucosidase in our study. This phenomenon was true for *Urtica dioica* (24), *Ficus carica* (25), *Peganum harmala* (26), *Laurus nobilis*(27), *Teucrium polium*(28), *Zingiber officinale* (29) and *Cuminum cyminum* (30) which previously had shown anti diabetic effect. So other alternative mechanism's for the hypoglycemic effect of these plants must be involved.

A component isolated from the methanolic extract of the galls of *Quercus infectoria*, significantly inhibited the alpha-glycosidases. The inhibitory activity on alpha-amylase was approximately 10 times lower than that of acarbose. So this plant might reduce the side effects, by reducing the inhibition of alpha-amylase. Our results indicated that the aqueous extract of *Quercus infectoria* can inhibit the alpha glucosidase as well as its methanolic extract (31).

Other plants such as *Verbascum kermanensis*, *Rosa damascene*, *Citrus aurantium*, *Rosmarinus officinalis*, *Levisticum officinale*, *Zataria multiflora*, *Sanguisorba minor*, *Alhagi camelorum*, *Pistacia vera*, *Vaccinium arcto-staphylus*, *Zhumeria majdae*, *Alpinia officinarum*, *Salvadora persica*,

Lawsonia inermis and *Thymus serpyllum* which presented more than 50% inhibitory effect on the alpha glucosidase in this study.

Conclusion

There was no record in the literature for their anti-diabetic effect but they might be a good candidate for the control of diabetes. It is necessary to investigate the effects of active

plants on blood glucose and the probable role of them on the insulin secretion; some of which are under investigation.

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References

1. Melo EBd, Gomesb AdS, Carvalhob I. Alpha- and beta-glucosidase inhibitors: chemical structure and biological activity. *Tetrahedron* 2006; 62:10277–10302.
2. Zanatta L, de Sousa E, Cazarolli LH, Junior AC, Pizzolatti MG, Szpoganicz B, et al. Effect of crude extract and fractions from *Vitex megapotamica* leaves on hyperglycemia in alloxan-diabetic rats. *J Ethnopharmacol* 2007; 109:151-155.
3. Li Y, Wen S, Kota BP, Peng G, Li GQ, Yamahara J, et al. *Punica granatum* flower extract, a potent alpha-glucosidase inhibitor, improves postprandial hyperglycemia in Zucker diabetic fatty rats. *J Ethnopharmacol* 2005; 99:239-244.
4. Ortiz-Andrade RR, Garcia-Jimenez S, Castillo-Espana P, vila GR-A, Villalobos-Molina R, Estrada-Soto S. Alpha-Glucosidase inhibitory activity of the methanolic extract from *Tournefortia hartwegiana*: An anti-hyperglycemic agent. *J Ethnopharmacol* 2007; 109: 48–53.
5. Shim YJ, Doo HK, Ahn SY, Kim YS, Seong JK, Park IS, et al. Inhibitory effect of aqueous extract from the gall of *Rhus chinensis* on alpha-glucosidase activity and postprandial blood glucose. *J Ethnopharmacol* 2003; 85:283-287.
6. Pluempunapat W, Adisakwattana S, Yibchok-Anun S, Chavasiri W. Synthesis of N-phenylphthalimide derivatives as alpha-glucosidase inhibitors. *Arch Pharm Res* 2007; 30:1501-1506.
7. Bhandari MR, Jong-Anurakkun N, Hong G, Kawabata J. A-glucosidase and a-amylase inhibitory activities of Nepalese medicinal herb Pakhanbhed (*Bergenia ciliata*, Haw.). *Food Chem* 2008; 106:247–252
8. Bhandari MR, Jong-Anurakkun N, Hong G, Kawabata J. A-Glucosidase and a-amylase inhibitory activities of Nepalese medicinal herb Pakhanbhed (*Bergenia ciliata*, Haw.). *Food Chem* 2008; 106:247–252.
9. Duncan AC, Jager AK, Vanstaden J. Screening of Zulu medicinal plants for angiotensin converting enzyme (ACE) inhibitors. *J Ethnopharmacol* 1999; 68:63-70.
10. Cope WA, Bell TA, Smart WWG. Changes in an enzyme inhibitor and tannin content in *Serica lespezda*. *Crop Sci Society Am* 1971; 11:893-895.
11. Sepici A, Gurbuz I, Cevik C, Yesilada E. Hypoglycaemic effects of myrtle oil in normal and alloxan-diabetic rabbits. *J Ethnopharmacol* 2004; 93:311-318.
12. Sabu MC, Kuttan R. Anti-diabetic activity of medicinal plants and its relationship with their antioxidant property. *J Ethnopharmacol* 2002; 81:155-160.
13. Mackenzie T, Leary L, Brooks WB. The effect of an extract of green and black tea on glucose control in adults with type 2 diabetes mellitus: double-blind randomized study. *Metabolism* 2007; 56:1340-1344.
14. Sabu MC, Smitha K, Kuttan R. Anti-diabetic activity of green tea polyphenols and their role in reducing oxidative stress in experimental diabetes. *J Ethnopharmacol* 2002; 83:109-116.
15. Afifi FU, Al-Khalidi B, Khalil E. Studies on the in vivo hypoglycemic activities of two medicinal plants used in the treatment of diabetes in Jordanian traditional medicine following intranasal administration. *J Ethnopharmacol* 2005; 100:314 -318.
16. Romani A, Coinu R, Carta S, Galardi C, Vincieri FF, Franconi F. Evaluation of antioxidant effect of different extract of *Myrtus communis* L. *Free Radic Res* 2004; 38:97-103
17. Swanston-Flatt SK, Day C, Bailey CJ, Flatt PR. Traditional plant treatments for diabetes. Studies in normal and streptozotocin diabetic mice. *Diabetologia* 1990; 33:462-464.
18. Kimura I, Nakashima N, Sugihara Y, Fu-jun C, Kimura M. The antihyperglycaemic blend effect of traditional chinese medicine byakko-ka-ninjin-to on alloxan and diabetic KK-CA(y) mice. *Phytother Res* 1999; 13:484-488.
19. Glombitza KW, Mahran GH, Mirhom YW, Michel KG, Motawi TK. Hypoglycemic and antihyperglycemic effects of *Zizyphus spina-christi* in rats. *Planta Med* 1994; 60:244 -247.
20. Gheibi N, Hashemi HJ, Parvizi M. the effect of cinnamon on glucose concentration of diabetic rats in presence or absence of insulin. *Qazvin Univ. Med. Sci* 2005; 9:2-7.
21. Swanston-Flatt SK, Day C, Flatt PR, Gould BJ, Bailey CJ. Glycaemic effects of traditional European plant treatments for diabetes. Studies in normal and streptozotocin diabetic mice. *Diabetes Res* 1989; 10:69-73.

22. Narendhirakannan RT, Subramanian S, Kandaswamy M. Mineral content of some medicinal plants used in the treatment of diabetes mellitus. *Biol Trace Elem Res* 2005; 103:109-115.
23. Saxena A, Vikram NK. Role of selected Indian plants in management of type 2 diabetes: a review. *J Altern Complement Med* 2004; 10:369-378.
24. Farzami B, Ahmadvand D, Vardasbi S, Majin FJ, Khaghani S. Induction of insulin secretion by a component of *Urtica dioica* leave extract in perfused islets of Langerhans and its in vivo effects in normal and streptozotocin diabetic rats. *J Ethnopharmacol* 2003; 89:47-53.
25. Pérez C, Domínguez E, Ramiro JM, Romero A, Campillo JE, Torres MD. A study on the glycaemic balance in streptozotocin-diabetic rats treated with an aqueous extract of *Ficus carica* (fig tree) leaves. *Phytother Res* 1998; 10:82-83.
26. Hussain Z, Waheed A, Qureshi RA, Burdi DK, Verspohl EJ, Khan N, *et al.* The effect of medicinal plants of Islamabad and Murree region of Pakistan on insulin secretion from INS-1 cells. *Phytother Res* 2004; 18:73-77.
27. Broadhurst CL, Polansky MM, Anderson RA. Insulin-like biological activity of culinary and medicinal plant aqueous extracts in vitro. *J Agric Food Chem* 2000; 48:849-852.
28. Esmaeili MA, Yazdanparast R. Hypoglycaemic effect of *Teucrium polium*: studies with rat pancreatic islets. *J Ethnopharmacol* 2004; 95:27-30.
29. Kar A, Choudhary BK, Bandyopadhyay NG. Comparative evaluation of hypoglycaemic activity of some Indian medicinal plants in alloxan diabetic rats. *J Ethnopharmacol* 2003; 84:105-108.
30. Talpur N, Echard B, Ingram C, Bagchi D, Preuss H. Effects of a novel formulation of essential oils on glucose-insulin metabolism in diabetic and hypertensive rats: a pilot study. *Diabetes Obes Metab* 2005; 7:193-199.
31. Hwang JK, Kong TW, Baek NI, Pyun YR. Alpha-glycosidase inhibitory activity of hexagalloylglucose from the galls of *Quercus infectoria*. *Planta Med* 2000; 66:273-274.