

The effects of supraphysiological levels of testosterone on neural networks upstream of gonadotropin-releasing hormone neurons

Mohammad Saied Salehi^{1,2}, Homayoun Khazali^{2*}, Fariba Mahmoudi³, Mahyar Janahmadi⁴

¹ Clinical Neurology Research Center, Shiraz University of Medical Sciences, Shiraz, Iran

² Department of Animal Physiology, Faculty of Life Sciences and Biotechnology, Shahid Beheshti University, Tehran, Iran

³ Faculty of Basic Sciences, University of Mohaghegh Ardabili, Ardabil, Iran

⁴ Neuroscience Research Center and Department of Physiology, Faculty of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran

ARTICLE INFO

Article type:
Original article

Article history:
Received: Nov 9, 2018
Accepted: Jan 12, 2019

Keywords:
GnRH
Kisspeptin
Neurokinin B
RFRP
Testosterone

ABSTRACT

Objective(s): Several pathological conditions are associated with hyper-production of testosterone; however, its impacts are not well understood. Hence, we evaluated the effects of supraphysiological levels of testosterone on gonadotropin-releasing hormone (GnRH) system in the hypothalamus of male rats. Also, we assessed the expression of two excitatory (kisspeptin and neurokinin-B) and two inhibitory (dynorphin and RFamide-related-peptide) neuropeptides upstream of GnRH neurons as possible routes to relay androgen information.

Materials and Methods: Gonadectomized (GDX) male rats received single injection of 100, 250 or 500 mg/kg testosterone undecanoate and three weeks later, posterior (PH) and anterior (AH) hypothalamus was dissected for evaluation of target genes using quantitative RT-PCR.

Results: We found that GnRH mRNA in the PH was high in GDX rats and 500 mg/kg testosterone reduced GnRH level expression. Finding revealed extremely high level of Kiss1 mRNA in the PH of GDX rats. However, in GDX rats treated with different levels of testosterone, Kiss1 expression was not significantly different than control. We also found that testosterone replacement increased the Kiss1 mRNA level in the AH. Moreover, neurokinin-B mRNA level in PH of GDX rats was similar to control. However, excess testosterone levels were effective in significantly inducing the down-regulation of neurokinin-B expression. The basal level of dynorphin mRNA was increased following testosterone treatments in the AH, where we found no significant difference in the level of RFamide-related-peptide mRNA between the experimental groups.

Conclusion: Excess levels of testosterone could act differently from its physiological concentration to regulate hypothalamic androgen sensitive neurons to control GnRH cell.

► Please cite this article as:

Salehi MS, Khazali H, Mahmoudi F, Janahmadi M. The effects of supraphysiological levels of testosterone on neural networks upstream of gonadotropin-releasing hormone neurons. Iran J Basic Med Sci 2019; 22:1065-1072. doi: 10.22038/ijbms.2019.36127.8605

Introduction

There are several pathological conditions that are associated with hyper-production of testosterone. Furthermore, androgen abuse among athletes has become a major public health issue (1). It has been reported that supraphysiological levels of androgen have adverse effects on various body systems, such as cardiovascular and central nervous system (2) and perhaps, hypothalamus is mostly affected by testosterone in the brain.

Gonadotropin-releasing hormone (GnRH) is the hypothalamic neuropeptide that controls the reproductive endocrine system and is considered as ultimate output of the central nervous system driving fertility in all mammals. GnRH induces gonadotropin release from the pituitary gland, leading to production of gonadal sex steroids, which then through positive and negative feedback mechanisms control GnRH neuronal activity. In males, the negative feedback actions of testosterone regulate GnRH, and in turn, gonadotropin (3). GnRH neurons do not express androgen receptors (4); therefore, steroid actions on GnRH neurons must be mediated via upstream steroid-sensitive neurons.

Discovery of kisspeptin as the most potent stimulator of GnRH neurons (5) has led to better understanding of neuroendocrine reproductive regulation, which is considered as a missing link in the sex steroid feedback control of GnRH. In the rat hypothalamus, kisspeptin cells are located in the anteroventral periventricular (AVPV) and arcuate (ARC) nuclei (6). Although our knowledge about the mechanisms involve in positive feedback action of estradiol are accumulating rapidly, the underlying mechanisms of steroids negative feedback remain unclear. It has been proposed that arcuate kisspeptin cells mediate negative feedback action of sex hormones on GnRH secretion in both male and female rodents; however, pathways through which ARC kisspeptin expressing neurons regulate GnRH neurons are not fully understood (7).

In addition, several lines of evidence have suggested that ARC kisspeptin neurons regulate pulsatile GnRH secretion via the autaptic activity of the neurokinin B (NKB) and dynorphin (DYN), involved in generating the rhythmic discharge of kisspeptin (8, 9). It was reported that NKB by interacting with kisspeptin controls GnRH secretion in the rat (10) and

*Corresponding author: Homayoun Khazali. Department of Animal Physiology, Faculty of Life Sciences and Biotechnology, Shahid Beheshti University, Tehran, Iran. Tel: +98-21-29903192; Email: h_khazali@sbu.ac.ir

DYN presumably relays inhibitory signals via kisspeptin cells (9). RFamide related peptide-3 (RFRP-3) belongs to the RFamide peptides family and in contrast to kisspeptin, hyperpolarizes GnRH neurons and inhibits the neuropeptide secretion. RFRP-3 is considered as a mammalian ortholog of avian gonadotropin-inhibitory hormone (11-13).

Since supraphysiological levels of testosterone might act differently from the physiological concentration of the steroid, we hypothesized that GnRH system might also be affected differently after the testosterone intervention. Accordingly, we investigated the impacts of abnormal testosterone levels on GnRH expression in hypothalamus of male rats. Furthermore, we evaluated the expression of two excitatory (kisspeptin and NKB) and two inhibitory (DYN and RFRP-3) neuropeptides upstream of GnRH neurons as possible messengers to relay testosterone signal.

Materials and Methods

Animals

In the present study, 30 randomly selected adult male Sprague-Dawley rats (weighing 200-250 g) were housed under controlled conditions (12 hr light/dark cycle, 22 °C temperature and ~40% humidity) and allowed *ad libitum* access to standard food and water. This experiment was approved by Animal Care Committee of the Shahid Beheshti University, Tehran, Iran (approval number: D/920/1010) and carried out in compliance with the recommendations of the Care and Use of Laboratory Animals (National Academy Press, 1996, Washington, USA).

Gonadectomy and testosterone treatments

All adult male rats were anesthetized with ketamine (100 mg/kg)/xylazine (10 mg/kg) and divided into five groups: one group of sham-operated non-gonadectomized (CTL group), one group of bilaterally

gonadectomized rats received normal saline (GDx group) and three groups of GDx rats received single intramuscular injection of 100 (GDx+T100), 250 (GDx+T250) and 500 (GDx+T500) mg/kg testosterone undecanoate (Nebido, Bayer Pharma, Berlin, Germany) to produce steady state supraphysiological levels of steroid hormone during the experiment. Testosterone undecanoate is a depot preparation allowing continuous slow release of free testosterone from the depot, which reaches the brain. These doses were chosen based on Callies *et al.* work who showed that a single injection of testosterone undecanoate is a well suited for long-term substitution in hypogonadism and is a convenient and effective tool for inducing a desire testosterone level for at least four weeks (14). A schema of experimental design is illustrated in Figure 1.

Tissue collection

Three weeks following interventions, rats were anesthetized, brains were immediately removed and hypothalamus was dissected rostral to the anterior border of the optic chiasm, caudal to the posterior border of the mammillary body, 2 mm lateral to the third ventricle and dorsal to the ventral border of the thalamus. Since kisspeptin cells are located in two distinct hypothalamic nuclei, the third coronal cut was made just rostral to the infundibulum in the middle of the optic tract (15). Finally, posterior (PH) and anterior (AH) hypothalami were snap-frozen and stored at -80 °C until further processing.

RNA extraction, cDNA synthesis and qRT-PCR

Total RNAs extraction (YTzol Pure RNA buffer, Yekta Tajhiz Azma, Iran), DNaseI treatment (Thermo Scientific, USA) and cDNA synthesis (Easy cDNA Synthesis kit, Pars Tous, Iran) were performed on the AH and PH samples based on the manufacturer’s instructions.

To evaluate GnRH, Kiss1, NKB, DYN and RFRP-3

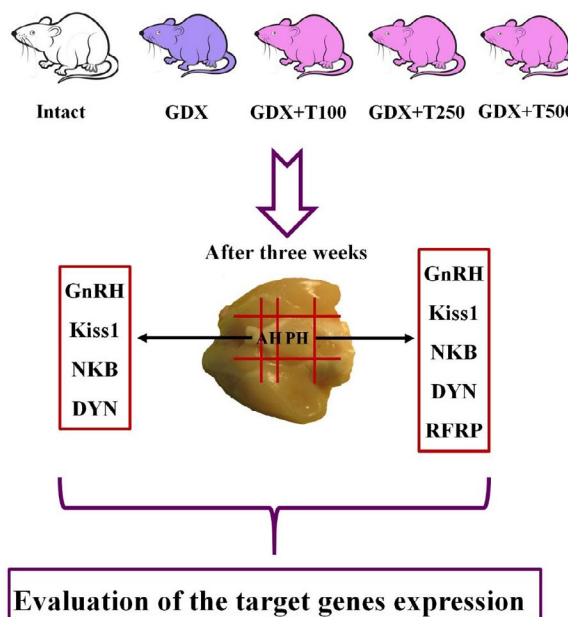


Figure 1. A schema of the experimental design. GDx: gonadectomized rats, GDx+T100: gonadectomized rat received 100 mg/kg testosterone undecanoate. GDx+T250: gonadectomized rat received 250 mg/kg testosterone undecanoate. GDx+T500: gonadectomized rat received 500 mg/kg testosterone undecanoate. PH: posterior hypothalamus, AH: anterior hypothalamus

Table 1. Primer sequences (5'-3') used in quantitative polymerase chain reaction (qPCR)

Gene	Accession number	Sequence	Amplicon (bp)
GnRH	NM_012767	F-GCCGCTGTTGTTCTGTTGACTG R-CCTCCTCCTGCCCATCTCTTG	133
Kiss1	NM_181692	F-TGATCTCGCTGGCTTCTTGGC R-GGGTTCAGGGTTCACCACAGG	98
RFRP-3	NM_023952	F-GAGTCCTGGTCAAGAGCAAC R-ACTGGCTGGAGGTTCCTAT	93
NKB	NM_019162	F-GGAAGGATGCTGAAAGTGCTGAG R-GGGAGTGTCTGGTTGGCTGTTT	130
DYN	NM_019374	F-CGGCGTCAGTCAAGTGGTG R-AGCAAGCGAAGGAATAAGCAGAGG	150
Actin, Beta	NM_031144	F-TCTATCCTGGCCTCACTGTC R-AACGCAGCTCAGTAACAGTCC	122

GnRH: gonadotropin-releasing hormone, RFRP-3: RFamide related peptide-3, NKB: neurokinin B, DYN: dynorphin

transcripts, triplicate reactions were carried out on the cDNA samples. In brief, twenty-microliter reaction mixture was prepared, consisting of RealQ Plus Master Mix Green (Ampliqon, Denmark), first strand cDNA template and appropriate qRT-PCR primer set (specific primers presented in Table 1). Thermo-cycling conditions were 95 °C for 10 min before 40 cycles of 95 °C for 30 sec, 56 °C for 30 sec and 72 °C for 30 sec on an ABI StepOne Real-Time PCR system (Applied Biosystems, USA). Melting curve analysis showed a single amplification peak for each reaction. The Ct value for each target was normalized to the copy number of the beta-actin transcript for each sample. Furthermore, a single band of the expected size was observed for amplified products (5 µl) that were subjected to electrophoresis on a 1% agarose gel stained with ethidium bromide. Fold changes in expression were calculated using the arithmetic formula $2^{-\Delta\Delta CT}$ (16).

Relative expression of GnRH, Kiss1, NKB and DYN genes were assessed on both the PH and AH samples, while expression of RFRP-3 was evaluated only on the PH.

Hormone assay

Serum testosterone and luteinizing hormone (LH) concentrations were measured just before tissue collection, using testosterone [¹²⁵I] RIA kit and rLH [¹²⁵I] RIA kit (Izotop, Hungary) according to the manufacturer’s instructions.

Statistical analysis

Obtained data were subjected to the normality test, and comparisons between groups were made by one-way ANOVA plus the Tukey *post hoc* test using GraphPad Prism (Version 7.03, GraphPad Software, Inc., San Diego, CA). The mean±SEM are reported in the text and $P < 0.05$

was considered to be statistically significant.

Results

Three weeks following interventions, serum testosterone concentration was measured to confirm the effectiveness of the drug in producing the desired hormone levels. As shown in Table 2, gonadectomy reduced mean testosterone concentration and androgen replacements, which dose dependently elevated the levels of steroid hormone. In addition, to determine whether different levels of testosterone have different ability to exert negative feedback action on gonadotropin secretion, serum LH concentration was also assessed. Although mean LH concentration was increased by more than 15-fold following gonadectomy, all three androgen treatments preserved gonadotropin secretion as much as the control (Table 2).

GnRH expression in response to supraphysiological levels of testosterone

Here, quantitative RT-PCR was used to evaluate the ability of gonadectomy and supraphysiological levels of testosterone replacement to regulate gene expression in the AH and PH three weeks after interventions. As shown in Figure 2, GnRH transcript in the PH was high in GDX rats ($P < 0.01$), and 500 mg/kg testosterone reduced the level of GnRH mRNA by about 50% ($P < 0.05$). GnRH expression in the PH of rats was unaffected by the GDX+T100 or GDX+T250 treatments ($P > 0.05$). Neither gonadectomy nor testosterone treatments affected the GnRH transcript in the AH ($P > 0.05$).

Effects of testosterone interventions on the kisspeptin and neurokinin-B transcripts

Next, we evaluated the effects of testosterone on

Table 2. Mean±SEM serum testosterone and luteinizing hormone (LH) concentration three weeks following interventions

	Experimental groups				
	CTL	GDX	GDX+T100	GDX+T250	GDX+T500
Mean testosterone concentration (ng/ml)	1.52±0.21	0.17±0.01	4.41±0.85	8.86±0.63	13.15±2.45
Mean LH concentration (ng/ml)	1.96±0.24	31.94±6.09	1.70±0.21	1.42±0.19	1.83±0.07

CTL: sham-operated non-gonadectomized, GDX: gonadectomized rats, GDX+T100: GDX rat received 100 mg/kg testosterone undecanoate. GDX+T250: GDX rat received 250 mg/kg testosterone undecanoate. GDX+T500: GDX rat received 500 mg/kg testosterone undecanoate

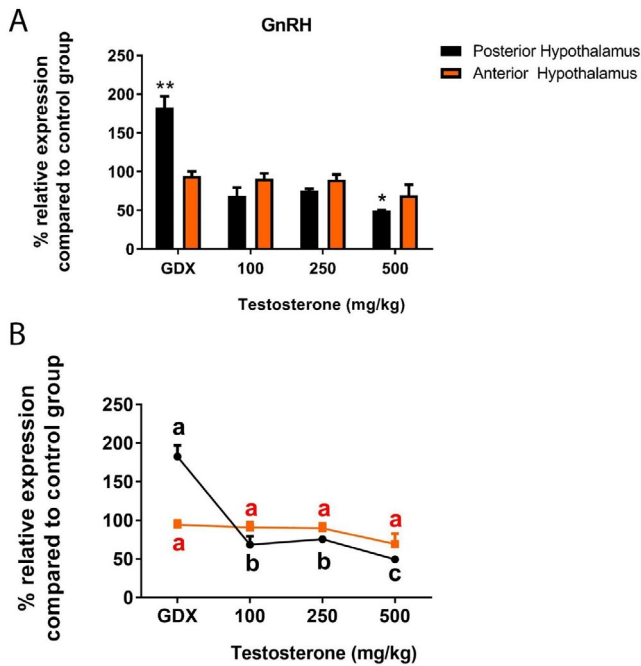


Figure 2. A) Relative expression of gonadotropin-releasing hormone (GnRH) mRNA in the rat posterior and anterior hypothalamus following testosterone interventions compared to control group. * $P < 0.05$, ** $P < 0.01$, ns: non-significant; B) Statistical comparison between groups; values with different superscripts are significantly different ($P < 0.05$). Data are represented as mean \pm SEM

two excitatory neuropeptides upstream of GnRH neurons. It was previously reported that kisspeptin in the ARC and AVPV is regulated by physiological level of testosterone (17). In the present work, we studied the effect of supraphysiological level of testosterone. We found, extremely high (more than 2600%) level of Kiss1

transcript in the PH of GDX rats ($P < 0.001$). However, in GDX rats treated with different levels of testosterone, Kiss1 expression was not significantly different than the control rats ($P > 0.05$) (Figure 3A). We also found that testosterone replacement elevated Kiss1 transcript level in the AH of GDX animals by around 700% when compared to the control group ($P < 0.001$) (Figure 3B).

NKB is another neuropeptide in the hypothalamus that has crucial function in regulating GnRH system. NKB mRNA in PH of GDX rats was similar to those of the control ($P > 0.05$). However, all three different treatments were able to induce a significant down-regulation in gene expression ($P < 0.05$) (Figure 4). On the other hand, the level of NKB mRNA in the AH of gonadectomized rats was not significantly different from that of AH in control rats or that of GDX rats receiving 100 mg/kg testosterone ($P > 0.05$). However, treatment with 250 or 500 mg/kg testosterone increased NKB transcript slightly but significantly in the AH ($P < 0.05$) (Figure 4).

Dynorphin and RFRP expression in response to testosterone replacements

Next, the effects of supraphysiological levels of testosterone on two inhibitory neuropeptides – DYN and RFRP– that can regulate GnRH cells, were investigated. DYN expression was not significantly different with/without testosterone in the PH ($P > 0.05$) (Figure 5). Furthermore, the basal level of DYN transcript elevated after testosterone interventions in the AH. It should be noted that testosterone only at its highest dose could statistically up-regulate DYN expression ($P < 0.05$) (Figure 5). RFRP mRNA in the hypothalamus of GDX rats with and without testosterone was also determined by qRT-PCR. We found no significant difference in the level of RFRP mRNA between the experimental groups ($P > 0.05$) (Figure 6).

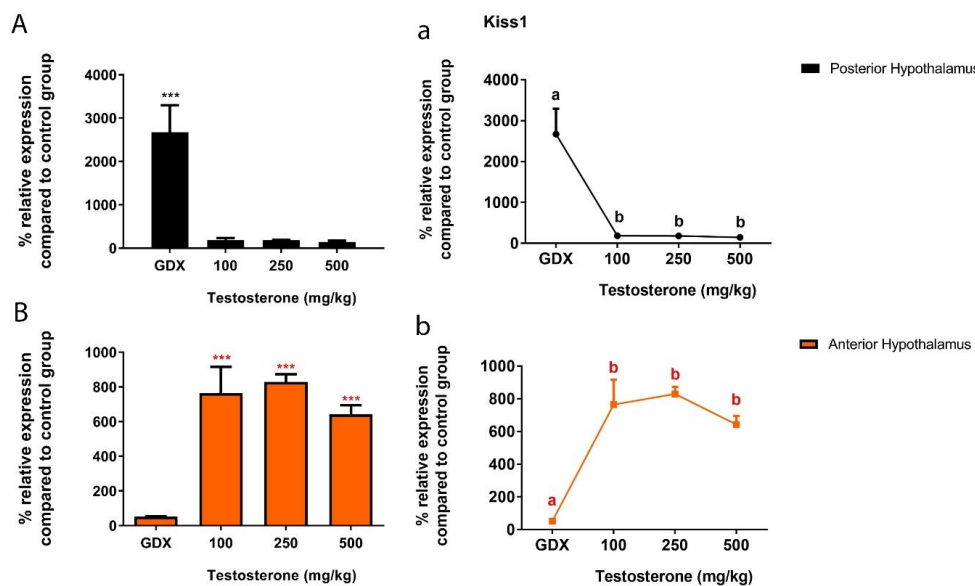


Figure 3. A and B) Relative expression of Kiss1 mRNA in the rat posterior and anterior hypothalamus following testosterone interventions compared to control group. *** $P < 0.001$. a and b) Statistical comparison between groups in the posterior and anterior hypothalamus; values with different superscripts are significantly different ($P < 0.05$). Data are represented as mean \pm SEM

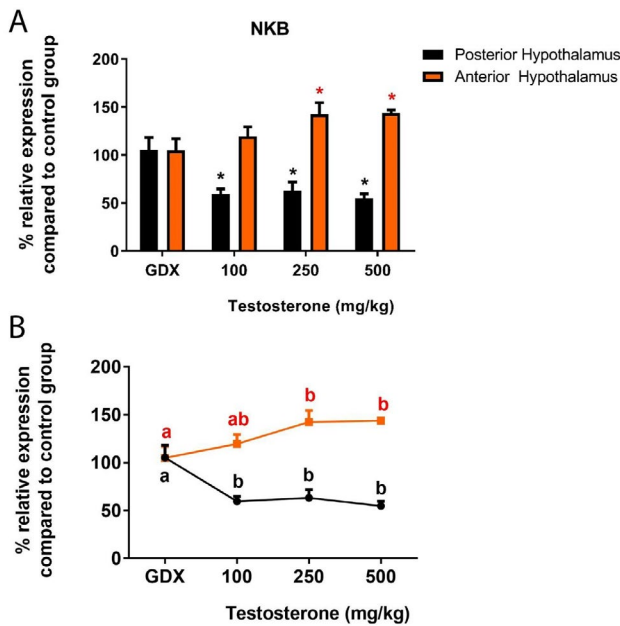


Figure 4. A) Relative expression of neurokinin-B mRNA in the rat posterior and anterior hypothalamus following testosterone interventions compared to control group. * $P < 0.05$. B) Statistical comparison between groups; values with different superscripts are significantly different ($P < 0.05$). Data are represented as mean \pm SEM

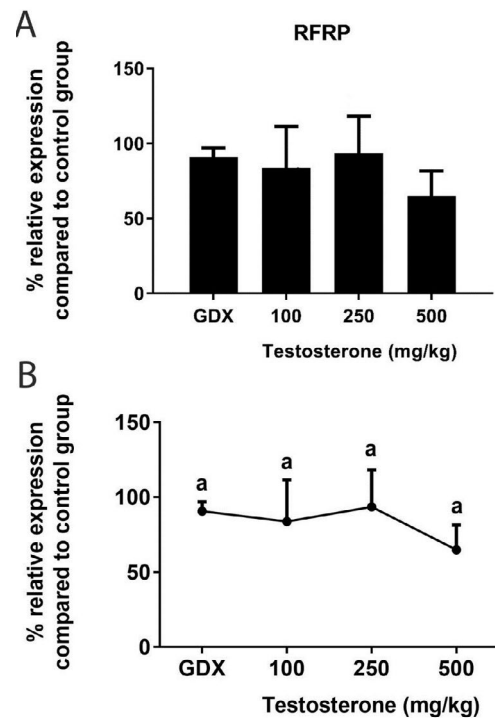


Figure 6. A) Relative expression of Rfamide-related-peptide-3 mRNA in the rat posterior hypothalamus following testosterone interventions compared to control group. B) Statistical comparison between groups; values with different superscripts are significantly different ($P < 0.05$). Data are represented as mean \pm SEM

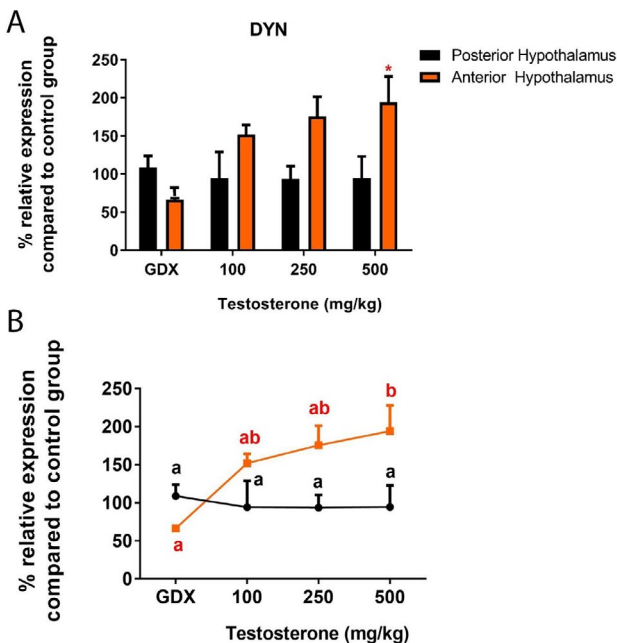


Figure 5. A) Relative expression of dynorphin mRNA in the rat posterior hypothalamus following testosterone interventions compared to control group. * $P < 0.05$. B) Statistical comparison between groups; values with different superscripts are significantly different ($P < 0.05$). Data are represented as mean \pm SEM

Discussion

In the present study, we confirmed the hypothesis that supraphysiological levels of testosterone can act differently from its physiological concentration to regulate hypothalamic androgen sensitive neurons in order to control GnRH cell. It has been known for while

that testosterone exerts negative feedback action on gonadotropin secretion, demonstrated by an elevated release of LH following castration. Although testicular steroid can act at pituitary gonadotrope and/or hypothalamus levels, obtained data from hypothalamo-pituitary disconnection models revealed that the predominant site at which testosterone act to regulate the secretion of LH is within the central nervous system (3). At odds with obvious LH response to testosterone, the effect of testicular steroid on GnRH level were often time inconclusive and contradictory. Previous studies showed either a decrease (18, 19), increase (20, 21) or no change (22, 23) in the content of hypothalamic GnRH following castration. Therefore, we first evaluated whether gonadectomy and testosterone treatments can alter hypothalamic GnRH expression. Our findings showed that gonadectomy increased GnRH mRNA in the PH, and the highest dose of testosterone treatment had a reverse effect on GnRH expression; however, neither gonadectomy nor testosterone treatments affected GnRH mRNA in AH.

There is no doubt that gonadal steroids exert their impacts on GnRH through upstream steroid-sensitive neurons. Kisspeptin neurons as the most important stimulatory input to GnRH cells have been suggested to have a role in testosterone negative feedback mechanism. In this regard, it was reported that castration caused a significant increase in Kiss1 expression in the arcuate of many species such as rats (24), mice (17), goat (25) and monkey (26), which was reversed with testosterone replacement. Similarly, in the present study we observed significant elevation of Kiss1 mRNA in the PH (where

ARC nucleus is located) following gonadectomy, while all supraphysiological levels of testosterone decreased GnRH expression to levels found in the control rats. Although kisspeptin cells in ARC are ideally located to act as the missing link in the sex steroid feedback control of GnRH, presently there is little evidence for anatomical connection between GnRH neurons and ARC kisspeptin neurons (7).

On the other hand, our results showed that excess testosterone exposure can lead to up-regulation of kisspeptin expression (more than 700%) in the AH where AVPV nucleus is located. Also, Kiss1 mRNA in the GDX rats decreased by around 50%, but it was not statistically significant. In female rodents, it is well-documented that AVPV kisspeptin neurons mediate estrogen positive feedback effect to stimulate the preovulatory GnRH/LH surge. In males, kisspeptin cells in AVPV have the same response similar to the female AVPV, but its physiological role is not well-defined as males normally do not generate sex steroid positive feedback (27). Since testosterone regulation of kisspeptin in the AVPV is mediated via estrogen signaling (17), supraphysiological levels of testosterone, which were employed in the present study, could increase Kiss1 expression in the AH after aromatization to estrogen.

NKB is another neuropeptide with crucial role in the control of reproductive function and inactivating mutations in the genes encoding NKB or its receptors, which results in hypogonadotropic hypogonadism (28). In the rat hypothalamus, majority of NKB mRNA expressing neurons locate in the lateral hypothalamic area (LHA) and ARC (29). Since arcuate NKB neurons co-express androgen receptor (30) and these neurons directly interact with GnRH cells, especially at the median eminence (31), we evaluated the effects of testicular steroid hormone on NKB expression. Here, gonadectomy did not affect NKB expression in the hypothalamus, although, supraphysiological doses of testosterone clearly reduced NKB mRNA in the PH. Previous reports showed either increase (32) or no change (33) in the level of arcuate NKB expression following castration.

Also in female rats, NKB expression in the ARC was reduced during the afternoon of pro-estrus when circulating estradiol is in its maximum level. Similarly, ovariectomized rats treated with estradiol had fewer NKB mRNAs in the ARC compare to non-treated group (10). Furthermore, based on a postmortem investigation, older men with lower testosterone concentrations had greater numbers of NKB neurons in the infundibular nucleus than young men (34).

On the other hand, we found that in GDX rats receiving 250 or 500 mg/kg testosterone, NKB mRNAs in the AH were slightly but significantly higher than the other groups. Interestingly, estradiol treatment induced NKB expression in the LHA of ovariectomized rats (35). Therefore, based on our findings, high testosterone concentrations might exert dual opposite effects on NKB expression in the ARC and LHA. However, further clarification is required to define whether the effects of testosterone on NKB neurons is direct or mediated via estrogen signaling.

DYN is another element, known for a long time as

an inhibitor of gonadotropin secretion (36, 37), which belongs to endogenous opioid peptides family and is considered to mediate the negative feedback effects of progesterone on GnRH pulse frequency (38). Also, a number of pharmacological evidences have proposed that endogenous opioid peptides might be involved in mediating the negative feedback effects of testosterone (3). Our results showed that DYN expression was not affected with or without testosterone in the PH; however, androgen treatment could elevate DYN mRNA in the AH. In line with our findings, Dudek *et al.* found that orchidectomy did not significantly change the number of DYN cells in the ARC of rats (33).

There are fewer DYN neurons in the infundibular nucleus of the postmenopausal women compared to the premenopausal group (39). Furthermore, ovariectomy reduces the number of DYN mRNA expressing neurons in the preoptic area, AH and ARC in ewes (40), which raise the possibility of inducing DYN expression by estradiol. However, it was reported that expression of DYN is inhibited by estradiol in female mice (8, 41). As DYN mRNAs have been widely distributed throughout the hypothalamus (42); hence, *in situ* analysis such as immunohistochemical study is required to understand the exact localization of the observed changes.

In rodents, RFRP neurons are exclusively located in the dorsomedial hypothalamic nucleus and inhibit GnRH neural activity by sending projections to these cells (11-13). Although androgen receptor was detected in RFRP neurons of male hamster (43), virtually no RFRP neurons in male and female mice co-expressed androgen receptor (44). In addition, estrogen receptor- α mRNA was expressed in approximately 40% of RFRP neurons in hamster (43), 18% in ovariectomized mice (45) and 25% in male and female intact mice (44). Accordingly, we hypothesized that excess testosterone levels might affect RFRP expression directly or following aromatization. Our findings showed that neither gonadectomy nor testosterone treatments affected RFRP mRNA in the hypothalamus. Similarly, there was no statistical difference in RFRP expression between castrated and sham-operated hamsters. Also, the RFRP expression in hamsters treated with testosterone for 4 weeks was not statistically different from castrated group (46). In addition, it was reported that RFRP expression is moderately regulated by estradiol in male and female mice and unaffected by dihydrotestosterone. Therefore, androgen-signaling seems not to be an important regulator of RFRP neurons.

Limitation

In the present study, we assessed target genes in the AH and PH; however, for more accurate evaluation, dissecting each hypothalamic nucleus is recommended. In addition, for better understating of supraphysiological levels of testosterone effects, evaluation of target genes expression at the level of protein using techniques such as immunohistochemistry or western-blot are recommended.

Conclusion

Our results showed that supraphysiological levels of testosterone elevate relative expression of kisspeptin,

NKB and DYN in the AH of male rats while decreasing GnRH and NKB mRNAs in the PH. Thus, excess level of the testicular steroid hormone affects neural networks upstream of GnRH neurons differently from its physiological concentration. Therefore, testosterone abuse might have long-term undesirable effects on the brain due to its adverse impacts on various neuropeptides system.

Acknowledgment

This study was financially supported by Shahid Beheshti University Grant No. 60092 and the results described in this paper were part of PhD student thesis. The authors are thankful to the Neuroscience Research Center, Shahid Beheshti University of Medical Sciences, for the technical support of this work. Also the authors wish to thank Mr. H. Argasi at the Research Consultation Center (RCC) of Shiraz University of Medical Sciences for his invaluable assistance in editing this manuscript.

Conflicts of Interest

The authors declare that they have no conflict of interest.

References

- Harmer PA. Anabolic-androgenic steroid use among young male and female athletes: is the game to blame? *Br J Sports Med* 2010; 44:26-31.
- Basaria S. Androgen abuse in athletes: detection and consequences. *J Clin Endocrinol Metab* 2010; 95:1533-1543.
- Tilbrook A, Clarke I. Negative feedback regulation of the secretion and actions of gonadotropin-releasing hormone in males. *Biol Reprod* 2001; 64:735-742.
- Huang X, Harlan RE. Absence of androgen receptors in LHRH immunoreactive neurons. *Brain Res* 1993; 624:309-311.
- Rønnekleiv OK, Kelly MJ. Kisspeptin excitation of GnRH neurons. In: Kauffman AS, Smith JT, editors. *Kisspeptin signaling in reproductive biology*. New York: Springer; 2013. p. 113-131.
- Mikkelsen JD, Simonneaux V. The neuroanatomy of the kisspeptin system in the mammalian brain. *Peptides* 2009; 30:26-33.
- Herbison AE. Physiology of the adult gonadotropin-releasing hormone neuronal network. In: Plant TM, Zeleznik AJ, editors. *Knobil and Neill's physiology of reproduction (Fourth Edition)*. San Diego: Academic Press; 2015. p. 399-467.
- Navarro VM, Gottsch ML, Chavkin C, Okamura H, Clifton DK, Steiner RA. Regulation of gonadotropin-releasing hormone secretion by kisspeptin/dynorphin/neurokinin B neurons in the arcuate nucleus of the mouse. *J Neurosci* 2009; 29:11859-11866.
- Wakabayashi Y, Nakada T, Murata K, Ohkura S, Mogi K, Navarro VM, et al. Neurokinin B and dynorphin A in kisspeptin neurons of the arcuate nucleus participate in generation of periodic oscillation of neural activity driving pulsatile gonadotropin-releasing hormone secretion in the goat. *J Neurosci* 2010; 30:3124-3132.
- Navarro VM, Castellano JM, McConkey SM, Pineda R, Ruiz-Pino F, Pinilla L, et al. Interactions between kisspeptin and neurokinin B in the control of GnRH secretion in the female rat. *Am J Physiol Endocrinol Metab* 2011; 300:E202-E210.
- Kriegsfeld LJ, Gibson EM, Williams W, Zhao S, Mason AO, Bentley GE, et al. The roles of RFamide-Related Peptide-3 in mammalian reproductive function and behaviour. *J Neuroendocrinol* 2010; 22:692-700.
- Salehi MS, Tamadon A, Shirazi MRJ, Namavar MR, Zamiri MJ. The role of arginine-phenylalanine-amide-related peptides in mammalian reproduction. *Int J Fertil Steril* 2015; 9:268-276.
- Wang H, Khoradmehr A, Jalali M, Salehi MS, Tsutsui K, Jafarzadeh Shirazi MR, et al. The roles of RFamide-related peptides (RFRPs), mammalian gonadotropin-inhibitory hormone (GnIH) orthologues in female reproduction. *Iran J Basic Med Sci* 2018; 21:1210-1220.
- Callies F, Kollenkirchen U, Von Zur Mühlen C, Tomaszewski M, Beer S, Allolio B. Testosterone undecanoate: a useful tool for testosterone administration in rats. *AMA Arch Gen Psychiatry* 2003; 111:203-208.
- Salehi MS, Namavar MR, Jafarzadeh Shirazi M, Rahmanifar F, Tamadon A. A simple method for isolation of the anteroventral periventricular and arcuate nuclei of the rat hypothalamus. *Int J Exp Clin Anat* 2012; 6-7:48-51.
- Livak KJ, Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the 2- $\Delta\Delta$ CT method. *Methods* 2001; 25:402-408.
- Smith JT, Dungan HM, Stoll EA, Gottsch ML, Braun RE, Eacker SM, et al. Differential regulation of KiSS-1 mRNA expression by sex steroids in the brain of the male mouse. *Endocrinology* 2005; 146:2976-2984.
- Park Y, Dai Park S, Kyoo CW, Kim K. Testosterone stimulates LH-RH-like mRNA level in the rat hypothalamus. *Brain Res* 1988; 451:255-260.
- Zanisi M, Celotti F, Ferraboschi P, Motta M. Testosterone metabolites do not participate in the control of hypothalamic LH-releasing hormone. *J Endocrinol* 1986; 109:291-296.
- Roselli C, Kelly M, Ronnekleiv O. Testosterone regulates progonadotropin-releasing hormone levels in the preoptic area and basal hypothalamus of the male rat. *Endocrinology* 1990; 126:1080-1086.
- Selmanoff M, Shu C, Petersen SL, Barraclough CA, Zoeller RT. Single cell levels of hypothalamic messenger ribonucleic acid encoding luteinizing hormone-releasing hormone in intact, castrated, and hyperprolactinemic male rats. *Endocrinology* 1991; 128:459-466.
- Rothfeld J, Hejtmancik J, Conn P, Pfaff D. LHRH messenger RNA in neurons in the intact and castrate male rat forebrain, studied by in situ hybridization. *Exp Brain Res* 1987; 67:113-118.
- Wiemann JN, Clifton DK, Steiner RA. Gonadotropin-releasing hormone messenger ribonucleic acid levels are unaltered with changes in the gonadal hormone milieu of the adult male rat. *Endocrinology* 1990; 127:523-532.
- Irwig MS, Fraley GS, Smith JT, Acohido BV, Popa SM, Cunningham MJ, et al. Kisspeptin activation of gonadotropin releasing hormone neurons and regulation of KiSS-1 mRNA in the male rat. *Neuroendocrinology* 2004; 80:264-272.
- Matsuyama S, Ohkura S, Mogi K, Wakabayashi Y, Mori Y, Tsukamura H, et al. Morphological evidence for direct interaction between kisspeptin and gonadotropin-releasing hormone neurons at the median eminence of the male goat: an immunoelectron microscopic study. *Neuroendocrinology* 2011; 94:323-332.
- Shibata M, Friedman R, Ramaswamy S, Plant T. Evidence that down regulation of hypothalamic KiSS-1 expression is involved in the negative feedback action of testosterone to regulate luteinising hormone secretion in the adult male rhesus monkey (*Macaca mulatta*). *J Neuroendocrinol* 2007; 19:432-438.
- Smith JT. Sex steroid regulation of kisspeptin circuits. In: Kauffman AS, Smith JT, editors. *Kisspeptin signaling in reproductive biology*. New York: Springer; 2013. p. 275-295.
- Grachev P, Millar R, O'byrne K. The role of neurokinin B signalling in reproductive neuroendocrinology.

- Neuroendocrinology 2013; 99:7-17.
29. Navarro VM, Ruiz-Pino F, Sánchez-Garrido MA, García-Galiano D, Hobbs SJ, Manfredi-Lozano M, *et al.* Role of neurokinin B in the control of female puberty and its modulation by metabolic status. *J Neurosci* 2012; 32:2388-2397.
30. Ciofi P, Krause JE, Prins GS, Mazzuca M. Presence of nuclear androgen receptor-like immunoreactivity in neurokinin B-containing neurons of the hypothalamic arcuate nucleus of the adult male rat. *Neurosci Lett* 1994; 182:193-196.
31. Krajewski SJ, Anderson MJ, Iles-Shih L, Chen KJ, Urbanski HF, Rance NE. Morphologic evidence that neurokinin B modulates gonadotropin-releasing hormone secretion via neurokinin 3 receptors in the rat median eminence. *J Comp Neurol* 2005; 489:372-386.
32. Danzer SC, Price RO, McMullen NT, Rance NE. Sex steroid modulation of neurokinin B gene expression in the arcuate nucleus of adult male rats. *Mol Brain Res* 1999; 66:200-204.
33. Dudek M, Kołodziejcki P, Pruszyńska-Oszmałek E, Ziarniak K, Sliwowska J. Effects of orchidectomy and testosterone replacement on numbers of kisspeptin-, neurokinin B-, and dynorphin A-immunoreactive neurones in the arcuate nucleus of the hypothalamus in obese and diabetic rats. *J Neuroendocrinol* 2017; 29.
34. Molnár CS, Vida B, Sipos MT, Ciofi P, Borsay B, Rác K, *et al.* Morphological evidence for enhanced kisspeptin and neurokinin B signaling in the infundibular nucleus of the aging man. *Endocrinology* 2012; 153:5428-5439.
35. Ruiz-Pino F, Navarro VM, Bentsen A, Garcia-Galiano D, Sanchez-Garrido MA, Ciofi P, *et al.* Neurokinin B and the control of the gonadotropic axis in the rat: developmental changes, sexual dimorphism, and regulation by gonadal steroids. *Endocrinology* 2012; 153:4818-4829.
36. Kinoshita F, Nakai Y, Katakami H, Imura H. Suppressive effect of dynorphin-(1-13) on luteinizing hormone release in conscious castrated rats. *Life Sci* 1982; 30:1915-1919.
37. Schulz R, Wilhelm A, Pirke KM, Gramsch C, Herz A. β -Endorphin and dynorphin control serum luteinizing hormone level in immature female rats. *Nature* 1981; 294:757-759.
38. Goodman RL, Coolen LM, Anderson GM, Hardy SL, Valent M, Connors JM, *et al.* Evidence that dynorphin plays a major role in mediating progesterone negative feedback on gonadotropin-releasing hormone neurons in sheep. *Endocrinology* 2004; 145:2959-2967.
39. Rometo A, Rance NE. Changes in prodynorphin gene expression and neuronal morphology in the hypothalamus of postmenopausal women. *J Neuroendocrinol* 2008; 20:1376-1381.
40. Foradori CD, Goodman RL, Adams VL, Valent M, Lehman MN. Progesterone increases dynorphin concentrations in cerebrospinal fluid and preprodynorphin messenger ribonucleic acid levels in a subset of dynorphin neurons in the sheep. *Endocrinology* 2005; 146:1835-1842.
41. Gottsch ML, Navarro VM, Zhao Z, Glidewell-Kenney C, Weiss J, Jameson JL, *et al.* Regulation of Kiss1 and dynorphin gene expression in the murine brain by classical and nonclassical estrogen receptor pathways. *J Neurosci* 2009; 29:9390-9395.
42. Lin S, Boey D, Lee N, Schwarzer C, Sainsbury A, Herzog H. Distribution of prodynorphin mRNA and its interaction with the NPY system in the mouse brain. *Neuropeptides* 2006; 40:115-123.
43. Kriegsfeld LJ, Mei DF, Bentley GE, Ubuka T, Mason AO, Inoue K, *et al.* Identification and characterization of a gonadotropin-inhibitory system in the brains of mammals. *Proc Natl Acad Sci U S A* 2006; 103:2410-2415.
44. Poling MC, Kim J, Dhamija S, Kauffman AS. Development, sex steroid regulation, and phenotypic characterization of RFamide-related peptide (Rfrp) gene expression and RFamide receptors in the mouse hypothalamus. *Endocrinology* 2012; 153:1827-1840.
45. Molnar C, Kallo I, Liposits Z, Hrabovszky E. Estradiol down-regulates RF-amide-related peptide (RFRP) expression in the mouse hypothalamus. *Endocrinology* 2011; 152:1684-1690.
46. Revel FG, Saboureaux M, Pevet P, Simonneaux V, Mikkelsen JD. RFamide-related peptide gene is a melatonin-driven photoperiodic gene. *Endocrinology* 2008; 149:902-912.