

Troxeutin affects the male fertility in prepubertal type 1 diabetic male rats

Zohreh Zavvari Oskuye¹, Fariba Mirzaei Bavi², Gholam Reza Hamidian³, Keyvan Mehri¹, Afsaneh Qadiri¹, Mahdi Ahmadi⁴, Hajar Oghbaei¹, Amir Mansour Vatankhah², Rana Keyhanmanesh^{2*}

¹ Department of Physiology, Faculty of Medicine, Tabriz University of Medical Sciences, Tabriz, Iran

² Drug Applied Research Center, Tabriz University of Medical Sciences, Tabriz, Iran

³ Department of Basic Sciences, Faculty of Veterinary Medicine, University of Tabriz, Tabriz, Iran

⁴ Tuberculosis and Lung Diseases Research Center, Tabriz University of Medical Sciences, Tabriz, Iran

ARTICLE INFO

Article type:
Original article

Article history:
Received: Jun 12, 2018
Accepted: Oct 14, 2018

Keywords:
Diabetes
FSH
LH
Oxidative stress
Prepubertal
Troxeutin

ABSTRACT

Objective(s): Diabetes can gradually cause damage to the function and structure of male gonads. This survey was conducted to investigate the effect of troxeutin on hormonal changes, serum oxidative stress indices, and testicular function and structure in prepubertal diabetic rats.

Materials and Methods: Fifty prepubertal (6 weeks old) male Wistar rats were divided into five groups including Control, Troxeutin, Diabetic, Diabetic+Troxeutin, and Diabetic+Insulin. Type I diabetes was induced by 55 mg/kg of streptozotocin intraperitoneally. The groups were treated with 150 mg/kg/day troxeutin via oral gavage or 4-6 IU/day insulin via subcutaneous injection for 4 consecutive weeks. Blood sugar (BS) and serum levels of insulin, FSH, LH, testosterone, glutathione peroxidase (GPX), superoxide dismutase (SOD), malondialdehyde (MDA), and total antioxidant capacity (TAC) were analyzed. Testis and epididymis were removed for histopathologic study and analysis of sperm parameters.

Results: Troxeutin significantly reduced the BS in the diabetic group similar to insulin but could not affect insulin, FSH, or LH significantly. Troxeutin caused a significant increase in testosterone and GPX but had no significant effect on serum MDA, TAC, and SOD levels. In addition, troxeutin had a better effect than insulin on diabetes-induced testicular structural damage. Sperm analysis results also revealed that troxeutin and insulin could improve sperm number, motility, and viability in diabetic rats.

Conclusion: According to these results, it can be derived that administration of troxeutin is a suitable protective strategy for side effects of diabetes in testis of prepubertal diabetic male rats.

► Please cite this article as:

Zavvari Oskuye Z, Mirzaei Bavi F, Hamidian GHR, Mehri K, Qadiri A, Ahmadi M, Oghbaei H, Vatankhah AM, Keyhanmanesh R. The effect of troxeutin on male fertility in prepubertal type 1 diabetic male rats. Iran J Basic Med Sci 2019; 22:197-205. doi: 10.22038/ijbms.2018.32678.7814

Introduction

Diabetes mellitus is a great endocrine and metabolic problem nowadays (1, 2). Diabetes mellitus Type 1 results from severe insulin deficiency while diabetes mellitus type 2 is characterized by insulin resistance, which may be merged with relatively reduced insulin secretion (3, 4). The report of the International Diabetes Federation in 2015 showed that the global prevalence of diabetes was estimated to be 415 million in adults and predicted that this figure will reach over 600 million in 2035 (5).

In diabetes mellitus type 1, pancreatic beta cells are damaged by the immune system, therefore, patients must use exogenous insulin to control blood sugar and inhibit risk of developing long-term complications (6, 7). Good glycemic control can avoid its complications (8).

One of the most common complications of diabetes is sexual dysfunction (9). It has been proposed that the reproductive complications of diabetes mellitus are caused by at least two different mechanisms including endocrine disorders (10) and oxidative stress (11-13). Some of sexual dysfunctions in diabetic men are disorders in ejaculation, libido (14), erection (15),

testicular tissue structure (16), sperm quality (17, 18), and testosterone and gonadotropins secretion (19, 20).

Since chemical drugs have many side effects, herbal drugs today are considered for control of diabetes complications. Herbal nutrition and major pharmaceutical companies are currently doing research on natural materials to find new herbal subjects with the least side effects (21, 22). One of these herbal subjects is troxeutin also known as vitamin P₄. It is a tri-hydroxyethylated derivative of natural flavonoids and can be found in tea, coffee, cereal grains, and some fruits and vegetables (23). This substance can be easily absorbed by the gastrointestinal system (24) and has many biological and pharmacological activities such as anti-oxidative (25), anti-inflammatory (26), anti-fatigue (27), anti-thrombotic (28), and anti-hyperglycemic (29) properties. Previous experiments confirmed that troxeutin has protective effects on the kidneys (23), liver (30), brain (1), and vascular injuries (24); and chronic venous insufficiency (CVI) disease could be treated by this flavonoid (31). Moreover, troxeutin could prevent nickel-induced testicular toxicity in Wistar rats (32).

Although there are reports for the anti-hyperglycemic effects of troxeutin, we did not find any studies about

*Corresponding author: Rana Keyhanmanesh. Drug Applied Research Center, Tabriz University of Medical Sciences, Tabriz, Iran. Tel/Fax: +98-4133364664; Email: keyhanmanesh@tbzmed.ac.ir

the protective effects of troloxerutin on the reproductive system in diabetic cases. Hence, this survey was designed to investigate the effect of troloxerutin on testicular function and structure in type 1 diabetic male rats.

Materials and Methods

Animal and experimental design

Fifty prepubertal (6 weeks old, weighing 90–115 g) male Wistar rats were attained from animal center of Tabriz University of Medical Sciences and transported to Drug Applied Research Center. Animals were kept in standard laboratory conditions; 12 hr light/12 hr dark cycle, 20–22 °C, 45–55% moisture, and water and food *ad libitum*. A week after transportation and adaptation, the animals were randomly divided into 5 groups (n=10);

1. Control group (C).
2. Troloxerutin group (T) which received troloxerutin (Merck, Germany) 150 mg/kg/day via oral gavage for 4 weeks (24).
3. Diabetic group (DM) (1).
4. Diabetic group treated with troloxerutin 150 mg/kg/day via oral gavage for 4 weeks (DT) (33).
5. Diabetic group treated with neutral protamine Hagedorn (NPH) insulin 4–6 IU/day subcutaneously (DI) (34).

For induction of diabetes, 55 mg/kg of liquefied streptozotocin (Sigma-Aldrich, Germany) in 10 mM sodium citrate (pH= 4.5) was injected intraperitoneally in DM, DI, and DT groups (35). Three days after streptozotocin injection, a blood sample of the tail vein was obtained and blood sugar (BS) was measured by means of a digital glucometer (Norditalia Elettromedicali S.r.l., Italy). If BS was more than 250 mg/dL, that animal was considered diabetic. The Ethics Committee of Tabriz University of Medical Sciences has supported all experimental procedures (No: IR.TBZMED.REC.1395.584).

Sampling

On the last day of experiments, animals were deeply anesthetized with intraperitoneal injection of a combination of ketamine and xylazine (80 and 12 mg/kg, respectively). Five milliliters of blood samples, taken from the inferior vena cava, were centrifuged at 3000 rpm for 10 min at room temperature. Then, serum aliquots were isolated and were kept at -80 °C for later hormonal and oxidative stress analysis. Finally, animals were sacrificed by decapitation and the left testis was removed. Length (longitudinal radius), width (transversal radius), and height (perpendicular to the transversal radius) of each testis was measured by caliper and after macroscopic evaluation tissue samples were assessed after fixation in 10% buffered neutral formalin.

Hormonal analysis of serum

The serum levels of insulin (Shanghai crystal Day Biotech Co, LTD, China, for insulin with 0.05 ng/ml sensitivity), FSH (Bioassay Technology Laboratory, China, with 0.12 mIU/ml sensitivity), LH (Bioassay Technology Laboratory, China, with 0.051 mIU/ml sensitivity) and testosterone (Diametra Co, Italy, with

0.07 ng/ml sensitivity) were measured using enzyme-linked immuno-absorbent assay according to the manufacturer's instructions (8).

Oxidative stress measurement

Serum level of glutathione peroxidase (GPX), superoxide dismutase (SOD), malondialdehyde (MDA), and total antioxidant capacity (TAC) were analyzed. The activity of GPX was measured using Randox kit, United Kingdom. SOD was evaluated using a spectrophotometric method on the basis of the inhibition of a superoxide-induced reduced nicotinamide adenine dinucleotide (NADH) oxidation and MDA was measured using the thiobarbituric acid (TBARS) and colorimetric method (33). TAC was measured using the Randox Kit according to the protocols of the manufacturer.

Sperm parameters

Immediately after the last intervention, animals were anesthetized and the testes, epididymis and vas deferens were removed for determination of the spermatozoid parameters. To calculate the epididymal sperm storage, epididymis were isolated from the testes. Accordingly, epididymis was cut from the tail, and the sperm content of epididymis was aspirated into a pre-weighed pipette. Then the pipette was re-weighed to measure the weight of the aspirated fluid (36).

A 1 ml sample of the suspension containing sperm extracted from the epididymis tail was diluted with 20 ml of Ham's F10 solution and the sperm count and motility were determined at $\times 400$ magnification. Subsequently, the total mean of motile sperms in ten fields was expressed as a percentage of motility according to the World Health Organization (WHO) guidelines (37).

Determination of the spermatozoa viability percentage was also performed according to the WHO guidelines, using eosin (1%) and nigrosin (10%) staining. Briefly, 1 ml sperm suspension was mixed with 2 ml of eosin. After incubation at 37 °C for 30 sec, an equal volume of nigrosin was added to this suspension. Then the percentage of viable sperms was calculated in different groups under a light microscope (3).

Moreover, the total number of sperms was counted in a hemocytometer. Concisely, the number of sperms in diluted sperm solution was counted in five large squares of a hemocytometer. Then the sperm count per milliliter was calculated (38, 39).

For evaluation of sperm motility, at least 200 spermatozoa in 100 μ l sperm suspension were evaluated under a light microscope and scored from A to D according to the WHO manual including (40, 41): grade A as fast progressive; grade B as slow progressive; grade C as non-progressive, and grade D as immotile sperm.

Histological analysis

Fixed testicular tissue samples were dehydrated in an ascending graded series of ethylic alcohol, cleared in xylol, and impregnated in paraffin. The testis was cut by a rotary microtome to 5 μ m thin sections and was stained by hematoxylin and eosin (H&E) according to a previously described protocol. At least 10 sections were checked for each animal with a light microscope. Testis

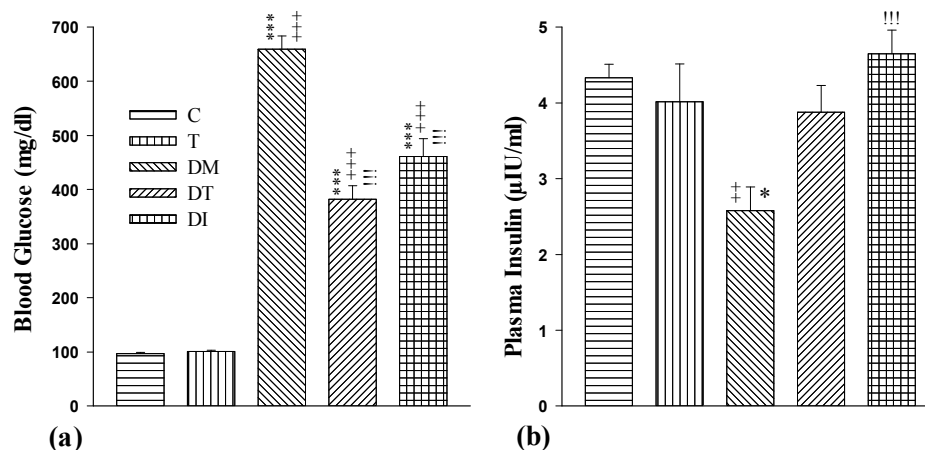


Figure 1. The effect of troloxerutin on blood glucose (a) and plasma insulin (b) in prepubertal type 1 diabetic male rats (n=10). Data are expressed as mean±SEM. Data were analyzed by using one way ANOVA followed by Tukey's *post hoc* test. Statistical differences between control and different groups: +++; $P<0.001$, ++; $P<0.01$, Statistical differences between troloxerutin and different groups: ***; $P<0.001$, *; $P<0.05$, Statistical differences between diabetic and different groups: !!!; $P<0.001$. Control (C), troloxerutin (T), diabetic (DM), diabetes+troloxerutin (DT), and diabetes+insulin (DI)

and seminiferous tubules were evaluated histologically (3).

Germinal epithelium organization and quality of 100 seminiferous tubules per animal were analyzed by Johnsen's score from 1 to 10 according to the following criteria (41):

Score 10: full spermatogenesis with regular germinal epithelium; Score 9: disorganized germinal epithelium with many spermatozoa; Score 8: existence of only a little spermatozoa; Score 7: no spermatozoa but many spermatids existed; Score 6: no spermatozoa and only a few spermatids were observed; Score 5: no spermatozoa, no spermatid but many spermatocytes existed; Score 4: only a few spermatocytes were observed; Score 3: only spermatogonia were present; Score 2: Sertoli cells were observed but no germ cells were present; Score 1: seminiferous tubule without any cells. All evaluations were done by an expert technician who was blinded to our study.

Statistical analysis

The results were expressed as mean ± SEM (standard error of the mean). Statistical analysis was performed by SPSS software version 22 (IBM company, SPSS Inc., 2010). One-way analysis of variance (ANOVA) followed by *post hoc* Tukey's test was used to assess the statistical significance of data between different groups. It was considered significant if $P<0.05$.

Results

The levels of blood glucose and serum insulin

The blood glucose levels in DM, DT, and DI groups increased significantly compared to the control and troloxerutin groups ($P<0.001$). Both insulin and troloxerutin treatment significantly diminished the blood glucose levels in comparison with the diabetic group ($P<0.001$) (Figure 1a).

Injection of streptozotocin in the DM group reduced insulin level compared to C and T groups ($P<0.01$ to $P<0.05$). There was an increased insulin level in the DI group compared to diabetic rats ($P<0.001$), however, administration of troloxerutin nonsignificantly increased the insulin level compared to the DM group (Figure 1b).

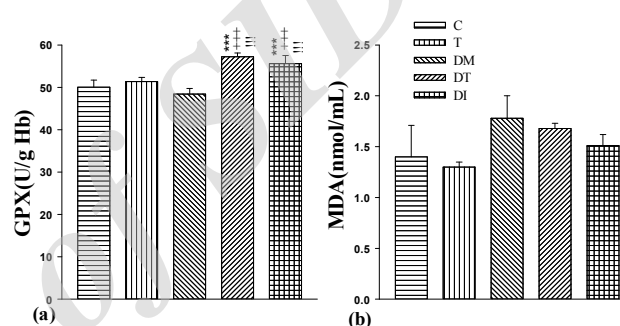


Figure 2. The effect of troloxerutin on serum glutathione peroxidase (GPX, a) and malondialdehyde (MDA, b) in prepubertal type 1 diabetic male rats (n=10) for 4 weeks. Data are expressed as mean±SEM. Statistical differences between control and different groups: +++; $P<0.001$, Statistical differences between troloxerutin and different groups: ***; $P<0.001$, Statistical differences between diabetic and different groups: !!!; $P<0.001$. Control (C), troloxerutin (T), diabetic (DM), diabetes+troloxerutin (DT), and diabetes+insulin (DI)

The levels of serum glutathione peroxidase (GPX) and malondialdehyde (MDA)

Induction of diabetes did not show any effect on the GPX level; however, in DI and DT groups, administration of insulin and troloxerutin increased GPX level significantly ($P<0.001$). It must be mentioned that insulin and troloxerutin did not have a significant difference in this regard (Figure 2a).

Although induction of diabetes increased the MDA level in comparison to the controls, this increment was not statistically significant. Treatment with insulin and troloxerutin in DI and DT groups nonsignificantly decreased the level of MDA. There was no significant difference between DI and DT groups (Figure 2b).

The serum levels of total antioxidant capacity (TAC) and superoxide dismutase (SOD)

Induction of diabetes and administration of troloxerutin and insulin did not change the TAC level significantly compared to the C group (Figure 3a).

Although induction of diabetes decreased the SOD level in comparison to the controls, it was not statistically significant. Treatment with insulin and troloxerutin in DI and DT groups nonsignificantly increased the level of

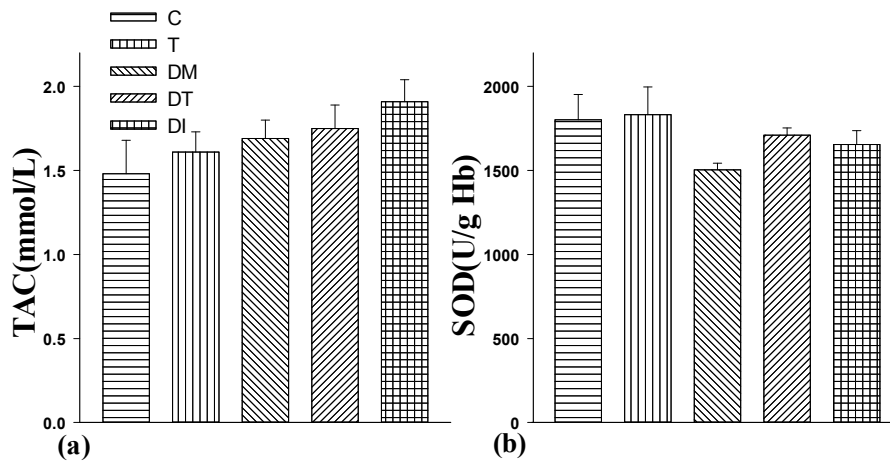


Figure 3. The effect of troloxerutin on total antioxidant capacity (TAC, a) and superoxide dismutase (SOD, b) in prepubertal type 1 diabetic male rats (n=10) for 4 weeks. Control (C), troloxerutin (T), diabetic (DM), diabetes+troloxerutin (DT), and diabetes+insulin (DI)

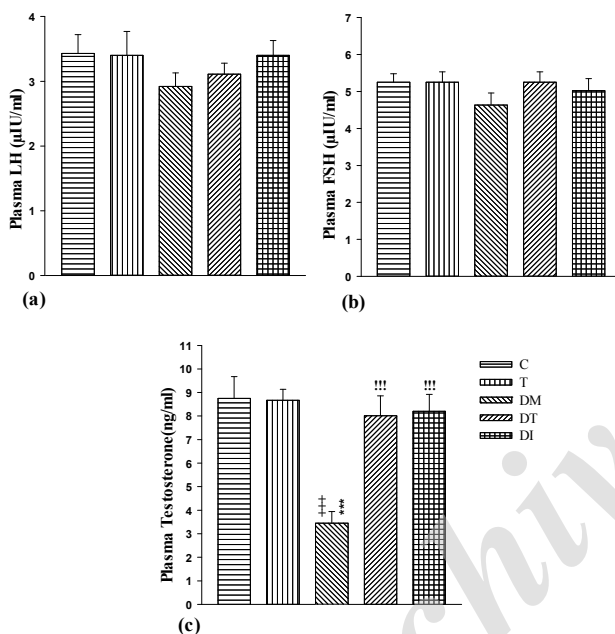


Figure 4. The effect of troloxerutin on LH (a), FSH (b), and testosterone (c) in prepubertal type 1 diabetic male rats (n=10) for 4 weeks. Data are expressed as mean±SEM. Statistical differences between control and different groups: +++; $P<0.001$, Statistical differences between troloxerutin and different groups: ***; $P<0.001$, Statistical differences between diabetic and different groups: !!!; $P<0.001$. Control (C), troloxerutin (T), diabetic (DM), diabetes+troloxerutin (DT), and diabetes+insulin (DI)

SOD in comparison to the DM group. There was no significant difference between treated diabetic groups (Figure 3b).

The serum levels of LH, FSH, and testosterone

The serum testosterone level decreased significantly after diabetes induction compared to C and T groups ($P<0.001$), whereas LH and FSH serum levels decreased nonsignificantly. Treatment with insulin and troloxerutin in DI and DT groups increased testosterone level significantly in comparison to diabetic rats ($P<0.001$), although the increments in the serum levels of LH and FSH in these groups were not statistically significant. There were not any significant differences between DT and DI groups (Figures 4a, b, and c).

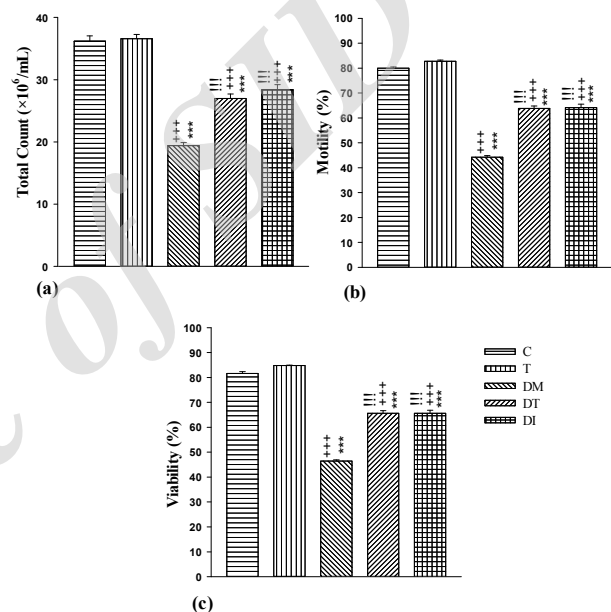


Figure 5. The effect of troloxerutin on total count (a), motility (b), and viability (c) of sperms in prepubertal type 1 diabetic male rats (n=10) for 4 weeks. Data are expressed as mean±SEM. Statistical differences between control and different groups: +++; $P<0.001$, Statistical differences between troloxerutin and different groups: ***; $P<0.001$, Statistical differences between diabetic and different groups: !!!; $P<0.001$. Control (C), troloxerutin (T), diabetic (DM), diabetes+troloxerutin (DT), and diabetes+insulin (DI)

Sperm parameters

The induction of diabetes reduced the total number, motility, and viability of sperms in comparison to C and T groups ($P<0.001$). Troloxerutin could not affect these parameters in control rats. Administration of troloxerutin and insulin significantly improved all of these parameters compared to the DM group ($P<0.001$). There were no significant differences between DT and DI groups (Figures 5a, b, and c).

Sperm motility grade

Induction of diabetes significantly decreased the percentages of fast progressive, slow progressive and non-progressive sperms ($P<0.001$ to $P<0.01$) but increased the percentage of immotile sperms ($P<0.001$) compared to control and troloxerutin groups. Treatment

Table 1. The effect of troxerutin on sperm motility grade in prepubertal type 1 diabetic male rats (n=10) for 4 weeks

| Motility Grades | Scores in groups | | | | |
|-------------------------------|------------------|-----------|----------------------|--------------------------|--------------------------|
| | % (Mean±SEM) | | | | |
| | Control | T | DM | DT | DI |
| Grade A (fast progressive) | 48.6±0.68 | 49.6±0.24 | 20.4±0.93 +++ *** | 30.6±0.87 +++ *** !!! | 29.4±0.60 +++ *** !!! |
| Grade B (slow progressive) | 22.0±0.55 | 25.0±0.63 | 12.2±0.58 +++ *** | 28.0±0.89 +++ *** !!! | 25.6±1.07 +++ *** !!! |
| Grade C (non-progressive) | 8.6±0.24 | 8.2±0.37 | 5.8±0.37 ++ ** | 5.4±0.24 ++ ** | 8.2±0.20 !! ## |
| Grade D (immotile) | 20.8±0.86 | 17.2±0.58 | 61.6±1.32 +++ *** | 36.0±1.00 +++ *** !!! | 36.8±1.24 +++ *** !! |

Data are expressed as mean±SEM. Statistical differences between control and different groups: +++; $P<0.001$, ++; $P<0.01$, Statistical differences between troxerutin and different groups: ***, $P<0.001$, **, $P<0.01$, Statistical differences between diabetic and different groups: !!!; $P<0.001$, !!; $P<0.01$, Statistical differences between DT and DI groups: ##; $P<0.01$. Control (C), troxerutin (T), diabetic (DM), diabetes+troxerutin (DT), and diabetes+insulin (DI)

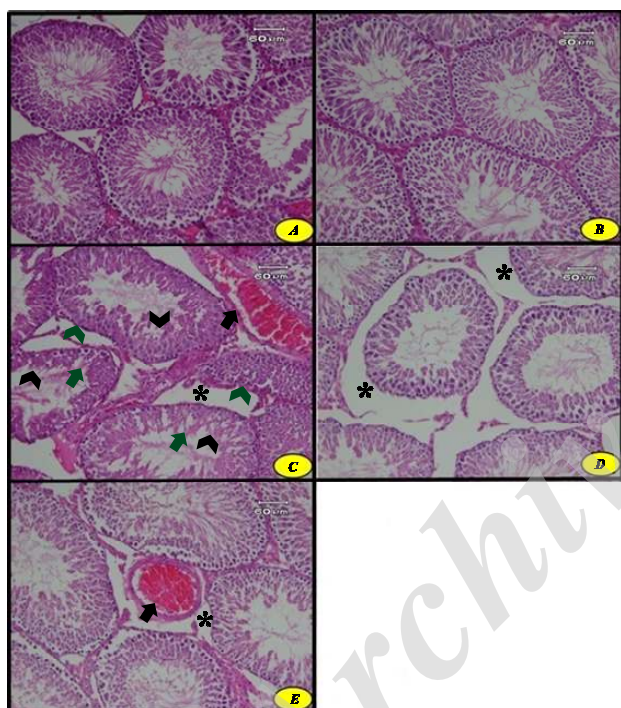


Figure 6. The effect of troxerutin on histological structure of testis in different groups (H&E, ×200) in prepubertal type 1 diabetic male rats (n=10) for 4 weeks. Control (A), troxerutin (B), diabetic (C), diabetic and troxerutin (D), and diabetic and insulin (E). An increase in interstitial space (*), vascular congestion (black arrow), presence of many vacuoles in the germinal epithelium (green arrow), destruction of germinal epithelium of seminiferous tubule (black arrowhead), shrinkage of tubule wall (green arrowhead) are observed

with troxerutin significantly increased the percentages of fast progressive and slow progressive sperms but decreased the percentage of immotile sperms ($P<0.001$). Moreover, in the DI group, insulin administration increased the percentages of fast progressive, slow progressive, and non-progressive sperms ($P<0.01$ to $P<0.001$) and decreased the percentage of immotile sperm ($P<0.001$) compared to the DM group. The percentage of non-progressive sperms of DT and DI groups was significantly different ($P<0.01$, Table 1).

Histological analysis of testis

Histological studies of testis tissue demonstrated

that the structure of the testis and seminiferous tubule are completely normal in the control and troxerutin-treated groups (Figures 6A and B). Microscopical analysis revealed that diabetes induction resulted in structural disturbance of the animals' testis including increased interstitial space and vascular congestion and destruction of germinal epithelium of seminiferous tubule. Existence of many vacuoles in germinal epithelium caused rupture in the cells integrity of this tissue. Irregular shape and shrinkage of seminiferous tubules basement membrane and disturbance of cellular arrangement and organization could be observed in diabetic rats (Figure 6C).

Histological studies of the testis in the diabetic group treated by troxerutin showed that the tissue structure was improved relatively compared to the diabetic group. The germinal epithelium of seminiferous tubule had proper structure and was similar to the control group. Increased interstitial space could be still observed in this group but vascular congestion was completely resolved (Figure 6D).

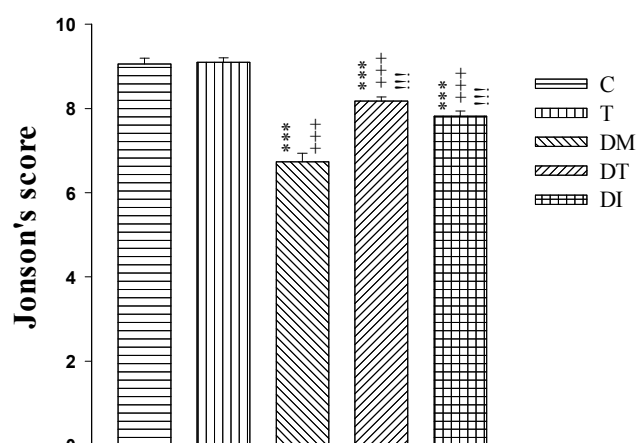


Figure 7. The effect of troxerutin on Jonhson's score for histopathological evaluation of testis tissues in prepubertal type 1 diabetic male rats (n=5) for 4 weeks. Data are expressed as mean±SEM. Statistical differences between control and different groups: +++; $P<0.001$, Statistical differences between troxerutin and different groups: ***, $P<0.001$, Statistical differences between diabetic and different groups: !!!; $P<0.001$. Control (C), troxerutin (T), diabetic (DM), diabetes+troxerutin (DT), and diabetes+insulin (DI)

Microscopic studies also revealed that treatment by insulin in the diabetic group resulted in evident structural improvement but increased volume in interstitial space could be still observed. Vascular congestion was less than in the diabetic rats but it was not totally eliminated. The effect of insulin administration on the improvement of structural changes of testis tissue was less compared with the group treated by troloxerutin for 4 weeks (Figure 6E).

Johnsen's score analysis illustrated a significant decrease in all diabetic rats (D, DT, and DI groups) compared to C and T groups ($P < 0.001$). Treatment with troloxerutin and insulin in DT and DI groups induced a significant increment compared to the DM group ($P < 0.001$) (Figure 7).

Discussion

Our results indicated that treatment of diabetic animals by insulin and troloxerutin resulted in significant increase of GPX, testosterone, total number, viability, and motility of sperm and Johnsen's score, although it reduced the blood sugar. The previous studies have shown that high blood sugar levels can affect fertility (42, 43). The results of a study have proven that hyperglycemia increased the testicular inflammatory cytokines (44). Other studies also suggested that hyperglycemia caused excessive production of ROS (45, 46) followed by the destruction of the testicular membrane (47).

Nowadays various drugs are used to treat diabetes, which may have a negative effect on other organs in addition to lowering blood glucose levels. One study reported that the use of sulfonylureas has led to apoptosis in beta cells and the failure of long-term treatment (48). Researchers also found that metformin and glibenclamide caused a significant reduction in the number and motility of the sperm and testicular damage, due to elevated lipid peroxidation and decreased antioxidant status in the testes (49).

Because of these side effects, scientists have tried to use alternative herbal drugs for controlling complications of diabetes. One of these medicines is troloxerutin. Regarding the positive effects of insulin on fertility, we decided to compare the drug with insulin. Insulin stimulates various functions of Sertoli cells, such as transferrin secretion, DNA and protein synthesis, glycine metabolism, lactate production (50, 51), and differentiation of spermatogonia through insulin-like growth factor receptor (IGF-1) (52).

In a study in 2014, increased sensitivity to insulin in the presence of troloxerutin has been reported as a major finding (53). Another study has also shown that routine supplementation effectively relieves symptoms of metabolic syndrome. In addition, they showed that the administration of troloxerutin in high-cholesterol-fed type 2 diabetic rats reduced the blood glucose levels, consistent with our results (54).

Kawamura *et al.* reported that SOD glycosylation percentage was significantly elevated in diabetic people compared with controls. The activity of glycosylated SOD is less than natural SOD (55). A study showed that MDA level of seminal plasma in diabetic men with normal sperm is more than that of non-diabetic men. Also, it has been shown that diabetic

men have lower levels of TAC compared to non-diabetic men (56). However, the current study showed that administration of troloxerutin (150 mg/kg) in immature diabetic rats had no significant effect on SOD, MDA, and TAC levels of serum in comparison to the diabetic group, but led to increment in serum level of GPX in comparison to the diabetic group. Previous study has also revealed that administration of troloxerutin to diabetic rats will not have a significant impact on the increase of SOD in comparison to those which had not received troloxerutin, although the serum level of GPX significantly increased (24), which is consonant with the results of our study. However, the current results did not coincide with the findings of Fan *et al* who investigated the effect of troloxerutin on D-galactose-induced renal injury in mice. These results indicated that it could increase the activity of antioxidant enzymes and reduce the lipid peroxidation products (23). The reasons for such differences can be attributed to the duration and severity of diabetes, method, and dosage of drug administration and method of diabetes induction.

Ballester *et al.* observed that induction of type 1 diabetes by streptozotocin for 3 months disturbed the function of Leydig cells and decreased the serum level of testosterone. This could be due to lack of stimulation effect of insulin on these cells. They also showed that the serum level of FSH and LH would also decrease in such conditions (57). In our study, the level of testosterone decreased significantly, although the levels of LH and FSH were decreased nonsignificantly. It seems that the duration of the experimental period of our study (4 weeks) can be the reason for these results, as the work of Ballester *et al.* (57) was conducted for 3 months. On the other hand, the age of rats could also make a difference. They worked on adult rats whereas our study was performed on prepubertal rats.

Previous study has revealed that diabetes can cause severe abnormalities in sperm by increasing the oxidative stress in testis and epididymis tissues (58). It has been shown that sperm cells of mammals contain high levels of lipids with high unsaturated fatty acids. On the other hand, spermatozoa use lipids as the main material for the peroxidation process. This can make the testis, epididymis, and released sperms in the seminiferous tubule a suitable site for production of free radicals as the result of lipid peroxidation during diabetes. The high rate of cell proliferation in germinal epithelium of the seminiferous tubule and reduction of anti-oxidative defense during diabetes can even intensify this issue (59). It has been also shown that hyperglycemia can increase the production of free radicals by increased glycolysis, activation of the sorbitol pathway in the cell, glucose self-oxidation, and proteins non-enzymatic glycation (45, 46), which is compatible with our findings in this investigation.

The results of qualitative and quantitative analysis of sperms in our study revealed that induction of diabetes by streptozotocin not only can affect the viability and the total number of sperms but also it can reduce the quality of sperm motility. Previous studies also suggested that diabetes and its consequent hyperglycemia can cause a reduction in quality and quantity parameters of sperm (3), disturb the spermatogenesis process (60), and also

decrease the sperm motility (57) by decreasing the production rate of testosterone.

Troxerutin has numerous biological protective effects against oxidative, fibrinolytic, inflammatory, γ -radiation, and diabetic damage and cancer (28, 54). It acts by affecting reactive oxygen species (ROS) and enzyme activities, probably indirectly by affecting the antioxidants acting on enzyme activities (61). The present study showed that troxerutin administration, similar to insulin, could relatively improve the diabetes-induced decrease in quantity and quality parameters of sperms. It seems that troxerutin can decrease the oxidative stress and blood sugar and relatively inhibit the side effects of diabetes on quality and quantity indices of sperm.

The results of this survey showed that induction of diabetes caused severe damage to the testicular tissue structure. Microscopical study showed that diabetes induced vascular hyperemia in the interstitial tissue of testis and increased the interstitial space of the seminiferous tubules due to interstitial edema. Kolahian *et al.* also reported these events in their study (3). The outcomes of our study displayed that induction of diabetes caused severe degeneration of spermatogenic cells as a result of structural changes in the germinal epithelium of seminiferous tubules. The disappearance of spermatogenic cells leads to the observation of many vacuoles in the germinal epithelium. In another study these reports were also observed (62). The results of the sperm analysis indicated that diabetes reduced testicular function and normal sperm production. The results of present investigation also showed that insulin or troxerutin therapy can importantly inhibit structural changes of the testis, surprisingly the effect of troxerutin was better than insulin and decreased the vascular congestion in testis, which was also consonant with the results of sperm analysis. So possible mechanisms that may be proposed for these effects of troxerutin include anti-inflammatory drug effect (28, 54), reduction of blood glucose (54, 63), improved insulin sensitivity (53), and indirect effect on antioxidants (61) because oxidative stress disrupts fertility (47).

Conclusion

According to these results, it can be concluded that administration of troxerutin is a suitable protective strategy for side effects of diabetes in testis of prepubertal diabetic male rats.

Acknowledgment

This report is based on a database from the thesis entitled "Effect of Troxerutin on blood levels of stress oxidative, testosterone, LH, FSH, insulin, and histological and stereological changes in type 1 diabetic adult and prepubertal male rat" registered in Drug Applied Research Center of Tabriz University of Medical Sciences, Tabriz, Iran. Moreover, the authors appreciate Professor S. Babri for gifting troxerutin.

Conflicts of Interest

The authors declare that they have no conflicts of interest to disclose.

References

- Baluchnejadmojarad T, Jamali-Raeufy N, Zabihnejad S, Rabiee N, Roghani M. Troxerutin exerts neuroprotection in 6-hydroxydopamine lesion rat model of Parkinson's disease: Possible involvement of PI3K/ER β signaling. *Eur J Pharmacol* 2017; 801:72-78.
- Ding GL, Liu Y, Liu ME, Pan JX, Guo MX, Sheng JZ, Huang HF. The effects of diabetes on male fertility and epigenetic regulation during spermatogenesis. *Asian J Androl* 2015; 17:948-53.
- Kolahian S, Sadri H, Larijani A, Hamidian G, Davasaz A. Supplementation of diabetic rats with leucine, zinc, and chromium: effects on function and histological structure of testes. *Int J Vitam Nutr Res* 2015; 85:311-21.
- Butler AE, Janson J, Bonner-Weir S, Ritzel R, Rizza RA, Butler PC. β -Cell deficit and increased β -cell apoptosis in humans with type 2 diabetes. *Diabetes* 2003; 52:102-110.
- Thomas MC, Cooper ME, Zimmet P. Changing epidemiology of type 2 diabetes mellitus and associated chronic kidney disease. *Nat Rev Nephrol* 2016; 12:73-81.
- Ashcroft FM, Rorsman P. Diabetes mellitus and the β cell: the last ten years. *Cell* 2012; 148:1160-1171.
- Regnell SE, Lernmark A. Early prediction of autoimmune (type 1) diabetes. *Diabetologia* 2017; 60:1370-1381.
- Zurita-Cruz JN, Nishimura-Meguro E, Villasís-Keever MA, Hernández-Méndez ME, Garrido-Magaña E, Rivera-Hernández AJ. Influence of the informal primary caretaker on glycemic control among prepubertal pediatric patients with type 1 diabetes mellitus. *J Pediatr (Rio J)* 2017; 93:136-141.
- Sexton WJ, Jarow JP. Effect of diabetes mellitus upon male reproductive function. *Urology* 1997; 49:508-513.
- Ali ST, Shaikh RN, Ashfaqsiddiqi N, Siddiqi PQ. Serum and urinary levels of pituitary-gonadal hormones in insulin-dependent and non-insulin-dependent diabetic males with and without neuropathy. *Arch Androl* 1993; 30:117-123.
- Ramvalho-Santos J, Amaral S, Oliveira PJ. Diabetes and the impairment of reproductive function: possible role of mitochondria and reactive oxygen species. *Curr Diabetes Rev* 2008; 4:46-54.
- Giacco F, Brownlee M. Oxidative stress and diabetic complications. *Circ Res* 2010; 107:1058-1070.
- Tiwari BK, Pandey KB, Abidi AB, Rizvi SI. Markers of oxidative stress during diabetes mellitus. *J Biomark* 2013; 2013:1-8.
- Watcho P, Mbiakop UC, Jeugo HG, Wankeu M, Nguenefack TB, Carro-Juarez M, Kamanyi A. Delay of ejaculation induced by Bersama engleriana in nicotine/streptozotocin-induced type 2 diabetic rats. *Asian Pac J Trop Med* 2014; 7:S603-S609.
- Ryan JG, Gajraj J. Erectile dysfunction and its association with metabolic syndrome and endothelial function among patients with type 2 diabetes mellitus. *J Diabetes Complications* 2012; 26:141-147.
- Abd El-Twab SM, Mohamed HM, Mahmoud AM. Taurine and pioglitazone attenuate diabetes-induced testicular damage by abrogation of oxidative stress and up-regulation of the pituitary-gonadal axis. *Can J Physiol Pharmacol* 2016; 94:651-661.
- Pomjunya A, Ratthanophart J, Fungfuang W. Effects of Vernonia cinerea on reproductive performance in streptozotocin-induced diabetic rats. *J Vet Med Sci* 2017; 79:572-578.
- Wankeu-Nya M, Florea A, Bălci S, Watcho P, Matei H, Kamanyi A. Dracaena arborea alleviates ultra-structural spermatogenic alterations in streptozotocin-induced diabetic rats. *BMC Complement Altern Med* 2013; 13:71-79.
- De A, Singh MF, Singh V, Ram V, Bisht S. Treatment effect of l-Norvaline on the sexual performance of male rats with streptozotocin induced diabetes. *Eur J Pharmacol* 2016; 771:247-254.
- Rovira-Llopis S, Bañuls C, de Marañón AM, Diaz-Morales

- N, Jover A, Garzon S, Rocha M, Victor VM, Hernandez-Mijares A. Low testosterone levels are related to oxidative stress, mitochondrial dysfunction and altered subclinical atherosclerotic markers in type 2 diabetic male patients. *Free Radic Biol Med* 2017; 108:155-162.
21. Patel DK, Kumar R, Laloo D, Hemalatha S. Diabetes mellitus: an overview on its pharmacological aspects and reported medicinal plants having antidiabetic activity. *Asian Pac J Trop Biomed* 2012; 2:411-420.
22. Goyal M. Traditional plants used for the treatment of diabetes mellitus in Sursagar constituency, Jodhpur, Rajasthan—An ethnomedicinal survey. *J Ethnopharmacol* 2015; 174:364-368.
23. Fan SH, Zhang ZF, Zheng YL, Lu J, Wu DM, Shan Q, Hu B, Wang YY. Troloxerutin protects the mouse kidney from d-galactose-caused injury through anti-inflammation and anti-oxidation. *Int Immunopharmacol* 2009; 9:91-96.
24. Badalzadeh R, Layeghzadeh N, Alihemmati A, Mohammadi M. Beneficial effect of troloxerutin on diabetes-induced vascular damages in rat aorta: histopathological alterations and antioxidant mechanism. *Int J Endocrinol Metab* 2015; 13:e25969.
25. Panat NA, Maurya DK, Ghaskadbi SS, Sandur SK. Troloxerutin, a plant flavonoid, protects cells against oxidative stress-induced cell death through radical scavenging mechanism. *Food Chem* 2016; 194:32-45.
26. Lu J, Wu DM, Zheng YL, Hu B, Cheng W, Zhang ZF, Li MQ. Troloxerutin counteracts domoic acid-induced memory deficits in mice by inhibiting CCAAT/enhancer binding protein β -mediated inflammatory response and oxidative stress. *J Immunol* 2013; 190:3466-3479.
27. Zamanian M, Hajizadeh MR, Esmaeili Nadimi A, Shamsizadeh A, Allahtavakoli M. Antifatigue effects of troloxerutin on exercise endurance capacity, oxidative stress and matrix metalloproteinase-9 levels in trained male rats. *Fund Clin Pharmacol* 2017; 31:447-455.
28. Liu C-M, Ma J-Q, Lou Y. Chronic administration of troloxerutin protects mouse kidney against D-galactose-induced oxidative DNA damage. *Food Chem Toxicol* 2010; 48:2809-2817.
29. Yu Y, Zheng G. Troloxerutin protects against diabetic cardiomyopathy through NF- κ B/AKT/IRS1 in a rat model of type 2 diabetes. *Mol Med Rep* 2017; 15:3473-3478.
30. Zhang ZF, Fan SH, Zheng YL, Lu J, Wu DM, Shan Q, Hu B. Troloxerutin protects the mouse liver against oxidative stress-mediated injury induced by D-galactose. *J Agric Food Chem* 2009; 57:7731-7736.
31. Gohel MS, Davies AH. Pharmacological agents in the treatment of venous disease: an update of the available evidence. *Curr Vasc Pharmacol* 2009; 7:303-308.
32. Elangovan P, Jalaludeen AM, Ramakrishnan R, Pari L. Protective effect of troloxerutin on nickel-induced testicular toxicity in wistar rats. *J Environ Pathol Toxicol Oncol* 2016; 35:133-46.
33. Kaya H, Sezik M, Ozkaya O, Dittrich R, Siebzehnrbil E, Wildt L. Lipid peroxidation at various estradiol concentrations in human circulation during ovarian stimulation with exogenous gonadotropins. *Horm Metab Res* 2004; 36:693-695.
34. Choi WS, Kwon OS, Cho SY, Paick JS, Kim SW. Effect of chronic administration of PDE5 combined with glycemic control on erectile function in streptozotocin-induced diabetic Rats. *J Sex Med* 2015; 12:600-610.
35. Alipour MR, Khamaneh AM, Yousefzadeh N, Mohammadnejad D, Soufi FG. Upregulation of microRNA-146a was not accompanied by downregulation of pro-inflammatory markers in diabetic kidney. *Mol Biol Rep* 2013; 40:6477-6483.
36. Amaral S, Moreno AJ, Santos MS, Seiça R, Ramalho-Santos J. Effects of hyperglycemia on sperm and testicular cells of Goto-Kakizaki and streptozotocin-treated rat models for diabetes. *Theriogenology* 2006; 66:2056-2067.
37. Oger P, Yazbeck C, Gervais A, Dorphin B, Gout C, Jacquesson L, Ayel JP, Kahn V, Rougier N. Adverse effects of hepatitis B virus on sperm motility and fertilization ability during IVF. *Reprod Biomed Online* 2011; 23:207-212.
38. Keegan BR, Barton S, Sanchez X, Berkeley AS, Krey LC, Grifo J. Isolated teratozoospermia does not affect in vitro fertilization outcome and is not an indication for intracytoplasmic sperm injection. *Fertil Steril* 2007; 88:1583-1588.
39. Bahmanzadeh M, Abolhassani F, Amidi F, Ejtemaiemehr Sh, Salehi M, Abbasi M. The effects of nitric oxide synthase inhibitor (L-NAME) on epididymal sperm count, motility, and morphology in varicocele rat. *Daru* 2008; 16:23-28.
40. Organization WH, WHO laboratory manual for the examination and processing of human semen 2010.
41. Johnsen SG. Testicular biopsy score count—a method for registration of spermatogenesis in human testes: normal values and results in 335 hypogonadal males. *Horm Res Paediatr* 1970; 1:2-25.
42. Oghbaei H, Alipour MR, Hamidian G, Ahmadi M, Ghorbanzadeh V, Keyhanmanesh R. Two months sodium nitrate supplementation alleviates testicular injury in streptozotocin-induced diabetic male rats. *Exp Physiol* 2018; doi: 10.1113/EP087198.
43. Keyhanmanesh R, Hamidian G, Alipour MR, Ranjbar M, Oghbaei H. Protective effects of sodium nitrate against testicular apoptosis and spermatogenesis impairments in streptozotocin-induced diabetic male rats. *Life Sci* 2018; 211:63-73.
44. Samir Zahkok, Nehal Abo-Elnaga, Amel FM Ismail, Esraa Mousa. Studies on fertility of diabetic male rats treated with olive leaves extract. *J Biomed Pharmaceut Res* 2016; 5:18-27.
45. Ahmed, N., Advanced glycation endproducts—role in pathology of diabetic complications. *Diabetes Res Clin Pract* 2005; 67:3-21.
46. Jakuš, V. and N. Rietbrock, Advanced glycation end-products and the progress of diabetic vascular complications. *Physiol Res* 2004; 53:131-142.
47. Tremellen K. Oxidative stress and male infertility—a clinical perspective. *Human Reproduction Update* 2008; 14:243-258.
48. Maedler K, Carr RD, Bosco D, Zuellig RA, Berney T, Donath MY. Sulfonylurea induced β -cell apoptosis in cultured human islets. *J Clin Endocrinol Metab* 2005; 90:501-506.
49. Adaramoye O, Akanni O, Adesanoye O, Labo-Popoola O, Olaremi O. Evaluation of toxic effects of metformin hydrochloride and glibenclamide on some organs of male rats. *Niger J Physiol Sci* 2012; 27:137-144.
50. Alves MG, Socorro S, Silva J, Barros A, Sousa M, Cavaco JE, Oliveira PF. In vitro cultured human Sertoli cells secrete high amounts of acetate that is stimulated by 17 β -estradiol and suppressed by insulin deprivation. *Biochim Biophys Acta* 2012; 1823:1389-1394.
51. Oliveira PF, Alves MG, Rato L, Laurentino S, Silva J, Sá R, Barros A, Sousa M, Carvalho RA, Cavaco JE, Socorro S. Effect of insulin deprivation on metabolism and metabolism-associated gene transcript levels of in vitro cultured human Sertoli cells. *Biochim Biophys Acta* 2012; 1820:84-89.
52. Nakayama Y, Yamamoto T, Abé SI. IGF-I, IGF-II and insulin promote differentiation of spermatogonia to primary spermatocytes in organ culture of newt testes. *Int J Dev Biol* 1999; 43:343-347.
53. Geetha R, Yagalakshmi B, Sreeja S, Bhavani K, Anuradha CV. Troloxerutin suppresses lipid abnormalities in the heart of high-fat-high-fructose diet-fed mice. *Mol cell biochem* 2014; 387:123-134.
54. Lu J, Wu DM, Zheng ZH, Zheng YL, Hu B, Zhang ZF. Troloxerutin protects against high cholesterol-induced cognitive deficits in mice. *Brain* 2011; 134:783-797.
55. Kawamura N, Ookawara T, Suzuki K, Konishi K, Mino M, Taniguchi N. Increased glycated Cu, Zn-superoxide dismutase levels in erythrocytes of patients with insulin-dependent

- diabetes mellitus. *J Clin Endocrin Metab* 1992; 74:1352-1354.
56. Karimi J, Goodarzi MT, Tavilani H, Khodadadi I, Amiri I. Relationship between advanced glycation end products and increased lipid peroxidation in semen of diabetic men. *Diabetes Res Clin Pract* 2011; 91: 61-66.
57. Ballester J, Muñoz MC, Domínguez J, Rigau T, Guinovart JJ, Rodríguez-Gil JE. Insulin-dependent diabetes affects testicular function by FSH-and LH-linked mechanisms. *J Androl* 2004; 25: 706-719.
58. La Vignera S, Condorelli R, Vicari E, D'Agata R, Calogero AE. Diabetes mellitus and sperm parameters. *J Androl* 2012; 33:145-153.
59. Kim ST, Moley KH. Paternal effect on embryo quality in diabetic mice is related to poor sperm quality and associated with decreased glucose transporter expression. *Reproduction* 2008; 136:313-322.
60. Baccetti B, La Marca A, Piomboni P, Capitani S, Bruni E, Petraglia F, De Leo V. Insulin-dependent diabetes in men is associated with hypothalamo-pituitary derangement and with impairment in semen quality. *Hum Reprod* 2002; 17:2673-2677.
61. Vinothkumar R, Vinoth Kumar R, Sudha M, Viswanathan P, Balasubramanian T, Nalini N. Modulatory effect of troloxerutin on biotransforming enzymes and preneoplastic lesions induced by 1, 2-dimethylhydrazine in rat colon carcinogenesis. *Exp Mol Pathol* 2014; 96: 15-26.
62. Kanter M, Aktas C, Erboga M. Protective effects of quercetin against apoptosis and oxidative stress in streptozotocin-induced diabetic rat testis. *Food Chem Toxicol* 2012; 50:719-725.
63. Lu J, Wu DM, Hu B, Cheng W, Zheng YL, Zhang ZF, Ye Q, Fan SH, Shan Q, Wang YJ. Chronic administration of troloxerutin protects mouse brain against D-galactose-induced impairment of cholinergic system. *Neurobiol Learn Mem* 2010; 93:157-164.

Archive of SID