REVIEW ARTICLE

HLA Typing from Serology to Sequencing Era

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ABSTRACT

The last two decades have seen a massive growth in the application of DNA technology in Histocompatibility and Immunogenetics. This review summarises the history and application of DNA methods in this area.

Keywords: DNA Typing; HLA Typing; Polymerase Chain Reaction

Serological Techniques

Traditionally HLA antigens have been defined using serological techniques. These techniques rely on obtaining viable lymphocyte preparations (for HLA class II typing, B lymphocytes are needed) and the availability of suitable antisera to recognise the HLA antigens. The advent of magnetic beads, coated with antibody used for isolation of B lymphocytes, made class II typing easier to accomplish. Reliable antisera was not available commercially and laboratories needed to screen to find their own reagents and exchange these with other such-minded laboratories. This meant a difference in the quality of reagents between laboratories and led to some laboratories producing more accurate results than others. In addition these reagents could not be replenished. Whilst serology performed adequately in typing family members, it proved unsatisfactory in donors for bone marrow typing unrelated transplantation, once the extent of polymorphism was known within "serological identical" specificities. Serological typing was also difficult in those cases of poor cell viability or poor expression and in confirming or refuting phenotypic homozygosity; by implementing a DNA technique for HLA-C alleles we have reduced the homozygosity role in our stem cell donor registry from 50% by serology to 21%. In addition typing of non-Caucasians by serological methods has always proved difficult when the majority of HLA sera available have been derived

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from Caucasian donors.

The last twenty years has seen an exponential growth in the application of DNA technology to the field of Histocompatibility and Immunogentics (H&I). Initially this was confined to a few research laboratories. However, development and application of several different DNA methods by many laboratories has led to the situation whereby nearly every H&I laboratory performs some DNA typing for the detection of HLA alleles.

Restriction Fragment Length Polymorphism (RFLP)

In 1982, Wake et al. described restriction fragment length polymorphisms (RFLPs) using a DRβ probe. 1 Complexities of binding patterns and inter-locus cross hybridisation of probes drew attention to the drawbacks of using full length HLA class II cDNA probes. In order to overcome these problems some investigators used short or exonspecific probes.²⁻⁴ Even at this early stage of development, there were indications that RFLP was more accurate than serology.5 The lack of locusspecific probes limited the characterization of class I RFLP, as cloned class I gene probes cross-hybridised with all members of the class I family. 6 Some HLA-A and -B probes were constructed 7,8 and RFLPs were defined which correlated with serologically defined HLA-A, -B and -C alleles. 9-14

In 1987, two reports using Taq I enzyme showed how short probes for DR β , DQ β and DQ α could be applied sequentially, after dehybridization, to a single membrane. The recognition site of Taq I includes

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the nucleotide dimer CpG and restriction sites containing this dimer show a higher frequency of polymorphism in human DNA than other restriction sites.¹⁷ Not only was there an excellent correlation between RFLP and serologically determined antigens, but heterogeneity was proven in several DR and DQ specificities, especially HLA-DR6.

One of the novel ideas (and a sign of the future) was the use of 19 base pair oligonucleotides as probes based on sequence information. 18-22 As the probes also hybridised to other genomic sequences, restriction enzyme digestion and gel electrophoresis were required to separate the target sequence from the bulk of the DNA. In addition, due to the small number of available copies of the relevant DNA this approach lacked sensitivity. A refinement of this method was to use the oligonucleotides to probe total RNA, as non-specific binding was not found on Northern blots. 23,24

The advent of DNA techniques highlighted discrepancies in HLA-DR assignment between serological and RFLP methods. ^{25,26} A subsequent report showed that by using RFLP and eliminating these antigen discrepancies from the determination of HLA-DR antigens, the success rate of HLA matched cadaveric transplants at one year (90%) approached that of HLA identical sibling grafts (93%). ²⁷ As a result of this report, many laboratories strove to establish molecular methods for routine typing. The introduction of these methods helped to improve the serological results in laboratories who complemented their serological method with a DNA typing method.

The RFLP methods were not without disadvantages. They did not directly identify the polymorphic coding sequences within the second exon of DRβ, DQβ and DQβ, but relied on polymorphic restriction sites situated generally outside these exons. In addition, they required the use of DR-DQ associations to discriminate between certain DR alleles that had identical DRB RFLP patterns. Thus, care was needed to apply the system to non-Caucasian populations. The method was cumbersome and could take up to 16 days to produce results for only 24 samples. A non-radioactive RFLP method was described²⁸ using digoxigenin and chemiluminescence, but by this time fundamentally different techniques were being developed. Eventually RFLPs were replaced, but not before the results from their

use had acted as a stimulus for the development of better methods for DNA typing.

Availability of Genomic Sequence

As developments were taking place in restriction enzyme methods, several research groups were concentrating on sequencing various class I and II HLA genes. The first HLA class I cDNA clones were obtained in 1980²⁹ and 1981⁶ and cDNA for HLA-DRA³⁰ and DRB^{31,32} genes were isolated in 1982. By 1984 representative genes for all of the classical HLA loci had been identified. Isolation of complete genomic clones led to the nucleotide sequencing of the complete genes encoding HLA-A3 and -Cw3. 33,34 Thereafter much of the sequencing concentrated on the exons. As early as 1984, samples considered to be homozygous by serological methods were found to be heterozygous by sequence analysis.³⁵ By 1986, although only two class II molecules (DRα and DRβ) were sequenced in their entirety, 30 different complete or almost complete amino acid sequences for class II genes were reported. 36 In 1987 a summary of sequence diversity data generated from studies on reference cell lines for the first domain of DR and DQ was reported.37

Polymerase Chain Reaction (PCR)

The advent of PCR^{38,39} was a seminal discovery fundamental to all subsequent techniques that have been developed. PCR had a revolutionary impact on molecular biology research in general and influenced multiple clinical applications. It swiftly became an accepted tool in many disciplines, including HLA. The simple idea of Mullis, which began as an exercise in trying to use oligonucleotides for detection of subtle differences between DNA sequences, has lived up to all its promises.

Emergence of nucleotide sequence data for the alleles of HLA genes permitted the rapid development of many PCR-based techniques and reagents. Conversely, the PCR technique greatly reduced the effort required in subsequent sequencing of new alleles. PCR-based methods may be broadly classified into three categories: (i) those which generate a product containing internally located polymorphisms which can be identified by a second technique, (e.g. PCR-sequence specific oligonucleotide (SSO) probing, PCR-RFLP, PCR followed by sequencing); (ii) those in which the polymorphism

is identified directly as part of the PCR process, although there are post-amplification steps, [e.g. PCR-sequence specific primer (SSP)]; and (iii) confor-mational analysis in which different mutations generate specific conformational changes in PCR products. The latter are identified by electrophoretic analysis e.g. heteroduplex analysis. The two main methods most frequently adopted to clinical histocompatibility have been SSO and SSP, although at regular intervals a novel method or a novel variation of an existing method are reported.

Many factors were found to influence the specificity of the PCR amplification, including sequence of the primer (i.e. length, GC content), free Mg²⁺ concentration, ratio of primer to target, buffer and polymerase concentration.⁴² Conditions must be optimised for each primer pair. It is extremely important to take precautions to prevent contamination of samples with previously amplified DNA.⁴³ Contamination of reagents can be minimised by preparing solutions in facilities which have not been exposed to amplified products, aliquoting reagents for single use and using dedicated equipment and consumables.

The use of thermostable Taq DNA polymerase⁴⁴ meant it was possible to avoid the inactivation of the polymerase, which had necessitated the addition of enzyme after each denaturation step. This development led to the automation of PCR by a variety of simple temperature - cycling devices. Taq polymerase significantly increased the specificity and overall yield of the reaction. In addition to the above, the ability to discard radioactive methods and replace them with detection systems that could be more easily and safely applied, led to the introduction of PCR techniques in many clinical laboratories. The implementation of the salting-out method for DNA extraction⁴⁵ provided a further advantage to the clinical laboratory. This procedure avoided the toxicity of organic solvents and was much cheaper and less time consuming.

Sequence Specific Oligonucleotides (SSO)

The literature contains many alternative methods for SSO typing. The major differences between these are the length and sequence of oligonucleotide probes, and the reporter molecule and its detection. Initially ³²P-labelled allele-specific oligonucleotides were hybridised to an amplified conserved region of

exon 2 of the HLA-DQ α gene,⁴⁶ but soon after biotin was used as a label.⁴⁷ The PCR-SSO method was quickly applied to other loci; DP β ,⁴⁸ DQ β ^{49,50} and DR β ^{51,52} with various procedures using ³²P, biotin or horseradish peroxidase-labeled probes. Methods in the clinical laboratory have tended to use either a substrate in a coloration development system⁵³ or a substrate which generates a chemiluminescent signal.⁵⁴

The SSO method can be customised for each application. For example, approximation of HLA-DR serological specificities requires detection of shared polymorphic sequences which encode the epitopes detected by antibodies. These shared sequences identify families of alleles that belong to the same serologically-defined specificity groups. This level of typing is often referred to as "low resolution" or "generic" SSO. Alternatively, "high resolution" SSO typing can distinguish all known alleles. High resolution SSO usually requires selective amplification of a group of related alleles. For example, all HLA-DRB1*04 alleles are specifically amplified with selected PCR primers and then the DNA is hybridised with a panel of probes which distinguish each HLA-DRB1*04 allele.55

Today, most laboratories prepare one membrane for each probe in the assay. This procedure is facilitated using a 96 well manifold. The use of automation not only eliminates the reuse of membranes after dehybridization, but it also minimizes sample to sample variation in loading and provides a relatively large surface area to aid in evaluation of hybridization dots of varying intensity. Other laboratories have used a robotic work station for both this and the amplification aspect of the technique. 56 Previously, when dot blot manifolds were not available, DNA was directly spotted on to membranes,⁵⁷ transferred from gels using a Southern transfer technique, 58 or spotted using a vacuum dot blot apparatus. Many laboratories reduce the number of different wash temperatures by the addition of tetramethylammonium chloride (TMAC), which reduces the effect of GC content on the stability of the hybrids and, providing the probes are of the same length, enables membranes to be washed at the same temperature. Some laboratories avoid the use of TMAC due to its toxic properties.

The SSO technique has proved very reliable, robust and accurate. Good amplification always gives

a clean and clear cut SSOP hybridisation while almost all the problematic typing results encountered are due to poor amplification. In a quality control exercise the percentage of correct HLA oligotypes was 99.8% for HLA-DR and 99.8% for HLA-DQB1 based on 9,244 HLA-DRB1 and 7,244 HLA-DQB1 assignments.⁵⁹ A recent study using two different SSOP methods in different laboratories found an error rate of 0.1% in testing 2,604 samples for HLA-DP.⁶⁰

DNA typing techniques were initially applied to class II genes rather than class I for several reasons. The requirement for replacing class II serology was thought to be more urgent as serological typing for class II antigens was difficult (HLA-DR and -DQ) or impossible (HLA-DP). [Although Parham made a compelling case that we were deluding ourselves into thinking that serology was good enough for typing HLA-A and -B specificities.⁶¹]. Class II was thought to be more important for transplantation and disease association. Furthermore DNA typing for class I was destined to be more complex because sequence polymorphisms in class I genes are located in two exons. However, once the benefits of developing and applying the methods to class II loci were apparent, attention turned to class I genes. Initially the methods were used to define alleles in selected specificity groups such as HLA-A2, -A68, -A69⁶² or HLA-A2, -A3, -B44,63 alleles of a single specificity e.g. HLA-B27, 64,65 or to identify specificities difficult to detect by serology e.g. HLA-Cw6.66 Two methods for the determination of a complete locus system proved to be the foundation for development of SSO class I methods. 67,68 These methods were later improved by better resolution and methods of probe labelling. 69-74

The biggest problem in class I typing has been the large number of probes required to give unambiguous results for certain heterozygote individuals, i.e. instances where two or more pairs of alleles give the same probe reaction pattern. The choice in SSO for class I has basically come down to whether a laboratory prefers performing one PCR amplification and use a very large number of probes, or use a two-stage typing system with an initial medium resolution followed by high resolution, depending on the results of the first typing. Regardless of which method is used, allele typing is much more difficult for HLA-B than HLA-A, due to the vast polymorphism at HLA-B

Although the SSO technique is perfectly suited for analysing large numbers of samples, it is not suitable for analysing individual or small numbers. The alternative technique of reverse dot blotting uses the same amplification procedure and SSO probes, but the panel of probes is bound to a solid support and the amplified sample is labelled and hybridised to that support. A single hybridization and stringency wash allows the detection of sequence polymorphisms present in the chosen sample. There are several variations of the reverse dot blot. One method uses poly-T tails to attach the oligonucleotide to the membrane, thus increasing the binding efficiency of the oligonucleotide and sparing the specific binding sequence from forming the attachment bonds. The immobilised array of probes hybridises to the PCR product which has incorporated biotinylated primers during the amplification process.⁷⁵ Streptavidin horseradish peroxidase conjugate is added and positive reactions are detected using a coloured substrate. Alternatively visualisation of soluble positive signals is performed by chemiluminescence.⁷⁶

PCR-RFLP

Initially this method used the availability of sites in the nucleotide sequences to employ restriction endonucleases which recognized allelic variations, to digest PCR amplified HLA genes (HLA-DR, -DQ, -DP). 77-79 However, small bands located close to each other on the polyacrylamide gels sometimes obscure precise analysis and some heterozygotes can not be discriminated. 80

These problems have been overcome for HLA-DR by a modified PCR-RFLP method using informative restriction enzymes, which have a single recognition site present in some alleles but not in others and using group specific primers to avoid cross hybridisation with other genes. 81,82 This method was also applied to HLA-DQB1, -DQA1 and -DPB1 genes 83,84 and, in some instances, simultaneous digestion of amplified DNA with two or more enzymes has been applied. 85 One of the first indications that HLA-DP matching may be important in bone marrow transplantation was reported using the PCR-RFLP method.86 A recent innovation was the use of consecutive rounds of PCR-RFLP. After the first digestion of the PCR product, the cleaved fragment was extracted from the gel and used as template for a second PCR-RFLP.87 For the

"allele walking" (the term used by authors to describe this method) to proceed, a previous "cutting" was required.

Sequence Specific Primers (SSP)

In the early 1990's publications appeared on a method called PCR-SSP, ^{88,89} the design of which was based on the amplification refractory mutation system (ARMS). ⁹⁰ The principal of this method is that a completely matched primer will be more efficiently used in the PCR reaction than a primer with one or several mismatches. Specificity is determined by the use of sequence specific primers in which a 3' single-base mismatch inhibits the priming of non-specific reactions. ⁹⁰ Because Taq polymerase lacks 3' to 5' exonuclease activity, even if primer pairs do anneal non-specifically, they will not amplify efficiently. Thus only the desired allele or alleles will be amplified and the amplified product can then be detected by agarose gel electrophoresis.

Other investigators have used multiplex PCR i.e. having several primer pairs in the same reaction. 91,92 Sizing of the PCR product is necessary for interpretation, necessitating that the gel be run longer to separate the PCR fragments. The SSP method is ideal for typing individual samples, but is costly and requires high capacity thermal cyclers for larger numbers of samples. One laboratory reduced this problem by instigating a two-stage technique - low resolution followed by high resolution according to the first result. 93 As the method takes less than five hours, it can be applied to cadaveric transplantation.

Much of the earlier work in class I was performed by Browning and colleagues who developed a low-resolution typing system for HLA-A, ⁹⁴ and quickly followed with a more extensive system to cover all HLA-A alleles. ⁹⁵ The same group designed a low resolution primer panel for HLA-B. ⁹⁶ Other laboratories used the SSP system as a supplement to serological specificities, ⁹⁷ or only determining HLA-B*27. ⁹⁸ Bunce et al. developed a SSP system for HLA-C. ⁹⁹ and a high resolution system for HLA-B. ¹⁰⁰

The products of many alleles of the HLA-C locus are difficult to detect by serological methods due to the low expression of HLA-C molecules at the cell surface and to the corresponding lack of suitable antisera. Thus having a DNA typing system for HLA-C proved very useful. In addition, systems were

developed for other loci which complemented each other, so that complete HLA-A, -B. -C, -DRB1, -DRB3, -DRB4, -DRB5 and -DQB1 typing could be performed simultaneously. This method, termed "phototyping", has a resolution equivalent to high quality serology and could be completed within three hours.

The SSP method suffers from the disadvantage that the end-step of gel electrophoresis is not suitable for large numbers of samples or for automation. A novel method has been reported which removes the electrophoresis and combines high throughput with speed and high resolution. The method uses fluorogenic probes, each of which has a reporter and a quencher dye. When the probe is intact the proximity of the two dyes results in suppression of the reporter fluorescence. During PCR-SSP, if the target of interest is present, the probe specifically anneals between the forward and reverse primer site. The nucleolytic activity of the Taq polymerase cleaves the probe resulting in an increase in fluorescence. Taq polymerase does not cleave free probe - the enzyme requires sequence complementarity between the probe and template for cleavage to occur. After cleavage, the shortened probe dissociates from the target and polymerisation of the strand continues. This process occurs in every cycle and allows direct detection of PCR product. Using the 14 sequence specific primer pairs selected originally by Olerup¹⁰² and three probes, Trucco and his colleagues applied the method to HLA-DQB1 allelic typing. 103 Other investigators have applied this method to class I typing using two different fluorogenic probes and 24, 48 and 16 primer mixes for HLA-A, -B and -C respectively. 104

Heteroduplexes

During the primer-annealing stage of each cycle of the PCR, a proportion of coding strands of each DRB locus allele may hybridise to the noncoding strands of a different DRB locus allele and vice versa. This double stranded DNA will thus be mismatched in some regions (heteroduplexes) leading to alteration in the conformation of the DNA molecule. This conformation varies for each DR haplotype and can be detected by the modified migration polyacrylamide nondenaturing gels, the heteroduplex will move more slowly than the homoduplexes (complementary strands). A single mismatch of nucleotide can cause a marked electrophoretic retardation and thus even subtypes involving a single substitution can be detected. PCR-heteroduplex analysis was first applied to HLA matching in 1990, in an analysis of HLA-DR allotypes encoded by the second exons of HLA-DRB genes. ¹⁰⁵

Additional bands are formed in heterozygotes which are not present in either of the patterns of the individual alleles and are caused by heteroduplex formation in trans i.e. between PCR products from two different haplotypes. This phenomenon was used in the DNA crossmatch test whereby DNA from two different individuals are co-amplified in the PCR. If the individuals are identical for HLA-DR then the banding pattern of the mixture will be the same as the banding pattern of each of the individuals, but if the individuals are not identical the mixture will contain extra bands. This method proved very useful in analysing HLA class I identical individuals for HLA-DR disparity in the selection of unrelated donors to be used for bone marrow transplantation. 106 modification of heteroduplex analysis is the inclusion of third party DNA to generate new heteroduplexes (spiking) which enhance discrimination between otherwise similar patterns. 107 A further refinement the use of temperature gradient gel electrophoresis which analyzes DNA not only on the basis of differential heteroduplex formation between different single strands, but also according to the thermal stability of the homoduplex, which is directly dependent on its nucleotide sequence. 108

In reference strand conformation analysis (RSCA), DNA from a homozygous reference sample is amplified using primers, one of which is fluorescent-labelled at its 5' end. 109 The sample under test is amplified and the PCR product mixed with the reference PCR product to form heteroduplexes. These are resolved in an automated DNA sequencer with only the fluorescent-labelled duplexes being observed and identified according to the distance they have migrated.

Sequence Based Typing (SBT)

One of the drawbacks of SSO or SSP is that, although they are capable of detecting a single base difference in DNA sequence between two alleles, they are not likely to detect a new undefined allele, unless the variation happens to be at the specific site

detected by the probe or the primer. Methods based on sequencing have come to the fore. The sequencing technology advanced with the introduction of dveprimers and fluorescent sequencing.¹¹⁰ Group-specific amplification performed in order to limit the number of allele sequences in any sequencing template, otherwise DNA from both haplotypes would be present. This, in turn, simplifies the software-based allele assignmentssuitable computer software was one of the greatest problems with this technique. Initially in typing for HLA-DR alleles PCR amplification used one set of primers with common sequence to all HLA-DRB1 alleles. However this meant the simultaneous amplification of alleles of other HLA-DRB loci (e.g. DRB3, DRB4) and led to difficulties in assignment of the DRB1 alleles. However by using a series of primers, the sequence of each being specific for some DRB1 alleles, but which do not share sequences with other DRB genes this problem has been nullified. This strategy has also been applied to SSOP typing.

In a recent comparative study between two centres, it was concluded that neither the manufacturer of the DNA sequencer nor the method of sequencing influenced the result. The most critical step was the amplification reaction. On some occasions preferential amplification was observed, whereby one allele was more likely to be amplified than another when they were present in the heterozygote state.

The great advantage of SBT is its accuracy. It is the only technique, which directly detects the nucleotide sequences of an allele, thus allowing an exact assignment. It requires very expensive equipment. Nevertheless, it should be only a matter of time until generally accepted, easy to perform protocols will be available, thus leading to wider use of SBT. The advent of capillary based sequencers has been a tremendous boost in reducing the sophistication of the approaches required in the laboratory; laboratories with no previous experience in sequencing have quickly adapted to this method. It is now feasible to lease the equipment required thus reducing the initial heavy financial outlay. Although it is not always necessary to have resolution to the allele level, the automation and reduction in time required for sequencing has mean that this is easy to obtain.

Choice of Method

The use of a specific technique will depend on the laboratory's requirements. The choice will be influenced by clinical urgency and requirement, sample numbers, availability of equipment, staff skills and budget. Some laboratories, depending on their needs, may use a combination of methods. Because of the ease of storage and transport of DNA samples, or reference cell lines, and the fact that reagents can be made and not continually searched for, as in serology, some laboratories have been able to assist laboratories to set-up the techniques.

According to the clinical application, high or low resolution typing may be required. Kidney transplant candidates, for instance, do not necessarily have to be typed at a high resolution level, because only the 'broad' serological specificities (eg. DR1-DR10) are usually taken into consideration for organ allocation. For unrelated bone marrow transplantation purposes, high resolution typing is required.

Many of the methods referenced in this article only cover polymorphisms for HLA class I occurring in exons 2 and 3 and for class II in exon 2. It should be remembered that some alleles only differ in sequence outside these regions. A list of these alleles can be found on the IMGT/HLA database at http://www.ebi.ac.uk/imgt/hla. At present there are 34 such pairs of alleles (including ten in which the pair of alleles only differ in silent substitutions) in class I, but only two at HLA-DR.

New Alleles

The advent of DNA techniques has led to a massive increase in the number of HLA alleles, either through the detection of alleles not recognised by serological methods or because alleles which differ from one another by one or a few nucleotides can be differentiated, a process not possible by serological methods. In 1992, the number of antigens detected by serology was HLA-A, (25), -B (32), -DR (34), whereas by January 2003 the number of alleles was HLA-A (263), -B. (503), -DR (327). A World Health Organisation committee for HLA nomenclature is responsible for the naming of new alleles and reports of new alleles are furnished monthly in H&I journals. Data is now available on the IMGT/HLA website.

On many occasions laboratories attempt, at great cost, to incorporate the detection of all alleles, even if many are absent in their population. Many alleles are

only present in one individual or his/her family, or may be the product of an erroneous sequencing – several alleles have subsequently been withdrawn after being present on the IMGT/HLA database. In the N.Ireland population, from our results of allele typing 1,000 individuals, we only found 103 of 422 HLA Class I alleles published at the time of testing; 30/90 (33.3%) for HLA-A; 50/254 (19.7%) for HLA-B; 23/78 (29.5%) for HLA-C. 112-114 Bearing in mind the number of new alleles determined since that study, it is likely that less than 15% of HLA Class I alleles identified at present are in this Caucasian population from Northern Europe. One way to alleviate this problem, would be for the laboratory to know the frequency of the alleles in the population under test. This would enable a prediction of the most probable alleles. Recently website www.allelefrequencies.net has been instigated to collect the frequencies of alleles in various populations. This enables an investigator to examine the frequency of all alleles, or a specific allele, in one or many populations. Data will be available, not only for HLA alleles, but for alleles of other polymorphic loci connected to the Histocompa-tibility and Immunogenetics field e.g. cytokine, TAP, MIC, KIR receptors.

Occasional difficulties arise in attempting to link the allele name based on sequence to the serology of the associated antigen. For example most laboratories have assigned the HLA-B7 serological type to the allele HLA-B*2708 which was given that name based on close sequence homology to HLA-B*2701. On other occasions, there is no serological information. Some alleles are hybrids of two "broad" alleles. For example HLA-DR*1107 which at the second exon is composed of two parts, (one from HLA-DRB1*11 and the other from HLA-DRB1*03) has been named because the initial half of the sequence is identical to HLA-DRB1*11 alleles. 115 Knowledge of serological functional behaviour of the allele would be important in such instances. Several alleles have been reported whose products are not expressed. The mutations causing these non-expressions are sometimes located outside the exons normally analysed by DNA methods. Until all these alleles are identified and DNA tests extended to include their polymorphisms, serology will have a role to play. 116-118 Although it must be pointed out that the frequency of these null alleles is very low.

HLA Associated Diseases

DNA techniques have provided a new impetus to the study of HLA and disease. They have removed the imprecise nature of serology allowing associations to be more accurately defined. HLA susceptibility to insulin-dependent diabetes mellitus has now been shown to be related to the presence of non-aspartic acid residues at position 57 on both DQB chains¹¹⁹ and to the presence of arginine at position 52 on the DQα chain. 120 More recent evidence has suggested a combination of DR and DQ molecules is involved. 121 It has been shown that whilst some alleles of HLA-DR4 are associated with susceptibility to rheumatoid arthritis (DRB1*0401, DRB1*0404) others are not (DRB1*0402, DRB1*0403). 122 It is now possible to denote common sequences in the HLA alleles associated with a disease. In rheumatoid arthritis the sequence (QKRRA/QRRAA) within the third hypervariable region of the HLA-DRB1 molecule is present in the alleles associated with this disease. 123 DNA techniques have shown that in nearly every case of coeliac disease the same DOαβ dimer is present in patients, either encoded in cis (on the same haplotype) or in trans (one chain encoded on the maternal haplotype and one chain encoded on the paternal haplotype). 124 The increased knowledge derived from all of the above studies will help to find the mechanisms responsible for the disease.

HLA in Anthropology

The extent of allelic diversity within the HLA complex makes it an especially valuable system for the study of population admixture. The more the HLA system has been studied, the greater the diversity discovered.

Among a group of subtypes, the common pattern is for one subtype to be widely distributed throughout the world's population and the others to be more localized. The sequence of each local subtype is usually identical to the common subtype, except within a short segment. For example, 65 alleles of the HLA-A2 serologically defined antigen have been discovered.

Only two of these are found in a Northern Ireland Caucasian population, with one allele (HLA-A*0201) representing 96% of HLA-A2. This allele also represents more than 90% of HLA-A2 alleles in a Mexican population. However, in a Singapore

Chinese population, only 20% of HLA-A2 alleles are HLA-A*0201, and HLA-A*0207, at 25%, is the most frequent HLA-A2 allele. Thus DNA techniques have been able to show differences in allele frequencies amongst populations which could not be found by serology.

Kidney Transplantation

Mention has previously been made of the beneficial effect on graft survival if HLA-DR typing can be performed more accurately.²⁷ Using serological techniques, it had been impossible to reach any conclusion on the effects of matching for HLA-DQ and -DP in kidney transplantation. With the advent of DNA technology, there has been some indication that matching for HLA-DQ may have some benefit in graft survival, 126,127 but this question remains difficult to answer because of the strong linkage disequilibrium between HLA-DR and -DQ. Initially no effect of matching for HLA-DP alleles was found in transplants from a single centre 127 or in transplants selected from multiple centres with no mismatching at the HLA-A, -B and -DR loci. 128 However an effect of HLA-DP matching in transplants with varying degrees of HLA-A, -B and -DR matching has been reported in second but not first transplants. 129 Although no allowance has been made for linkage disequilibrium, it is known that linkage between HLA-DP and -DR is limited compared to that between HLA-DQ and -DR.

Using serological techniques it was previously shown in kidney cadaveric transplantation that matching for HLA-A and -B splits was more influential than matching for the broad antigens. ¹³⁰ Additionally, a study examined HLA-A and -B specificities using SSO and SSP in kidney cadaveric transplants, that had zero mismatches for HLA-A and -B (by serology) and HLA-DR (by RFLP). ¹³¹ This study did not take the resolution of typing to the allele level, but still found an increase in graft survival of 15% between those transplants with 0mm for HLA-A, and -B after testing with medium resolution SSO and SSP, compared to transplants which were found to have mismatches at either HLA-A or -B by SSO or SSP.

Retrospective studies such as the above are required to determine at what resolution matching should be applied and whether polymorphic sequences at certain parts of the HLA molecule are more important than others. At present in transplantation, a difference of one HLA specificity is called a mismatch. One has only to look at the sequence of alleles to see that DRB1*0101 (HLA-(HLA-DRBR) DR1) and DRB1*0103 significantly fewer amino acid differences than and DRB1*0401 (HLA-DR4). In DRB1*0101 addition, analysis of matching on transplantation success has mainly been performed using serological typing at the broad level whereby specificities such as HLA-B44 and -B45 have been considered as matched. However they are quite different in sequence and, in fact, HLA-B45 is closely related to HLA-B50.

Naturally any structural difference must be shown to be functionally important and the peptide-binding specificity of the allele will need to be considered. ¹³² The more relevant parameter in the choice of a donor may not be the number of mismatches but the kind of mismatch.

Hematopoietic Cell Transplantation

The DNA methods have meant that most laboratories typing for bone marrow transplantation no longer rely on the mixed lymphocyte culture test. In addition, the methods have enabled HLA testing before birth, in order to ascertain if the cord blood would be suitable to use for a sibling requiring a stem cell transplantation. The application of high resolution DNA typing for HLA alleles in bone marrow transplantation has shown that nearly 20% of transplants that were initially typed as identical for HLA-DR by serology were not matched at the allele level, and that there was a significant correlation between DRB1 mismatches and decreased survival. 133 It has already been shown that the one nucleotide difference between HLA-B*0707 and HLA-B*0702 can be recognised by CTLs. 134 This difference is functionally relevant and similar differences have also been shown in other recipient/donor pairs serologically matched for B14, 135 B35, 136,137 B41 138 and B44. 139 With the ability to type to the allele level it will be very difficult for many recipients to receive a transplant from an allelic-matched donor. Hopefully new studies will be able to find the answer on what must be matched for and which mismatches can be ignored.

In the compilation of stem cell donor registries one of the problems has been the recording of donor types obtained by DNA methods when these methods have given alternate types which cross the existing serological boundaries.

A suitable method needs to be found which alleviates this problem and permits existing serological and DNA data to "communicate" with each other. In addition, an SSO probe pattern is interpreted according to the sequence information available at that time. However, as new HLA alleles are found they may have the same probe pattern. Therefore the type assigned to a probe pattern will differ with time. The same problem would also arise with SSP.

As suggested by Hurley, the recording on registries of sequence polymorphisms may go some way toward solving these problems. 140

Future

The chemistry of probe synthesis has evolved to the stage where probes can be synthesised on surfaces such as glass or silicon. The use of large arrays of oligonucleotides on a solid support (DNA chips) which can then be hybridised with a labelled target sequence sequence sequence the result of the needs in other fields for technological improvements in direct sequencing technology and automation.

The tests that become available should have flexibility in their resolution enabling laboratories to economically purchase what they require. For example a medium resolution system would at present be adequate for renal transplantation whereas high resolution to the allele level will probably be required for marrow transplantation.

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