Iranian Journal of Basic Medical Sciences

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Liver ischemia preconditions the heart against ischemiareperfusion arrhythmias

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ARTICLE INFO

Article type:

Original article

Article history:

Received: Feb 21, 2014 Accepted: Sep 27, 2014

Keywords:

Ischemia-reperfusion arrhythmia Isolated heart Liver Remote ischemic preconditioning

ABSTRACT

Objective(s): This study aimed to examine the hypothesis that an antiarrhythmic effect might be obtained by ischemic preconditioning of the liver, and also to characterize the potential underlying mechanisms.

Materials and Methods: Male Wistar rats were anesthetized by thiopental sodium (50 mg/kg, IP) followed by IV injection of heparin (250 IU). Remote ischemic preconditioning (RIPC) was induced by 3 cycles of 5 min liver ischemia followed by 5 min of reperfusion. The hearts were excised within 5 min after the final cycle of preconditioning and perfused using Langendorff's system. The isolated perfused hearts were subjected to 30 min global ischemia followed by 90 min reperfusion. The myocardial arrhythmias induced by ischemia- reperfusion (I/R) were determined in accordance with the guidelines of Lambeth Conventions. The potential role of K_{ATP} channels on RIPC was assessed by injection of glibenclamide (nonselective K_{ATP} blocker) or 5-hydroxydecanoate (mitochondrial K_{ATP} blocker) on rats 30 and 15 min before induction of RIPC in the liver, respectively.

Results: Hepatic remote preconditioning of the heart significantly (P<0.0001) prevented the incidence of myocardial arrhythmias induced by I/R in the perfused hearts (5.33±1.54 vs. 32.33±6.44,). However, the protective effects of remote preconditioning was significantly (P<0.01) abolished by the K_{ATP} blocker, glibenclamide (25.5±4.9 vs. 5.33±1.54,).

Conclusion: Hepatic RIPC may prevent the arrhythmias induced by I/R in the isolated perfused hearts via K_{ATP} channels.

► Please cite this paper as:

Noorbakhsh MF, Arab HA, Kazerani HR. Liver ischemia preconditions the heart against ischemia-reperfusion arrhythmias. Iran J Basic Med Sci 2015; 18:80-88.

Introduction

Severe ventricular arrhythmias represent a major challenge for therapeutic intervention due to complexity of pathophysiological mechanisms initiating arrhythmias in the ischemic heart disease (1), so new strategies are urgently needed to prevent and manage the condition. In the ischemiareperfusion (I/R) condition, the imbalance between the production of reactive oxygen species (ROS) and the availability of endogenous antioxidants plays an important role in the genesis of myocardial injury (2-4). This can result in malignant I/R-induced arrhythmias (5, 6). Deleterious effect of an oxidative load has been shown by the ability of exogenous free radical scavengers to improve functional recovery of the post-ischemic reperfused heart (7-9). Different approaches have been developed to prevent myocardial arrhythmias induced by I/R in the heart. It has also been shown that myocardial I/R injury can be dramatically reduced by subjecting the heart to one or more episodes of non-lethal myocardial I/R prior to the sustained coronary artery occlusion (10). This endogenous cardioprotective phenomenon termed as ischemic preconditioning (IPC) exerts different beneficial effects including decreased myocardial damage (11-14), improvement of functional recovery (15), and antiarrhythmic effects (16-18). However, the IPC is an invasive procedure being applied directly to the heart tissues in order to obtain myocardial protection. This treatment in some clinical settings can be impractical and may cause harmful effects. An alternative and more compliant strategy is to apply the cardioprotective stimulus to an organ or tissue far from the heart (19). The classical preconditioning at a distance in the heart itself is induced by a brief occlusion of one coronary artery followed by prolonged occlusion of other coronary artery (20). This approach entitled as remote ischemic preconditioning (RIPC) can dramatically prevent the myocardial injury induced by I/R (21, 22).

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Although the exact molecular mechanisms responsible for cardioprotection afforded by IPC are unclear, the results of some studies suggest that ATPsensitive K⁺ (K_{ATP}) channels can be the key players in this process (23-25). Opening of sarcolemmal K_{ATP} channels has been initially proposed as an endmechanism the IPC in Glibenclamide, a KATP channel blocker, has been able to abolish the IPC protection in dogs (26). Moreover, it has been reported that K_{ATP} openers can also mimic the IPC-induced protection (23, 27). However, there is evidence that K_{ATP} channel modulators may exert both anti- and pro-arrhythmic effects (28), other reports suggested though that cardioprotection may occur independent of any alteration in the action potential duration as the main determinant of arrhythmogenesis and a target for sarcolemmal KATP openers (29). It has been shown that 5-hydroxydecanoate (5-HD), a selective mitochondrial K_{ATP} (mito K_{ATP}) inhibitor, abolished protection against contractile dysfunction in guinea pig papillary muscle conferred by hypoxic PC (30). However, a cardioprotection has been induced by IPC (31) in rabbits and rats without affecting the shortened action potential (32). Garlid et al reported that mitochondrial (mito) K_{ATP} channel is 2000-fold more sensitive than the sarcolemmal one to the K_{ATP} opener diazoxide, which is the most likely endeffector involving in IPC (33). It seems that the opening of KATP channel not only plays as the endeffector in the preconditioning cascade, but also induces an upstream mechanism of protein kinase activation (34). It has been suggested that some underlying pathways and subsequent signal transductions activated in remotely preconditioned cardiomyocytes may be similar to those recruited in IPC procedure (35). Steen et al have reported that the cardioprotection induced by RIPC is exerted via mito K_{ATP} channels (36), but the mechanism of the antiarrhythmic protection induced by mito K_{ATP} opening remains still unclear.

The cardioprotective effect of RIPC is under intensive investigations and many unsolved problems remains to be examined. There is no evidence whether a protection against the myocardial arrhythmias induced by I/R could be obtained by preceding previous brief ischemic episodes of the liver as RIPC for the heart. Therefore, the present study was conducted to examine the hypothesis that hepatic RIPC can induce antiarrhythmic effects on I/R-exposed isolated hearts through the opening of K_{ATP} channels.

Materials and Methods Animals

Male Wistar rats (250 to 300 g) were purchased and housed in standard conditions (12 hr light/day cycle with 20 to 22°C temperature and 40 to 50% humidity). The animals had access to commercial

chow and water *ad libitum*. The project had prior approval from the Institutional Animal Care and Use Committee, and all procedure conducted on the animal was in accordance with the guidelines described by the Helsinki Declaration, as revised in Edinburgh 2000.

The surgery for hepatic ischemic preconditioning and isolated rat heart preparations

The animals were anesthetized by thiopental sodium (50 mg/kg, IP) and then they received heparin (250 IU) from femoral artery (37). The ventral midline abdomen of the animals was excised, and the liver was exposed by disconnecting its connective ligaments to the abdominal wall. The hepatic RIPC was induced by intermittent portal triad clamping for 5 min episodes (38). The heart was isolated immediately after the final episode of hepatic ischemia, and then perfused using Langendorff's setup. To achieve this, the heart was removed; the aorta was cannulated for retrograde perfusion at a constant flow rate of 10 ml/min with a non-recirculating Krebs-Henseleit buffer solution (KHB). The KHB solution contained (mmol/l) NaCl 118.0, NaHCO₃ 25.0, KCl 4.7, KH₂PO₄ 1.2, MgSO₄.7H₂O 1.2, CaCl₂ 1.25, and glucose 11.0, which was saturated with a mixture of 95% O₂ and 5% CO₂ and kept at pH =7.4 and 37°C. A global ischemia was induced by occlusion of the KHB inflow (30 min) followed by reperfusion for 90 min (37). To monitor the occurrences of I/R induced arrhythmias, two stainless steel electrodes were connected to the apex and right atrium of the heart (39). The coronary perfusion pressure (CPP) was measured through a three-way stop cock using a pressure transducer (MLT844 Physiological Pressure Transducer, ADInstruments). Left ventricular pressure was measured by an elastic water-filled balloon inserted into the left ventricle via the left atrium (adjusted to obtain end-diastolic pressure of 10 mmHg) and connected to a pressure transducer (MLT844 Physiological Transducer, AD Instruments) (40).

Experimental protocol

Rats were randomly divided into eight experimental groups of six rats in each. In group I chosen as sham-operated, the isolated heart was perfused for a period of 140 min. In group II (I/R), the isolated heart was exposed to 30 min ischemia followed by 90 min reperfusion. The procedure in group III (control) was the same as I/R group except that a sham operation was performed on the liver for RIPC. Group IV (treatment) underwent a hepatic RIPC by 3 cycles of 5 min occlusion of the portal triad prior to myocardial I/R. The rats in group V were treated the same as the group III but received 0.3 mg/kg glibenclamide (Gliben) through IV injection (36) 30 min prior to the sham operation on the liver. The procedure in group VI designated as 5-



hydroxydecanoate (5-HD) treated control group, was the same as in group III but the rats were treated with 5 mg/kg of IV 5-HD (41) 15 min before sham operation on the liver. Group VII (Gliben treated treatment group) received 0.3 mg/kg Gliben through IV 30 min before induction of RIPC in the liver, and

the remaining protocol was the same as described in group IV. In group VIII labeled as 5-HD treated treatment group, the rats were administered 5-HD (5 mg/kg, IV) 15 min before induction of RIPC, and then the procedure was performed the same as described in group IV.

Table 1. Pre-ischemic and post-ischemic values of hemodynamic parameters in different groups of isolated perfused rat hearts. The ischemia-reperfusion was induced in the isolated heart by 30 min ischemia followed by 90 min reperfusion, the remote ischemic preconditioning was induced by 3 cycles of 5 min liver ischemia followed by 5 min of reperfusion, and the K_{ATP} channel blocker was injected into the animal 30 and 15 min before induction of remote ischemic preconditioning

	Preischemia	End of ischemia	Reperfusion time (min)				
			2	15	30	60	90
HR							
Sham	216.8±4.1	209.7±11.9	211.5±8.1	212.2±7.5	208.3±9.3	197.7±10.6	193±13.4
Ischemia/Reperfusion	247.5±11.4	a0	246.2±13.4	242.2±12.5	235.6±16.6	214.47±7.2	214.5±6.3
Control	238.9±11.3	a0	236.6±17.8	217.19±13.7	210.8±17.4	214.7±5.9	203.2±4.87
Treatment	249.3±5.8	a0	260.7±17.04	232.4±16.2	241.1±10.1	232.2±7.3	226.7±11.3
Control (Gliben)	240.46±11.4	a0	239.21±13.4	235.2±12.5	228.55±16.6	207.41±7.2	207.51±6.3
Control (5-HD)	249.00±13.4	a0	226.63±12.6	225.93±9.8	227.6±12.6	220.96±14.7	205±4.8
Treatment (Gliben)	235.80±12.5	a0	239.06±11.8	224.21±15.8	209.60±9.5	211.00±9.6	207.35±9.5
Treatment (5-HD)	254.6±8.6	a0	239.14±15.7	228.80±12.2	215.16±7.1	216.18±9.9	215.33±11.4
СРР			0				
Sham	90.5±5	88.75±6.5	90.96±6.1	82.09±9.8	89.02±7.8	86.72±7.8	85.22±7.9
Ischemia/Reperfusion	94.66±1.8	a0	71.02±4.5	77.09±6.7	77.71±4.7	71.56±3.5	65.4.±1.2
control	76.42±4.3	a0	61.47±7.6	54.47±4.2	55.64±6.7	50.94±8.1	47.42±6.2
Treatment	92.69±3.6	a0	87.66±3.4	98.08±6.5	97.68±2.7	94.03±3.3	91.62±5.2
Control (Glibenclamide)	51.38±2.6	a0	50.36±2.3	48.95±1.9	53.12±3.3	50.61±3.1	50.03±3.0
Control (5-HD)	58.62±2.5	a0	55.33±1.3	53.49±1.8	55.95±3.8	53.93±2.0	50.95±2.9
Treatment (Glibenclamide)	49.97±2.7	a0	50.61±1.75	47.7±1.62	46,16±1.8	48.9±2.5	48.98±2.5
Treatment (5-HD)	53.17±4.0	a0	48.25±1.8	46.10±2.7	45.17±2.1	47.58±2.8	49.47±1.4
Max dp/dt							
Sham	2620±213	2707±186	2676± 238	2527±303	2567±270	2553±316	2476±366
Ischemia/Reperfusion	2835±88	a25±0.6	a1917±41	^b 2022±94	^b 2111±131	ab 1843±71	ab 1825.±79
Control	2254±157	a26±0.1	a1976±171	ab 1863±204	b 1983±169	ab 1651±153	ab 1653±178
Treatment	2806±127	^a 26±0.1	2353±71	3125±185	3286±30	3373±81	3204±158
Control (Glibenclamide)	3161±157	a26±0.6	2147±41	^b 2252±94	b 2341±131	2040±66b	b 1988±42
Control (5-HD)	2667±199	a26±0.2	2007±76	^b 2140±36	^b 2096±146	b 1982±85	b 1938±120
Treatment (Glibenclamide)	2390±206	^a 26±0.1	1783±170	^b 2097±253	^b 2057±173	^b 1976±235	b 1908±162
Treatment (5-HD)	2851±147	a26±0.05	1971±99	^b 2364±79	b 2383±45	^b 2146±98	^b 2138±51

Data are means \pm SEM, n = 6 in each group, HR – heart rate (beats/min), CPP– coronary perfusion pressure (ml min⁻¹), Max dp/dt (mmHg/s), a=P<0.0001vs. sham, b=P<0.05 vs. test

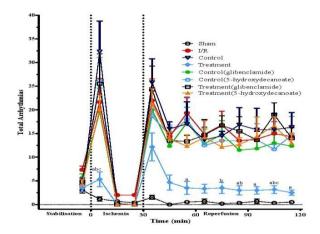


Figure 1. Effects of hepatic remote ischemic preconditioning and K_{ATP} channel blockers on the ischemia-reperfusion (I/R) -induced total arrhythmias in the isolated perfused rat heart. Values are expressed as means \pm S.E.M. of 6 isolated hearts per group in which a= P < 0.05 vs. control, b=P < 0.05 vs. treatment (glibenclamide), c=P < 0.05 vs. treatment (5-hydroxydecanoate)

Measurement and classification of ventricular arrhythmias

Ventricular arrhythmias induced by I/R in the isolated perfused rat hearts were monitored and analyzed using the data obtained from ECG recordings. Identification and classification of the I/R-induced arrhythmias in the experimental groups was based on the Lambeth Convention (15). Ventricular premature beats (VPBs) were characterized by the presence of QRS complexes not proceeded by P waves. Single isolated VPBs were defined as singles, whereas two or three consecutive VPBs were considered as salvos, and four or more consecutive VPBs were defined as ventricular tachycardia. Consistent with the Lambeth Convention, ventricular fibrillation was defined as a signal for which individual QRS complexes cannot be distinguished from each other.

Statistical analysis

Data analysis and drawing of the graphs were performed using GraphPad Prism Software v5.0 (GraphPad Software, USA). Data were expressed as mean±SEM obtained from at least six isolated perfused rat heats in each experimental group. Statistical comparisons between different experimental groups were performed using analysis of variance (ANOVA) followed by Bonferroni's *Post-hoc* test to compare the differences of means. A *P-value* less than 0.05 were considered as significant.

Results

Characteristics of isolated hearts

The values for different hemodynamic parameters in terms of heart rate, coronary perfusion pressure and max dp/dt in different experimental groups are shown in Table 1. As the table shows, there were no significant

differences among the groups in the hemodynamic values before the induction of ischemia.

Effects of hepatic RIPC on post-ischemic recovery of myocardial contractile dysfunction

There was a significant post-ischemic contractile dysfunction in the isolated perfused hearts exposed to I/R as determined by changes in the level of max dp/dt. However, the RIPC markedly (P<0.0001) attenuated post-ischemic contractile dysfunction by increasing max dp/dt recovery from 1653±178 (in the control) to 3204±158 (in the treatment group). Pretreatment of test animals with glibenclamide and 5-HD did not affect max dp/dt recovery in non-preconditioned hearts, but reversed the cardioprotective effect induced by RIPC in the Gliben and 5-HD treated treatment groups. As table 1 shows, the levels of max dp/dt at 90 min of reperfusion was significantly (P<0.05) declined from 3204±158 (in the treatment group) to 1908±162 and 2138±51 in the Gliben and 5-HD treated groups, respectively.

Effect of remote ischemic preconditioning, glibenand 5-HD on susceptibility to ventricular arrhythmias

The myocardial injury induced by I/R significantly (P<0.05) caused incidence of arrhythmia in the control and I/R groups, and the maximal occurrences was shown during 0 and 10 min of ischemia and at the beginning of reperfusion (Figure 1). The rates of single arrhythmias were also significantly (P<0.0001) higher in the I/R and control groups compared to the sham-operated group. However, there was no significant difference between the treatment and the sham groups. The application of RIPC significantly (P<0.0001) reduced single arrhythmias from 14.67±2.9 in the control group to 2.33±0.61 in the treatment group. Glibenclamide and

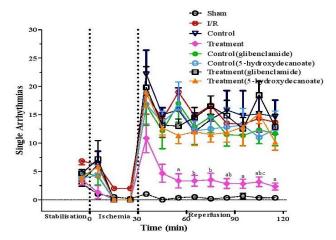


Figure 2. Effects of hepatic remote ischemic preconditioning and K_{ATP} channel blockers on ischemia-reperfusion (I/R) -induced single arrhythmias in the isolated perfused rat heart. Values are expressed as means \pm SEM of 6 hearts per group in which a= P<0.05 vs. control, b=P<0.05 vs. treatment (glibenclamide), c=P<0.05 vs. treatment (5-hydroxydecanoate)

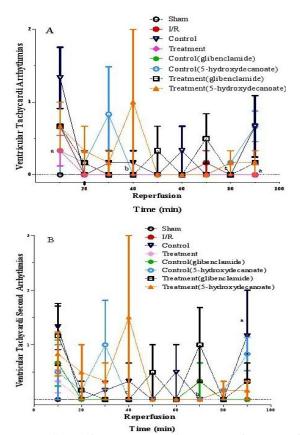


Figure 3. Effects of hepatic remote ischemic preconditioning and K_{ATP} channel blockers on ischemia-reperfusion (I/R) -induced ventricular arrhythmias in the isolated perfused rat heart. A: Number of episodes of ventricular tachycardia. B: Duration of ventricular tachycardia. VT-ventricular tachycardia. Values are expressed as means±SEM of 6 hearts per group in which a= P < 0.05 vs. control, b = P < 0.05 vs. treatment (glibenclamide), c = P < 0.05 vs. treatment(5-hydroxydecanoate)

5-HD significantly (P<0.01) abolished the antiarrhythmic effect induced by hepatic RIPC (18.5±2.40 and 15.17±1.72 vs. 3.16±0.83 in treatment). However, administration of these K_{ATP} channel blockers in the non-preconditioned hearts exposed to I/R had no effect on the incidence of single arrhythmias (Figure 2). Gliben or 5-HD in the control groups did not significantly change the incidence of salvo in explanted hearts.

Ventricular tachyarrhythmia (tachycardia and fibrillation) was the most severe form of arrhythmia that occurred in all isolated hearts exposed to I/R. Marked attenuation of arrhythmias was observed in the preconditioned hearts. However, pretreatment with Gliben or 5-HD in preconditioned hearts improvement reversed the of ventricular tachyarrhythmia induced by hepatic RIPC (Figures 3, 4). The hepatic RIPC significantly (P<0.01) decreased the mean number of the episodes of VT from 1.33 ± 0.42 to 0.33 ± 0.21 , and also shortened the total duration of VT $(0.64\pm0.33 \text{ vs. } 2.3\pm0.4 \text{ s in the control})$ Figure 3). As Figures 4A and 4B show, the exposure of rat to RIPC significantly (P<0.0001) reduced the number of episodes of VF and its total duration

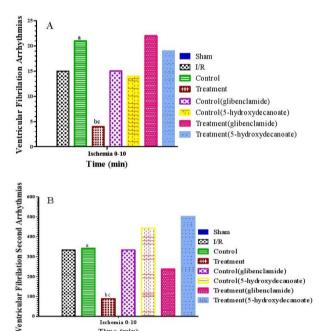


Figure 4. Effects of hepatic remote ischemic preconditioning and KATP channel blockers on ischemia-reperfusion (I/R) -induced ventricular arrhythmias in the isolated perfused rat heart. A: Number of episodes of ventricular fibrilation. B: Duration of ventricular fibrilation.VF- ventricular fibrillation. Values are expressed as means \pm S.E.M. of 6 hearts per group in which a= P < 0.0001 vs. treatment, b=P < 0.001 vs. treatment (glibenclamide), c=P < 0.001 vs. treatment (5-hydroxydecanoate)

compared to the control groups. The maximal effect of RIPC was shown between 0 and 10 min of ischemia. The total numbers of arrhythmias was also significantly (P < 0.0001) reduced in the isolated perfused hearts previously preconditioned by liver ischemia (5.33±1.54 vs. 32.33±6.44 in the control, Figure 1). Although pretreatment with glibenclamide or 5-HD in non-preconditioned hearts did not change the total number of arrhythmias, blockade of KATP before preconditioning exacerbated arrhythmias and partially attenuated the effect of RIPC. This has been shown by increased incidence of VT in treated hearts (81%) compared to the nontreated preconditioned hearts (P<0.01, Figure 3). In addition, blockade of non-selective KATP and mito K_{ATP} channels significantly (P<0.0001) increased the number of episodes of VF from 4.1±0.40 to 18.5±4.53 and 14.33±3.40 in gliben and 5-HD treatment groups, respectively (Figure 4A).

Discussion

The present study was conducted to examine the hypothesis that brief episodes of ischemia in the liver as a remote preconditioning might cause an antiarrhythmic effect against myocardial I/R injury in isolated perfused hearts and also to characterize the potential mechanism of this protective procedure. The common definition of IPC is used for a procedure in which a cardioprotective effect can be

obtained by induction of one or several episodes of ischemia in the heart against the damage induced by a subsequent prolonged period of cardiac ischemia (10). It is known that an episode (usually 3 to 5 min) of regional ischemia induced by the coronary ligation either in the intact animal or in the isolated heart is protective against the deleterious electrophysiological, biochemical, and mechanical effects of a longer ischemia within the same region of the heart (35, 13). The induction of ischemia at a distance in the heart itself by ligation of a different coronary artery has been reported to be cardioprotective (42). Such a protective effect of RIPC in general may be mediated by different endogenous substances including adenosine and calcitonin-gene related peptide released from tissues into the blood that subsequently affect the organ in danger at a distance (43-45). The antiarrhythmic effect is one of the cardioprotective outcomes of ischemic preconditioning that is well documented in various species using different experimental models (16, 18). Tatyana et al reported that brief ischemia of an extremity as a non-invasive preconditioning of the heart can be protective against reperfusiontachyarrhythmia (46). Consistently, Heidi et al demonstrated that induction of RIPC during coronary occlusion increased the ventricular arrhythmia threshold in conscious rats (47). Thus, in the present work, we used a model of the explanted heart in which protection is manifested by attenuation of I/R-induced arrhythmias to explore the possibility of obtaining the antiarrhythmic effect using RIPC of the liver.

In the present study, we found that RIPC of liver can improve the profile of arrhythmias as it significantly reduced total incidences of arrhythmias and the occurrence and the total duration of VT. In addition, cardiac performance including Max dp/dt was improved by hepatic RIPC. There is evidence that different factors including the size of the occluded zone and the changes in the heart rate may influence the occurrences of arrhythmias (48, 49). However, in the method used in the present study, the impact of the influencing factors have been excluded by exposing the experimental groups to a similar method of global ischemia, and there were no differences in the size of ischemic area between the groups correlating with the reduction of coronary flow or the heart rate (50). These findings are in agreement with the findings of other investigators (24, 46) who demonstrated an arrhythmias reduction in the preconditioned rats and rabbits.

The attenuation of spatial dispersion of repolarization between the epi- and endocardial layers of the myocardium as a substrate for reentry arrhythmias induced by ischemia has been suggested as the physiological mechanisms underlying antiarrhythmic effect induced by RIPC (51). However, the role of K_{ATP} channels in the

antiarrhythmic effects is still a matter of dispute. In the present study, we showed the involvement of K_{ATP} channels in the protection against I/R-induced arrhythmias by application of glibenclamide as a non-selective K_{ATP} blocker and 5-HD as a selective mito K_{ATP} blocker. It was found that the use of these K_{ATP} channel inhibitors before the preconditioning phase, attenuated the antiarrythmic effect induced by RIPC and increased the incidence of ventricular tachycardia. These results are consistent with those of Munch-Ellingsen *et al* and Végh and Parratt studies who found that 5-HD reduced the cardioprotection in the preconditioned rats and dogs (25, 32).

K_{ATP} channel activation may be attributed to the release of reactive oxygen species (ROS) during the preconditioning (52). It has been proposed that Superoxide (O2-) ROS generated during IPC may activate mito K_{ATP} channels through direct action on the sulfhydryl groups of the channel proteins subsequently leading to the opening of K_{ATP} channels (53). Kazuaki et al have reported that hepatocyte protection was mediated through ROS generation by Kupffer cells after IPC (54). This is in agreement with the study of Matejikova et al who reported that myocardial IPC were associated with a temporal moderate increase in generation of ROS prior to sustained ischemia (17). There is also evidence that cardioprotective effect of IPC and diazoxide may be manifested during the phase of prolonged ischemia by reduction of ROS generation and mobilization of antioxidant reserves (54-56). The use of diazoxide and preconditioning is found to be associated with an improved mitochondrial recovery after I/R injury (58), and that myocardial IPC attenuated ROS production at the end of sustained ischemia (17). There is also evidence that hepatic ischemic preconditioning is able to prevent excessive ROS generation and subsequent injury induced by IR in the liver and lung (59, 60). It seems that opening of the mito K_{ATP} channel occurs upstream of the mitochondrial activities and the generation of ROS in the protective pathway. Since this protection mechanism is abolished by two free radical scavengers, 2-mercapto-propionylglycine and nacetyl cysteine (53, 61-63).

The activation of K_{ATP} channels as a final step in the cardioprotective signaling mechanisms has been supported by the findings that activated protein kinase C (PKC) phosphorylates sarcolemmal K_{ATP} channels (64), and that NO, PKC and mitogenactivated protein kinase (MAPK)-mediated mechanisms facilitate the opening of mito K_{ATP} channels (65, 66). It is possible that the opening of a K_{ATP} channels may act as both a trigger and a mediator of preconditioning (67, 36). Different mechanisms that may be involved in this process include depolarization of the mitochondrial inner



membrane in conjunction with limitation of calcium uptake by mitochondria (68), regulation of mitochondrial volume and rate of respiration (69) and modulation of ROS production (67). The regulation of antiapoptotic proteins in the mitochondria has been proposed as another potential mechanism of attenuation of cell death induced by opening of mito K_{ATP} channels (70).

Conclusion

This study showed that short episodes of liver ischemia followed by reperfusion caused significant cardioprotective effect against the I/R- induced arrhythmias in the isolated rat heart similar to the preconditioning of the heart with regional ischemia in the heart itself. In an attempt to characterize the potential mechanism of this endogenous protective procedure, it was found that the protection process is dependent on the mito K_{ATP} channels activities. These findings provide evidence that the short episodes of liver ischemia can precondition the heart against the I/R- induced arrhythmias.

Acknowledgment

The results described in this paper were part of student thesis. We appreciate the University of Tehran for financial support. We are also grateful to Mohammad Noghre'ei for his technical assistance.

References

- 1. Bril A. Cellular mechanisms of cardiac arrhythmias in the ischemic and reperfused heart. EXS 1996; 76:135-153.
- 2. Mccord JM. Oxygen-derived free radicals in postischemic tissue injury. N Engl J Med 1985; 312:159-163.
- 3. Kloner RA, Przyklenk K, Whittaker P. Deleterious effects of oxygen radicals in ischemia/reperfusion. Resolved and unresolved issues. Circulation 1989; 80:1115-1127.
- 4. Dhalla NS, Elmoselhi AB, Hata T, Makino N. Status of myocardial antioxidants in ischemia-reperfusion injury. Cardiovasc Res 2000; 47:446-456.
- 5. Yang CS, Tsai PJ, Chou ST, Niu YL, Lai JS, Kuo JS. The roles of reactive oxygen species and endogenous opioid peptides in ischemia-induced arrhythmia of isolated rat hearts. Free Radic Biol Med 1995; 18:593-598.
- 6. Ravingerova T, Slezak J, Tribulova J, Dzurba A, Uhrik B, Ziegelhoffer A. Reactive oxygen species contribute to high incidence of reperfusion-induced arrhythmias in isolated rat heart. Life Sci 1999; 65:1927-1930.
- 7. Downey J, Omar B, Ooiwa H, Mccord J. Superoxide dismutase therapy for myocardial ischemia. Free Radic Res Commun 1991; 12:703-720.
- 8. Tang LD, Tang ZM. Protective effects of SH-compounds on ischemia reperfusion induced arrhythmias in the isolated rat heart. Yao Xue Xue Bao 1991; 26:91-95.

- 9. Qiuy, Galinanes M, Ferrari R, Cargnoni A, Ezrin A, Hearse DJ. PEG-SOD improves post-ischemic functional recovery and antioxidant status in bloodperfused rabbit hearts. Am J Physiol 1992; 263: H1243-H1249.
- 10. Murry CE, Jennings RB, Reimer KA. Preconditioning with ischemia: a delay of lethal cell injury in ischemic myocardium. Circulation 1986; 74: 1124-1136.
- 11. Li,Y, P.Whittaker, Kloner R. The transient nature of the effect of ischemic preconditioning on myocardial infarct size and ventricular arrhythmia. Am Heart J 1992; 123:346–353.
- 12. Liu Y, Downey JM. Ischemic preconditioning protects against infarction in rat heart. Am J Physiol 1992; 263: H1107-H1112.
- 13. Murry, CE, Richard VJ, Reimer KA, Jennings RB. Ischemic preconditioning slows energy metabolism and delays ultrastructural damage during a sustained ischemic episode. Circ Res 1990; 66:913–931.
- 14. Efstathios KI, Antigone L, Dimitrios TK. Ischemic preconditioning: Protection against myocardial necrosis and apoptosis. Vasc Health Risk Manag 2007; 3: 629–637.
- 15. Walker MJA, Curtis MJ, Hearse DJ, Campbell RWF, Janse MJ, Yellon DM, *et al.* The Lambeth Conventions: guidelines for the study of arrhythmias in ischaemia, infarction, and reperfusion. Cardiovasc Res 1988; 22: 441-455.
- 16. Takayuki M, Uno I, Munetaka F, Jun S, Hirofumi A, Kazu-ichi Y. Effects of ischemic and Sevoflurane-induced preconditioning on myocardial infarction and arrhythmias in rabbits *in vivo*. J Anesth Clin Res 2013; 4:361-366.
- 17. Matejikova J, Kucharska J, Pinterova M, Pancza D, Ravingerova T. Protection against ischemia-induced ventricular arrhythmias and myocardial dysfunction conferred by preconditioning in the rat heart: involvement of mitochondrial KatpChannels. Physiol Res 2009; 58:9-19.
- 18. Evrengul H, Seleci D, Tanriverdi H, Kaftan A. The antiarrhythmic effect and clinical consequences of ischemic preconditioning. Coron Artery Dis 2006 May; 17:283-288.
- 19. Derek J, Hausenloy M, Derek MY. Remote ischaemic preconditioning: underlying mechanisms and clinical application. Cardiovascular Research 2008; 79:377–386.
- 20. Przyklenk K, Bauer B, Ovize M, Kloner R, Whittaker P. Regional ischemic preconditioning protects remote virgin myocardium from subsequent sustained coronary occlusion. Circulation 1993; 87: 893–899.
- 21. Jose FC, Ricardo F-C, Adelino FLM. Myocardial remote ischemic preconditioning: From pathophysiology to clinical application. Rev Port Cardiol 2013; 32:893-904.
- 22. Das DK, Maulik A. Preconditioning potentiates redox signaling and converts death signal into survival signal. Arch Biochem Biophys 2003; 420:305-311.
- 23. Gross G, Fryer RM. Sarcolemmal versus mitochondrial ATP-sensitive K+ channels and myocardial preconditioning in dogs. Circ Res 1999; 9: 973-979.

- 24. Das B, Sarkar CH. Is the sarcolemmal or mitochondrial K_{ATP} channel activation important in the antiarrhythmic and cardioprotective effects during acute ischaemia/reperfusion in the intact anesthetized rabbit model. Life Sci 2005; 77: 1226-1248.
- 25. Vegh A, Parratt JR. The role of mito K_{ATP} channels in antiarrhythmic effects of ischaemic preconditioning in dogs. Br J Pharmacol 2002; 137: 1107-1115.
- 26. Gross GJ, Auchampach JA. Blockade of ATP-sensitive potassium channel prevents myocardial preconditioning in dogs. Circ Res 1992; 70: 223-233.
- 27. Grover GJ, D'alonzo AJ, Sleph PG, Dzwonczyk S, Hess T, Darbenzio RB. The cardioprotective and electrophysiological effects of cromakalim are attenuated by meclofenamate through a cyclooxygenase independent mechanism. J Pharmacol Exp 1994; 269: 536-540.
- 28. Tosaki A, Szerdahelyi P, Das DK. Reperfusion-induced arrhythmias and myocardial ion shifts: a pharmacologic interaction between pinacidil and cicletanine in isolated rat hearts. Basic Res Cardiol 1992; 87: 366-384.
- 29. Hamada K, Yamazaki J, Nagao T. Shortening of action potential duration is not prerequisite for cardiac protection by ischemic preconditioning or a K_{ATP} channel opener. J Mol Cell Cardiol 1998; 30: 1369-1379.
- 30. Ravingerova T, Lokebo JE, Sundset R, Ytrehus K. Preconditioning against contractile dysfunction in guinea pig papillary muscle depends on the opening of K_{ATP} -sensitive channels. Exp Clin Cardiol 1998; 3: 184-188.
- 31. Sato T, Sasaki N, Seharaseyon J, O'rourke B, Marban E. Selective pharmacological agents implicate mitochondrial but not sarcolemmal K_{ATP} channels in ischemic cardioprotection. Circulation 2000; 101: 2418-2423.
- 32. Munch-Ellingsen J, Lokebo JE, Bugge E, Jonassen AK, Ravingerova T, Ytrehus K: 5-HD abolishes ischemic preconditioning independently of monophasic action potential duration in the heart. Basic Res Cardiol 2000; 95: 228-234.
- 33. Garlid KD, Dos Santos P, Xie Z-J, Costa ADT, Paucek P. Mitochondrial potassium transport: the role of the mitochondrial ATP-sensitive K channels in cardiac function and cardioprotection. Biochim Biophys Acta 2003; 1606: 1-21.
- 34. Yue Y, Qin Q, Cohen MV, Downey JM, Critz SD. The relative order of mito K_{ATP} channels, free radicals and p38 MAPK in preconditioning's protective pathway in rat heart. Cardiovasc Res 2002; 55: 681-689.
- 35. Hausenloy DJ, Yellon DM. Preconditioning and postconditioning: united at reperfusion. Pharmacol Ther 2007; 116: 173–191.
- 36. Steen BK, Ole H, Rajesh KK, Jens E N-K, Michael RS, Andrew N. Remote preconditioning reduces ischemic injury in the explanted heart by a $K_{\rm ATP}$ channel-dependent mechanism. Am J Physiol Heart Circ Physiol 2005; 288: H1252–H1256.
- 37. Harlokesh N Y, Manjeet S, P.L. Sharma, Dhiraj M, Tapan B, Atinder P K. Possible Role of Cyclooxygenase-2 in Remote Aortic Preconditioning Induced Cardioprotection in Rat Heart. Pharmacologia 2012; 3: 1-8.

- 38. Lloris-Carsi JM, Cejalvo D, Toledo-Pereyra LH, Calvo MA, Suzuki S. Preconditioning: Effect upon lesion modulation in warm liver ischemia. Transplant Proc 1993; 25: 3303–3304.
- 39. Najafi M, Garjani A. The effect of L-carnitine on arrhythmias in the ischemic rat heart. Iran J Basic Med Sci 2005; 8: 38-44.
- 40. Najafi M, Shaseb E, Ghaffary Sa, Fakhrju A, Oskouei Eteraf T. Effects of chronic oral administration of natural honey on ischemia-reperfusion-induced arrhythmias in isolated rat heart. Iran J Basic Med Sci 2011; 14:75-81.
- 41. Shahid M, Tauseef M, Sharma K K, Fahim M. Brief femoral artery ischemia provides protection agains myocardial ischaemia–reperfusion injury in rats: the possible mechanisms. Exp Physiol 2008; 93: 954–968.
- 42. Przyklenk K, Whittaker P. Remote ischemic preconditioning current knowledge, unresolved questions, and future priorities. J Cardiovasc Pharmacol Ther 2011; 16: 255-259.
- 43. Clanahan MC, Nao B, Wolke L, Martin BJ, Mezt TE. Brief renal occlusion and reperfusion reduces myocardial infarct size in rabbits. FASEB J 1993; 7: A18.
- 44. Gho BC, Schoemaker RG, van den Doel MA, Duncker DJ, Verdouw PD. Myocardial protection by brief ischemia in noncardiac tissue. Circulation 1996; 94: 2193-2200.
- 45. Tang ZL, Dai W, Li YJ, Deng HW. Involvement of capsaicin-sensitive sensory nerves in early and delayed cardioprotection induced by a brief ischaemia of the small intestine. Naunyn Schmiedebergs Arch Pharmacol 1999; 359: 243-247.
- 46. Tatyana O, Michael A, Rodica K, Natalie A, Babeth R. Limb ischemia preconditions the heart against reperfusion tachyarrhythmia. Am J Physiol Heart Circ Physiol 1997; 273: H1707-H1712.
- 47. Heidi LL, Stephen ED. Partial Hind limb Occlusion Reduced the Susceptibility to Sustained Ventricular Tachycardia in Conscious Rats. J Cardiovasc Pharmacol Ther 2009; 14(3): 199–206.
- 48. Curtis MJ. Characterisation, utilisation and clinical relevance of isolated perfused heart models of ischemia-induced ventricular fibrillation. Cardiovasc Res 1998; 9: 194-215.
- 49. Bernier M, Ccurtis MJ, Hearse DJ. Ischemiainduced and reperfusion-induced arrhythmias: importance of heart rate. Am J Physiol 1989; 256: H21-H31.
- 50. Ravigerova T, Tibulova N, Slezak J, Curtis MJ. Brief intermediate and prolonged ischemia in the isolated crystalloid perfused rat heart: relationship between susceptibility to arrhythmias and degree of ultrastructural injury. J Mol Cell Cardiol 1995; 27: 1937-1951.
- 51. Botsford MW, Lukas A. Ishemic preconditioning and arrhythmogenesis in the rabbit heart: effects on epicardium vs. endocardium. J Mol Cell Cardiol 1998; 30: 1723-1735.
- 52. Cohen MV, Yang X-M, Liu GS, Heusch G, Downey JM. Acetylcholine, bradykinin, opioids, and henylephrine, but not adenosine, trigger preconditioning by generating free radicals and openingmitochondrial K_{ATP} channels. Circ Res 2001; 89: 273-278.
- 53. Zhang DX, Chen YF, Campbell WB, Zou AP, Gross GJ, Li PL. Characteristics and superoxide-induced



- activation of reconstituted myocardial ATP-sensitive potassium channels. Circ Res 2001; 89: 1177-1183.
- 54. Kuazaki T, Masahiro A, Hitoshi I, Tomoaki T, Mikio Yanase, Yukiko I, *et al.* Ischemic Preconditioning Protects Hepatocytes Via Reactive Oxygen Species Derived From Kupffer Cells in Rats. Gastroenterology 2004; 127: 1488–1496.
- 55. Morihira M, Hasebe N, Baljinnyam E, Sumitomo K, Matsusaka T, Izawa K, *et al.* Ischemic preconditioning enhances scavenging activity of reactive oxygen species and diminishes transmural difference of infarct size. Am J Physiol 2006; 290: H577-H583.
- 56. Maczewski M, Duda M, Pawlak W, Reresewiks A. Endothelial protection from reperfusion injury by ischemic preconditioning and diazoxide involves a SOD-like anti-O2-mechanism. J Physiol Pharmacol 2004; 55: 537-550.
- 57. Glantz L, Avramovich A, Trembovler V, Gurvits V, Kohen R, Eidelman LA, ShohaMI E. Ischemic preconditioning increases antioxidants in the brain and peripheral organ after cerebral ischemia. Exp Neurol 2005; 192: 117-124.
- 58. Honda HM, Korge P, Weiss JN. Mitochondria and ischemia/reperfusion injury. Ann NY Acad Sci 2005; 1047: 248-258.
- 59. Glanemann M, Vollmar B, Nussler AK, Schaefer T, Neuhaus P, Menger MD. Ischemic preconditioning protects from hepatic ischemia/reperfusion-injury by preservation of microcirculation and mitochondrial redox-state. J Hepatol 2003; 38: 59–66.
- 60. Fernandez L, Heredia N, Grande L, Gomez Ga, Rimola A, Marco A. Preconditioning Protects Liver and Lung Damage in Rat Liver Transplantation: Role of Xanthine/Xanthine Oxidase. Hepatology 2002; 36: 562-572.
- 61. Yue Y, Qin Q, Cohen MV, Downey JM, Critz SD. The

- relative order of mito K_{ATP} channels, free radicals and p38 MAPK in preconditioning's protective pathway in rat heart. Cardiovasc Res 2002; 55: 681-689.
- 62. Tang LD, Tang ZM. Protective effects of SH-compounds on ischemia reperfusion induced arrhythmias in the isolated rat heart. Yao Xue Xue Bao 1991; 26: 91-95.
- 63. Forbes RA, Steenbergen CH, Murphy E. Diazoxide-induced cardioprotection requires signaling through a redox-sensitive mechanism. Circ Res 2001; 88: 802-809. 64. Hu K, Duan D, Li GR, Nattel S. Protein kinase C activates ATP-sensitive K+current in human and rabbit ventricular myocytes. Circ Res 1996; 78: 492-498.
- 65. Sato T, O'Rourke B, Marban E: Modulation of mitochondrial ATP-dependent K+channels by protein kinase C. Circ Res 1998; 83: 110-114.
- 66. Murphy E. Primary and secondary signaling pathways in early preconditioning that converge on the mitochondria to produce cardioprotection. Circ Res 2004; 94: 7-16.
- 67. Pain T, Yang XM, Critz SD, Yue Y, Nakano A, Liu GS, *et al.* Opening of mitochondrial K_{ATP} channels triggers the preconditioned state by generating free radicals. Circ Res 2000; 87: 460-466.
- 68. Holmuhamedov EL, Wang L, Terzic A. ATP-sensitive K+channel openers prevent Ca++ overload in rat cardiac mitochondria. J Physiol Lond 1999; 519: 347-360.
- 69. Lim KH, Javadov SA, Das M, Clarke SJ, Suleiman MS, Halestrap AP. The effects of ischaemic preconditioning, diazoxide and 5-hydroxydecanoate on rat heart mitochondrial volume and respiration. J Physiol Lond 2002; 545: 961-974.
- 70. Shimizu S, Narita M, Tsujimito Y. Bcl-2 family proteins regulate the release of apoptogenic cytochrome c by the mitochondrial channel VDAC. Nature 1999; 399: 483-487.