

Review Paper: Application of Hair Follicle Bulge Stem Cells in Wound Healing

Fatemeh Heidari¹ , Maliheh Nobakht², Alireza Shams³, Abazar Yari^{3*} 

1. Department of Anatomy, School of Medicine, Qom University of Medical Sciences, Qom, Iran.
2. Department of Anatomy, School of Medicine, Iran University of Medical Sciences, Tehran, Iran.
3. Department of Anatomy, School of Medicine, Alborz University of Medical Sciences, Karaj, Iran.

Citation: Heidari F, Nobakht M, Shams A, Yari A. Application of Hair Follicle Bulge Stem Cells in Wound Healing. *Anatomical Sciences*. 2017; 14(3):77-88.



Abazar Yari is assistant professor in the Department of Anatomy, Alborz University of Medical Sciences, Karaj. He received PhD of Anatomy from Iran University of Medical Sciences in 2015. His interest is working on wound healing and stem cell transplant.



Funding: See Page 84

Article info:

Received: 10 December 2016

Accepted: 25 March 2017

Available Online: 01 August 2017

Keywords:

Wound healing, Regenerative medicine, Hair Follicle Stem Cell (HFSC), Bulge

ABSTRACT

Despite the significant advances in regenerative medicine, wound healing has remained a challenging clinical problem. Skin is the largest human organ with many vital functions; therefore, any damage to its normal structure should be treated as soon as possible. Easy access to skin stem cells has created a lot of excitement in therapeutic applications. "Cell therapy" is considered a novel method in regenerative medicine, especially when conventional treatments fail. Candidate cell populations for therapeutic applications include embryonic, induced pluripotent, adult mesenchymal, and hair follicle stem cells. It is possible to differentiate stem cells separated from the bulge area of hair follicle into neurons, melanocytes, keratinocytes, glia and smooth muscle cells that are negative for the keratinocyte marker *kr15*.

This review discusses the plasticity of skin stem cells, especially stem cells located in the hair follicle and their involvement in wound healing, gene expression profile in wound healing, hair follicle stem cells, and their surrounding epidermis. Moreover, the ability of hair follicle stem cells for treating wounds and regenerative medicine is going to be discussed. Eventually we suggest the hair follicle as an ideal source of stem cells for cell therapy and regenerative medicine because they are abundant with easy access and great differentiation ability.

* Corresponding Author:

Abazar Yari, PhD

Address: Department of Anatomy, School of Medicine, Alborz University of Medical Sciences, Karaj, Iran.

Tel: +98 (26) 34349802

E-mail: yari@abzums.ac.ir

1. Introduction

This study reviews the latest advances in the field of stem cells and their potential uses in cell therapy with special focus on wound healing. Despite the advances in regenerative medicine, wound healing has remained a challenging medical problem, mainly following operation and severe damage of the skin that increases the risk of infections, postoperative hospitalizations [1, 2], early and late complications, and eventually morbidity and mortality rate [3].

Skin is the largest human organ with many important functions including thermoregulation, sweat production, and protection against pathogens; thus, any damage to its normal structure should be cured as soon as possible [4]. The skin structure comprised two layers (dermis and epidermis) that are distinct functionally, anatomically, and developmentally [5].

The term “wound” can be defined as any disruption in the normal structure of the skin resulting in loss of its normal function [6]. Current therapies including surgery, wound bandage, common negative pressure, and substitutes of skin

are not sufficient in all conditions and new methods should be introduced in the field of wound healing as soon as possible [3, 7, 8]. Wound healing process included three phases (inflammation, proliferation, maturation or remodeling) which overlap in the time and space [9]. Common wound curing is a complex mechanism, required coordinated interactions between different biological and immunological systems at many different levels such as molecular mediators, cells, and structural elements [10, 11].

Conventional treatments of wounds is not effective in all conditions so modern approaches should be developed. In this regard, growth factors and cell therapy have been widely used [12]. Based on the reports, the pluripotent nestin-positive, keratin (K15)-negative stem cells in the hair follicle of mouse can differentiate into glia, neurons, keratinocytes, melanocytes and smooth muscle cells. Nestin-expressing cells differentiate into neuronal and glial cells following transplantation to the damaged spinal cord and help in repairing injury and recovery of locomotor system. The nestin-expressing pluripotent stem cells from the dermal papilla are called Skin-derived Precursor (SKP) cells. These outcomes propose that stem cells of hair-follicle in bulge-area can be considered as an autologous and accessible source for multipotent stem cells to treat wounds.

Table 1. Cytokines and growth factors that accelerate keratinocyte proliferation and differentiation

Cytokines/Growth Factors	Key References
Epidermal growth factor	[13-15]
Insulin-like growth factor	[15, 16]
Epiregulin	[17]
Fibroblast growth factor	[18]
Transforming growth factor	[19]
Keratinocyte growth factor	[20]
Stem cell factor	[21]
Bone morphogenetic protein	[22]
Angiopoietin-related growth factor	[23]
Growth differentiation factor-5	[24]
Granulocyte/macrophage colony-stimulating factor	[25]
Tumor necrosis factor-a	[26]
Interleukin-1	[27]
Thymocyte-activating factor	[28]
Nerve growth factor	[29]

2. Growth Factors and Cytokines Involved in Wound Healing

Many studies demonstrated that keratinocyte proliferation and differentiation play a major role in wound healing. Many growth factors and cytokines are involved in this process. Here we have summarized the results in Table 1.

3. Cell Therapy and Regenerative Medicine

Growth factors, on their own are not effective in wound treatment [12]. Nowadays, researchers have focused on the application of cell therapy for treating several pathologies such as wound healing [30, 31]. Cell therapy is considered as a new method in regenerative medicine, especially when conventional treatments fail. Cell therapy involves applying live stem cells to repair or restore the function of a damaged tissue [32, 33]. Therefore, the application of stem cells in cell therapy are investigated in numerous fields of regenerative treatment.

4. History of Stem Cells Therapies for Wound Healing

The term “stem cells” was first defined at the end of the 19th century as a theoretical assumption for self-renewing ability of the certain tissues (skin, blood, etc.) for the lifetime of an organism while they are made of short-lived cells. Many years later and following advancements in isolation of stem cell candidates, along with their potential testing after transplantation in vivo models, stem cells were identified as distinct cellular population. In 1966, Friedenstein et al. first isolated bone marrow-derived Mesenchymal Stem Cells (MSCs) [34], from humans by aspiration from the iliac crest. Later, the cells were extended in culture and topically applied to wounds to improve repairing tissue.

5. Stem Cells and Regenerative Medicine

By definition, stem cell are specific cell types that are self-healing and can proliferate and differentiate into other cell lines [35]. So far, the suitable source of stem cells has remained a major challenge, because their underlying mechanism is not completely understood [36]. Stem cells are able not only to produce cell types of their own tissue, but also cell types presenting in other tissues [37]. Cell therapy employs embryonic [38] or adult stem cells [39] to reconstruct injured tissues. Applications of embryonic stem cells has risen ethical concerns. The differentiation ability of adult stem cells has provided a chance for researchers to apply them in regenerative medicine [36]. Obtaining adult stem cells is possible from numerous organs of the body [40]. More studies

are needed to focus on the function of stem cells in wound treatment to improve their efficacy (Table 3) [9].

Stem cells are undifferentiated cell populations which have capacity to differentiate into progenitor, or precursor cells of other cell types [37]. Adult stem cells are frequently used in therapeutic applications; some characteristics of adult stem cells have been mentioned in some research studies. It is simple to experiment on and culture them in vitro and it does not create moral concerns. Such stem cells as embryonic cells with high proliferative ability and differentiation potential can be collected from almost any tissue [41]. Table 2 lists some stem cells found in the hair follicle and surrounding skin.

6. Stem Cell Populations for Cutaneous Repair

Numerous sources of stem cells have been tested in preclinical and clinical settings for their potential to improve wound treatment. They have shown positive clinical outcomes and demonstrated autologous stem cell therapy to be safely tolerated. For example, Lee et al. showed that application of adipose tissue-derived stem cell implanting in a patient suffering from critical limb ischemia, could decrease the frequency of minor limb amputation, pain rating scales, and significantly improve claudication walking distance [58, 65].

Therapeutic capacity of the most stem cell populations including Embryonic Stem Cells (ESCs), induced Pluripotent Stem Cells (iPSCs), hair follicle bulge stem cells and adult MSCs, for wound healing have already been tested [61, 66].

7. Clinical Applications of Embryonic and Induced Pluripotent Stem Cells

Embryonic stem cells are pluripotent cells. They result from the internal cell mass corresponding to the blastocyst and have capacity to form a complete organism [67]. In an effort to use ESCs for repairing cutaneous tissue, they were differentiated by Guenou et al. into functional keratinocytes and applied for regeneration of the epidermis [61]. However, current clinical application of ESCs because of the possibility of immunogenicity, tumorigenicity, and ethical controversy is not promising [68].

Induced Pluripotent Stem Cell (iPSC) is a novel source of stem cells that possibly has the benefits of ESCs. These cells were generated by Takahashi and Yamanaka by reprogramming of adult fibroblasts into pluripotent immature state [66]. The iPSC technology allows to create populations of autologous pluripotent stem cell from adult differentiated cells. iPSCs are autologous and non-immunogenic, so spare the ethical issues associated with human ESCs. In vitro, 3-D

Table 2. Stem cells in the hair follicle and surrounding skin

Stem Cells	Location	Derivative Cell Types	References
Stem cells of dermal-sheath	Hair follicle dermal sheath	Dermal papilla cells and wound treating fibroblasts	[42, 43]
Stem cells of epidermis	Basal layer of the epidermis	Transient intensifying cells and terminal-differentiated epidermal cells	[44]
Stem cells of endothelium	Dermis	Endothelial cells	[45]
Stem cells of follicle multipotent	Bulge region of hair follicle	Hair follicle epithelium (containing external root sheath, inside root sheath, hair shaft, etc.), sebaceous gland cells and epidermal cells	[46-48]
Stem cells of hematopoietic tissue	Dermal papillae of hair follicle	All erythroid and myeloid lineages	[49, 50]
Stem cells of neural-crest	Dermal papillae of hair follicle	All neural cell types and a number of mesodermal derivatives	[51]
Stem cells of melanocyte	Bulge region of hair follicle	Melanocytes	[52]
Mesenchymal stem-cell-like cells	Dermis	Mesodermal derivatives and several neural cell types	[53]

ANATOMICAL SCIENCES

skin equivalents were generated by Itoh et al. [63] which composed mainly of human iPSC-derived keratinocytes and fibroblasts. Differentiation of iPSCs into folliculogenic human epithelial stem cells was demonstrated by Yang et al. [69] that restored all parts of the hair follicle. These results could improve the iPSC-based generation corresponding to full cutaneous equivalents, including epidermal appendages for wound healing.

iPSCs show the hybrid benefits of the ESCs and MSCs. Though, there are still many risks, including their tumorigenicity in an undifferentiated state, which must be solved prior to extensive clinical application. In spite of advantages of iPSC-based therapies in wound healing, it is essential to raise their safety ethics and improve the current approaches for their differentiating into keratinocytes, fibroblasts, and

related cells in the wound bed with a focus on its cost-effectiveness and efficiency [70].

8. Clinical Applications of Mesenchymal Stem Cells (MSCs) in Wound Healing

The clinical usefulness of MSCs in wound treatment has already been published in several studies. The most used source of adult stem cells is Bone Marrow (BM). Several studies demonstrated that cells collected from the BM contribute in regenerating or repairing numerous tissues, comprising the bone, myocardium, cartilage, tendons, and skin [71].

BM contains a variety of heterogeneous cell populations, such as adipocytes, fibroblasts [72], Mesenchymal Stem Cells (MSCs), and Hematopoietic Stem Cells (HSCs). Both

Table 3. Summary of cells applied in wound healing

Cell Type	References
BM-MNC* BM-MSc	[31, 54-57]
ASC	[12, 58-60]
ESCs	[61]
iPSC	[62-64]

* BM-MNC: Bone Marrow Mononuclear Cells

ANATOMICAL SCIENCES

MSCs and HSCs have a great potential of plasticity, and can contribute into hematopoietic and non-hematopoietic tissues [73]. Both MSCs and HSCs mobilize from the BM to the wound site, once a wound occurs. In the wound place, they control proliferation and migration of cells in the inflammation phase [74]. MSCs produce several growth factors which induce dermal fibroblast proliferation, angiogenesis and also collagen deposition [75]. Recent studies demonstrated that BM stem cells play an important role in skin regeneration and vascularization [60]. Furthermore, MSCs have antimicrobial activity [76].

Clinical uses of BM derived MSCs (BM-MSCs) in wound treatment have been documented. Badivas et al. demonstrated that injecting BM directly into the wound edges followed by using cultured MSCs, lead to full closure of the wound and tissue reconstruction in 3 patients with chronic ulcers where traditional treatment regimens had failed [54]. In 2009, Dash et al. showed the effect of intramuscular application of autologous BM-MSCs at the edges of the wound in 24 subjects with wounds in the lower edges because of vasculitis or diabetes. The results demonstrated that implanting autologous BM-MSCs in non-treating wounds improves clinical parameters and accelerates the healing process significantly [57]. In systemic administration of MSCs carried out by Lu et al. on diabetic subjects suffering from lower limb ischemia, the pain relief and increase in treatment rate of the wound was significant [77]. Remarkable improvement in the regeneration of the dermis has been achieved through transplanting BM-MSCs. In fact, application of these cells with stem cell properties might be helpful in wound healing and reducing scar formation [78].

During the repair process, cells extracted from BM can be an important source of endothelial progenitor cells and inflammatory cells to repair cutaneous wounds [79], moreover, the contribution of BM-MSCs derived fibroblasts and endothelial cells in wound healing has been documented. In these cases, circulating fibrocytes (contributing to the myofibroblast population) have an important role in wound closure [80]. There is a wealth of evidence supporting the existence of BM-derived epithelial cells in the tissues of skin [71]; BM cells can form epidermal keratinocytes [81, 82]. These outcomes also confirm the application of BM-derived cells in epidermal healing and repair. Further study suggests that BM-derived cells are able to constitute the functional skin cells and restore cutaneous tissue [71, 82].

9. Medical Applications of Adipose-Tissue-Derived MSCs (ASCs) in Wound Treatment

As mentioned before, it is possible to obtain MSCs from diverse tissues but harvesting them is invasive and painful

[83]; thus, Zuk and associates characterized and defined adipose tissue derived MSCs (ASCs) from lipoaspirates, in 2001 [84].

ASCs secrete different cytokines and growth factors similar to those released by BM-derived MSCs [85]. These properties make ASCs a common source for cell treatment, and currently they are applied in diverse clinical treatment, such as wound healing [12, 41, 86, 87]. In 2012, Lee et al. reported the result of intramuscular use of ASC in 15 patients with critical limb ischemia. In their findings, decrease in the pain rating scale and improve in claudication walking distance was significant. This study concluded that ASC application may be a safe method to attain healing angiogenesis in subjects suffering from critical limb ischemia where other treatment modalities failed [58].

10. Plasticity of Skin Stem Cells

Plasticity is a concept to define the multipotency characteristic of adult cell populations. This means that adult cells have the capacity to transdifferentiate and or be reprogramed and fuse, or persist corresponding to multipotential stem cells of adult tissues and organs [88, 89].

Skin cell plasticity has many therapeutic advantages. There are some multipotent stem cells in skin; epidermal stem cells in the epidermis basal layer, mesenchymal stem cell like cells and skin-derived progenitor cells in the dermis [53], and Hair Follicle Stem Cells (HFSCs). These cells like BM-MSCs are able to differentiate into mesodermal derivatives and neural cell types; therefore, they could be a source for fibroblasts that are essential in wound treatment events [78, 90].

Epidermal stem cells of adult human, in both the hair follicle and the epidermis have a high expansion capacity, and can form colonies in vitro: studies reported the successful transplantation of cultured epithelia for large and deep burn wounds, especially if this method was mixed with fibrin matrices to ease the application corresponding to epidermal stem cells [91].

11. Stem Cells Located in Hair Follicle

Recent developments have been made in locating and detecting diverse adult stem cell progenies in the skin and hair follicle, for example, epidermal stem cells located in the basal layer of epidermis. They are unipotent and provide regeneration of the epidermis in adult skin [44]. These unipotent stem cells arise from multipotent stem cells, located in the bulge of hair follicles. Researches

show that bulge of hair follicle is an important place for multipotent stem cells [47, 92].

Subgroups of these multipotent stem cells are able to migrate out of the hair follicles to repair the damaged epithelium in the wounded site; but they little participate in the intact epidermal layer. Moreover, these hair-follicle-derived stem cells are able to participate in the follicles reconstruction (comprising the external root-sheath, internal root-sheath and hair shaft) and also the sebaceous gland. Furthermore, melanocyte stem cells are also present in the hair follicle bulge region [52].

Hair follicle dermal cells also contain hematopoietic cell population, which are CD 45-positive, and mesenchymal stem cell population [50, 93]. Follicle dermal cells have capacity to generate hematopoietic cells and could reconstitute the hematopoietic system in lethally irradiated mice [49]. The human scalp hair follicle dermal sheath can induce the formation of hair follicles, form new dermal papilla, and when transplanted onto the skin, can produce hair shafts [94]. In the case of implanting the follicle dermal cells into skin wounds, they have the ability to reconstitute the fresh dermis like skin wound-healing fibroblasts [43].

According to the available evidence, HFSCs are a source for dermal and epidermal cell populations [95]. In allograft transplantation of dermal sheath cells from one individual to another, they generate follicles that result in normal growth of hair with no rejection [94]; thus the immune privilege of such stem cells make them universal donors in cell-based therapy applications and also suitable targets [96]. Skin cells are largely available, especially hair follicles. They can be exploited as unique populations, a perfect source of autologous or allogeneic applications. The hair follicle displays immune privilege consequence of a unique immunological profile: having no expression for MHC class I and low amount of hair follicle immune cells [96].

12. Hair Follicle Bulge Stem Cells

The accessibility of stem cell sources in therapeutics approaches is a key point. Skin stem cells ease of access created much excitement in therapeutic applications. Skin stem cells provide hope to induce adult wound healing like embryonic ones, with fast regeneration, no scarring, and full reconstitution of hair and glands [79]. Moreover stem cells of skin have the therapeutic potential for treating wounds and diseases in other tissues [95].

Stem cells of the hair follicle bulge of adults is one of the candidate sources for regenerative drugs [97]. The hair follicle reconstructs itself via the cycle comprising three phas-

es of anagen (growing phase), catagen (regression phase), and telogen (resting phase), all proposing the existence of its own stem cells [43, 92]. Some of HFSCs are located in the bulge region, which is between the arrector pili muscle insertion and the sebaceous gland duct [98].

13. Bulge Stem Cells and Wound Healing

Recent studies show that cell therapy with endogenous stem cell populations placed in the bulge region of hair follicle is a suitable method in coetaneous wound treatment [99].

Hair Follicle bulge Stem Cells (HFSCs) are appropriate source for pluripotent adult stem cells used in regenerative medicine because these cells are available, can be cultured easily, and unlike embryonic and fetal stem cells, are not associated with ethical issues. Also they prevent additional surgery complications [46, 100, 101].

Recent research demonstrated that population of stem cell in bulge region has positive expression of nestin, neural stem cell marker, and could differentiate into keratinocytes, neurons, glia, melanocytes, smooth muscle cells, and blood vessels [102-105]. These stem cells may be an accessible, autologous source and have high therapeutic value in regenerative medicine. Also their application does not raise any immunological problems or ethical concerns [106].

14. Wound Healing and Hair Follicle Gene Expression

Following full-thickness wounds, cells from the Interfollicular Epidermis (IFE) and the hair follicles migrate into the damaged area [48, 107]. Studies demonstrate that delayed wound healing occurs in mice lacking hair follicles [108]. That means that few hair follicle derived cells are involved in the re-epithelialization of epidermis in full-thickness wounds. Lineage detecting of Lrig1+ cells top of the bulge and Gli1+ cells in the junctional area indicate that hair follicle fate is able to persist for longer time [109, 110] but following repair, these hair follicles offspring are mainly substituted with epidermal progeny [111]. Thus, Interfollicular Epidermis (IFE) plays a major role in wound re-epithelialization rather compared to hair follicles. Overall, the source of new hair follicle placodes in repaired skin are not Keratin 15+ bulge stem cells from normal periwound hair follicles [112], but other populations of hair follicles stem cells which are Lrig+ and Lgr6+ stem cells [113].

Gene expression in hair follicle stem cells and its surrounding epidermis Skin epithelia display distinct populations of stem cell in hair follicles and epidermis. Epithelial

Table 4. Distribution of different genes in Hair follicle stem cell

Marker	Location	References
CD34+	Bulge	[116]
CD200+	Bulge	[117, 118]
K15+	Bulge	[119]
Sox9+	Bulge	[115]
Lhx2+	Bulge	[115]
Tcf3	Bulge	[115]
Nfatc1	Bulge	[115]
Lgr1+	Isthmus	[120]
Plet1+	Isthmus	[115]
Gli1+	Isthmus	[115]
Lgr6+	Isthmus	[121]
Blimp1+	SG	[122]
Sca1+	Infundibulum	[123]
Lrig1+	Junctional zone	[115]
CD200+	Secondary hair germ	[115]
K15+	Secondary hair germ	[115]
Gli1+	Secondary hair germ	[115]
Lgr5+	Secondary hair germ	[115]

ANATOMICAL SCIENCES

stem cells have diverse lineage capacity in different locations [114]. Physiologically, stem cells in the follicular infundibulum and interfollicular epidermis are limited to the fate of epidermal cells.

Stem cells of interfollicular epidermal can be offered as slow-cycling populations in label retaining research, but specific indicators are unknown. Hair follicles have several specific epithelial cell populations in the isthmus and junctional area that are Lrig1+ (yellow), Lgr6+, and Gli1+ stem cells (green), which preserve the isthmus and contribute to infundibulum, sebaceous gland, and to interfollicular epidermis. Progenitors of sebaceous gland are unipotent and identified by Blimp [1] (orange). The stem cells of bulge (blue) normally reconstitute all lineages of hair follicle and can be detected with the expression of CD34, Krt15, Sox9, CD200, Lhx2, Lgr5, Nfatc1, and Tcf3. The telogen hair follicles secondary germ (purple) express CD200, Gli1 and Lgr5 (Table 4) [115].

15. Conclusion

Any damage to the normal anatomical structure of the skin resulting disrupts its normal function and can be defined as wound. Despite the advances in regenerative medicine, wound healing has still remained a challenging clinical problem. A new field of medicine is cell therapy which applies embryonic or adult stem cells to reconstruct injured tissues. Numerous sources of stem cells are being studied in clinical treatment for their potential to improve wound treatment and autologous stem cell therapy to be safely tolerated. The easy access of stem cell sources is a key point in therapeutics approaches.

While the wound healing with stem cells has shown promising results, identifying the cells that have the most beneficial effect will be an effective approach. For this purpose, the hair follicle stem cells especially in bulge area are a suitable source, because they can differentiate into different lineages with no expression of MHC class I. Therefore, the

immune privilege of these stem cells make them universal donors in cell-based therapy. Given that in normal mode, stem cells of hair follicle are involved in wound treatment, the similar studies about the bulge area of hair follicle and wound healing may lead to novel techniques for application of stem cells in clinical treatment of wound healing.

Ethical Considerations

Compliance with ethical guidelines

There is no ethical principle to be considered in this paper.

Funding

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Authors contributions

All authors have read and approved the manuscript.

Conflict of interest

The authors declared no conflict of interest.

References:

- [1] Smith MA, Dahlen NR. Clinical practice guideline surgical site infection prevention. *Orthopedic Nursing*. 2013; 32(5):242-8. [DOI:10.1097/NOR.0b013e3182a39c6b]
- [2] Smith RL, Bohl JK, McElearney ST, Friel CM, Barclay MM, Sawyer RG, et al. Wound infection after elective colorectal resection. *Annals of Surgery*. 2004; 239(5):599-605.
- [3] Natarajan S, Williamson D, Stiltz AJ, Harding K. Advances in wound care and healing technology. *American Journal of Clinical Dermatology*. 2000; 1(5):269-75. [DOI:10.2165/00128071-200001050-00002] [PMID]
- [4] Hanson SE, Bentz ML, Hematti P. Mesenchymal stem cell therapy for nonhealing cutaneous wounds. *Plastic and Reconstructive Surgery*. 2010; 125(2):510-6. [DOI:10.1097/PRS.0b013e3181c722bb] [PMID] [PMCID]
- [5] Lazic T, Falanga V. Bioengineered skin constructs and their use in wound healing. *Plastic and Reconstructive Surgery*. 2011; 127, 75S-90S. [DOI:10.1097/PRS.0b013e3182009d9f] [PMID]
- [6] Lazarus GS, Cooper DM, Knighton DR, Percoraro RE, Rodeheaver G, Robson MC. Definitions and guidelines for assessment of wounds and evaluation of healing. *Wound Repair and Regeneration*. 1994; 2(3):165-70.
- [7] de Laat EH, van den Boogaard MH, Spauwen PH, van Kuppevelt DH, van Goor H, Schoonhoven L. Faster wound healing with topical negative pressure therapy in difficult-to-heal wounds: a prospective randomized controlled trial. *Annals of Plastic Surgery*. 2011; 67(6):626-31. [DOI:10.1097/SAP.0b013e31820b3ac1] [PMID]
- [8] Lutz NW, Confort Gouny S, Casanova D, Andrac Meyer L, Magalon G, Cozzone PJ. Conditions of wound healing and cutaneous growth affect metabolic performance of skin following plastic surgery. *Wound Repair and Regeneration*. 2007; 15(4):491-6.
- [9] Teng M, Huang Y, Zhang H. Application of stems cells in wound healing-an update. *Wound Repair and Regeneration*. 2014; 22(2):151-60.
- [10] Velnar T, Bailey T, Smrkolj V. The wound healing process: An overview of the cellular and molecular mechanisms. *The Journal of International Medical Research*. 2009; 37(5):1528-42. [DOI:10.1177/147323000903700531] [PMID]
- [11] Chodorowska G, Rogus Skorupska D. Cutaneous wound healing. *Annales Universitatis Mariae Curie-Skłodowska*. 2004; 59(2):403-7. [PMID]
- [12] Cherubino M, Rubin JP, Miljkovic N, Kelmendi Doko A, Marra KG. Adipose-derived stem cells for wound healing applications. *Annals of Plastic Surgery*. 2011; 66(2):210-5. [DOI:10.1097/SAP.0b013e3181e6d06c] [PMID]
- [13] Ristow HJ. Studies on stimulation of DNA synthesis with epidermal growth factor and insulin-like growth factor-I in cultured human keratinocytes. *Growth Regulation*. 1996; 6(2):96-109. [PMID]
- [14] Kwon YB, Kim HW, Roh DH, Yoon SY, Baek RM, Kim JY, et al. Topical application of epidermal growth factor accelerates wound healing by myofibroblast proliferation and collagen synthesis in rat. *Journal of Veterinary Science*. 2006; 7(2):105-9. [DOI:10.4142/jvs.2006.7.2.105] [PMID] [PMCID]
- [15] Bhora FY, Dunkin BJ, Batzri S, Aly HM, Bass BL, Sidawy AN, et al. Effect of growth factors on cell proliferation and epithelialization in human skin. *The Journal of Surgical Research*. 1995; 59(2):236-44. [DOI:10.1006/jsre.1995.1160] [PMID]
- [16] Kamalati T, Howard M, Brooks RF. IGF I induces differentiation in a transformed human keratinocyte line. *Journal of Biological Chemistry*. 1989; 106(2):283-93. [PMID]
- [17] Shirakata Y, Komurasaki T, Toyoda H, Hanakawa Y, Yamasaki K, Tokumaru Sh et al. Epiregulin, a novel member of the epidermal growth factor family, is an autocrine growth factor in normal human keratinocytes. *The Journal of Biological Chemistry*. 2000; 275(8):5748-53. [DOI:10.1074/jbc.275.8.5748] [PMID]
- [18] Song YH, Zhu YT, Ding J, Zhou FY, Xue JX, Jung JH, et al. Distribution of fibroblast growth factors and their roles in skin fibroblast cell migration. *Molecular Medicine Reports*. 2016; 14(4):3336-42. [DOI:10.3892/mmr.2016.5646]
- [19] Wang G, Higgins PJ, Gannon M, Staiano Coico L. Transforming growth factor-beta 1 acts cooperatively with sodium n-butyrate to induce differentiation of normal human keratinocytes. *Experimental Cell Research*. 1992; 198(1):27-30. [DOI:10.1016/0014-4827(92)90144-W]
- [20] Chomiski V, Gragnani A, Bonucci J, Correa SA, Noronha SM, Ferreira LM. Keratinocyte growth factor and the expression of wound-healing-related genes in primary human keratinocytes from burn patients. *Acta Cirurgica Brasileira*. 2016; 31(8):505-12.

- [21] Grabbe J, Welker P, Rosenbach T, Nürnberg W, Krüger Krasagakes S, Artuc M, et al. Release of stem cell factor from a human keratinocyte line, HaCaT, is increased in differentiating versus proliferating cells. *The Journal of Investigative Dermatology*. 1996; 107(2):219-24. [DOI:10.1111/1523-1747.ep12329664] [PMID]
- [22] D'Souza SJ, Pajak A, Balazsi K, Dagnino L. Ca²⁺ and BMP-6 signaling regulate E2F during epidermal keratinocyte differentiation. *The Journal of Biological Chemistry*. 2001; 276(26):23531-8. [DOI:10.1074/jbc.M100780200] [PMID]
- [23] Oike Y, Yasunaga K, Ito Y, Matsumoto SI, Maekawa H, Morisada T, et al. Angiopoietin-related Growth Factor (AGF) promotes epidermal proliferation, remodeling, and regeneration. *Proceedings of the National Academy of Sciences of the United States of America*. 2003; 100(16):9494-9. [DOI:10.1073/pnas.1531901100] [PMID] [PMCID]
- [24] Kim DS, Korting HC, Schafer Korting M. Effects of growth factors on the proliferation of human keratinocytes and fibroblasts in vitro. *Die Pharmazie*. 1998; 53(1):51-7. [PMID]
- [25] Braunstein S, Kaplan G, Gottlieb AB, Schwartz, M., Walsh, G., Abalos, R.M., et al. GM-CSF activates regenerative epidermal growth and stimulates keratinocyte proliferation in human skin in vivo. *The Journal of Investigative Dermatology*. 1994; 103(4):601-4. [DOI:10.1111/1523-1747.ep12396936] [PMID]
- [26] Bikle DD, Pillai S, Gee E, Hincenbergs M. Tumor necrosis factor- α regulation of 1, 25-dihydroxyvitamin D production by human keratinocytes. *Endocrinology*. 1991; 129(1):33-8. [DOI:10.1210/endo-129-1-33] [PMID]
- [27] Maas Szabowski N, Stark HJ, Fusenig NE. Keratinocyte growth regulation in defined organotypic cultures through IL-1-induced keratinocyte growth factor expression in resting fibroblasts. *The Journal of Investigative Dermatology*. 2000; 114(6):1075-84. [DOI:10.1046/j.1523-1747.2000.00987.x] [PMID]
- [28] Al Refu K, Edward S, Ingham E, Goodfield M. Expression of hair follicle stem cells detected by cytokeratin 15 stain: Implications for pathogenesis of the scarring process in cutaneous lupus erythematosus. *The British Journal of Dermatology*. 2009; 160(6):1188-96. [DOI:10.1111/j.1365-2133.2009.09074.x] [PMID]
- [29] Di Marco E, Mathor M, Bondanza S, Cutuli N, Marchisio PC, Cancedda R, et al. Nerve growth factor binds to normal human keratinocytes through high and low affinity receptors and stimulates their growth by a novel autocrine loop. *The Journal Of Biological Chemistry*. 1993; 268(30):22838-46. [PMID]
- [30] Darkazalli A, Ismail AA, Abad N, Grant SC, Levenson CW. Use of human mesenchymal stem cell treatment to prevent anhedonia in a rat model of traumatic brain injury. *Restorative Neurology and Neuroscience*. 2016; 34(3):433-41. [DOI:10.3233/RNN-150628] [PMID]
- [31] Garcia Gomez I, Elvira G, Zapata AG, Lamana ML, Ramirez M, Garcia Castro J, et al. Mesenchymal stem cells: biological properties and clinical applications. *Expert Opinion on Biological Therapy*. 2010; 10(10):1453-68. [DOI:10.1517/14712598.2010.519333] [PMID]
- [32] Watt FM, Hogan BL. Out of eden: Stem cells and their niches. *Science*. 2000; 287(5457):1427-30. [DOI:10.1126/science.287.5457.1427] [PMID]
- [33] Weissman IL. Stem cells: Units of development, units of regeneration, and units in evolution. *Cell*. 2000; 100(1):157-68. [DOI:10.1016/S0092-8674(00)81692-X]
- [34] Friedenstien AJ, Piatetzky-Shapiro II, Petrakova KV. Osteogenesis in transplants of bone marrow cells. *Journal of Embryology and Experimental Morphology*. 1966; 16(3):381-390. [PMID]
- [35] Levy YS, Stroomza M, Melamed E, Offen D. Embryonic and adult stem cells as a source for cell therapy in Parkinson's disease. *Journal of Molecular Neuroscience*. 2004; 24(3):353-86. [DOI:10.1385/JMN:24:3:353]
- [36] Amoh Y, Li L, Campillo R, Kawahara K, Katsuoka K, Penman S, et al. Implanted hair follicle stem cells form Schwann cells that support repair of severed peripheral nerves. *Proceedings of the National Academy of Sciences of the United States of America*. 2005; 102(49):17734-8. [DOI:10.1073/pnas.0508440102] [PMID] [PMCID]
- [37] Barry FP, Murphy JM. Mesenchymal stem cells: Clinical applications and biological characterization. *The International Journal of Biochemistry & Cell Biology*. 2004; 36(4):568-84. [DOI:10.1016/j.biocel.2003.11.001] [PMID]
- [38] Gerecht Nir S, Itskovitz Eldor J. Cell therapy using human embryonic stem cells. *Transplant Immunology*. 2004; 12(3-4):203-9. [DOI:10.1016/j.trim.2003.12.013] [PMID]
- [39] Gerlach JC, Zeilinger K. Adult stem cell technology-prospects for cell based therapy in regenerative medicine. *The International Journal of Artificial Organs*. 2002; 25(2):83-90. [DOI:10.1177/039139880202500202] [PMID]
- [40] Neuss S, Becher E, Woltje M, Tietze L, Jahnen Dechent W. Functional expression of HGF and HGF receptor/c-met in adult human mesenchymal stem cells suggests a role in cell mobilization, tissue repair, and wound healing. *Stem Cells*. 2004; 22(3):405-14. [DOI:10.1634/stemcells.22-3-405] [PMID]
- [41] Strioga M, Viswanathan S, Darinskas A, Slaby O, Michalek J. Same or not the same? Comparison of adipose tissue-derived versus bone marrow-derived mesenchymal stem and stromal cells. *Stem Cells and Development*. 2012; 21(14):2724-52. [DOI:10.1089/scd.2011.0722] [PMID]
- [42] Jahoda CA, Reynolds AJ. Hair follicle dermal sheath cells: Unsung participants in wound healing. *Lancet*. 2001; 358(9291):1445-8. [DOI:10.1016/S0140-6736(01)06532-1]
- [43] Gharzi A, Reynolds AJ, Jahoda CA. Plasticity of hair follicle dermal cells in wound healing and induction. *Experimental Dermatology*. 2003; 12(2):126-136. [DOI:10.1034/j.1600-0625.2003.00106.x] [PMID]
- [44] Morasso MI, Tomic Canic M. Epidermal stem cells: The cradle of epidermal determination, differentiation and wound healing. *Biology of the Cell*. 2005; 97(3):173-83. [DOI:10.1042/BC20040098] [PMID] [PMCID]
- [45] Brouard M, Barrandon Y. Controlling skin morphogenesis: hope and despair. *Current Opinion in Biotechnology*. 2003; 14(5):520-5. [DOI:10.1016/j.copbio.2003.09.005] [PMID]
- [46] Hoffman RM. The pluripotency of hair follicle stem cells. *Cell Cycle*. 2006; 5(3):232-3. [DOI:10.4161/cc.5.3.2397] [PMID]
- [47] Morris RJ, Liu Y, Marles L, Yang Z, Trempus C, Li S, et al. Capturing and profiling adult hair follicle stem cells. *Nature Biotechnology*. 2004; 22(4):411-7. [DOI:10.1038/nbt950] [PMID]
- [48] Levy V, Lindon C, Zheng Y, Harfe BD, Morgan BA. Epidermal stem cells arise from the hair follicle after wounding. *The FASEB Journal*. 2007; 21(7):1358-66. [DOI:10.1096/fj.06-6926com] [PMID]

- [49] Lako M, Armstrong L, Cairns PM, Harris S, Hole N, Jahoda CA. Hair follicle dermal cells repopulate the mouse haematopoietic system. *Journal of Cell Science*. 2002; 115(20):3967-74. [DOI:10.1242/jcs.00060] [PMID]
- [50] Shi C, Mai Y, Cheng T. Identification of hematopoietic cell populations from the dermal papillae of human hair follicles. *Transplantation Proceedings*. 2004; 36(10):3208-11. [DOI:10.1016/j.transproceed.2004.11.104] [PMID]
- [51] Toma JG, McKenzie IA, Bagli D, Miller FD. Isolation and characterization of multipotent skin-derived precursors from human skin. *Stem cells*. 2005; 23(6):727-37.
- [52] Steingrimsson E, Copeland NG, Jenkins NA. Melanocyte stem cell maintenance and hair graying. *Cell*. 2005; 121(1):9-12.
- [53] Young HE, Steele TA, Bray RA, Hudson J, Floyd JA, Hawkins K, et al. Human reserve pluripotent mesenchymal stem cells are present in the connective tissues of skeletal muscle and dermis derived from fetal, adult, and geriatric donors. *The Anatomical Record*. 2001; 264(1):51-62. [DOI:10.1002/ar.1128] [PMID]
- [54] Badiavas EV, Abedi M, Butmarc J, Falanga V, Quesenberry P. Participation of bone marrow derived cells in cutaneous wound healing. *Journal of Cellular Physiology*. 2003; 196(2):245-250. [DOI:10.1002/jcp.10260] [PMID]
- [55] Falanga V, Iwamoto S, Chartier M, Hudson J, Floyd JA, Hawkins K, et al. Autologous bone marrow-derived cultured mesenchymal stem cells delivered in a fibrin spray accelerate healing in murine and human cutaneous wounds. *Tissue Engineering*. 2007; 13(6):1299-1312. [DOI:10.1089/ten.2006.0278] [PMID]
- [56] Yoshikawa T, Mitsuno H, Nonaka I, Sen Y, Kawanishi K, Inada Y, et al. Wound therapy by marrow mesenchymal cell transplantation. *Plastic and Reconstructive Surgery*. 2008; 121(3):860-77. [DOI:10.1097/01.prs.0000299922.96006.24] [PMID]
- [57] Dash NR, Dash SN, Routray P, Mohapatra S, Mohapatra PC. Targeting nonhealing ulcers of lower extremity in human through autologous bone marrow-derived mesenchymal stem cells. *Rejuvenation Research*. 2009; 12(5):359-66. [DOI:10.1089/rej.2009.0872] [PMID]
- [58] Lee HC, An SG, Lee HW, Park JS, Cha KS, Hong TJ, et al. Safety and effect of adipose tissue-derived stem cell implantation in patients with critical limb ischemia: A pilot study. *Circulation Journal*. 2012; 76(7):1750-60. [DOI:10.1253/circj.CJ-11-1135] [PMID]
- [59] Amini N, Vousooghi N, Hadjighassem M, Bakhtiyari M, Mousavi N, Safakheil H, et al. Efficacy of human adipose tissue-derived stem cells on neonatal bilirubin encephalopathy in rats. *Neurotoxicity Research*. 2016; 29(4):514-24. [DOI:10.1007/s12640-016-9599-3] [PMID]
- [60] Asahara T, Masuda H, Takahashi T, Kalka C, Pastore C, Silver M, et al. Bone marrow origin of endothelial progenitor cells responsible for postnatal vasculogenesis in physiological and pathological neovascularization. *Circulation Research*. 1999; 85(3):221-8. [DOI:10.1161/01.RES.85.3.221] [PMID]
- [61] Guenou H, Nissan X, Larcher F, Feteira J, Lemaitre G, Saidani M, et al. Human embryonic stem-cell derivatives for full reconstruction of the pluristratified epidermis: A preclinical study. *Lancet*. 2009; 374(9703):1745-53. [DOI:10.1016/S0140-6736(09)61496-3]
- [62] Yang R, Zheng Y, Burrows M, Liu S, Wei Z, Nace A, et al. Generation of folliculogenic human epithelial stem cells from induced pluripotent stem cells. *Nature Communications*. 2014; 5:3071. [DOI:10.1038/ncomms4071] [PMID] [PMCID]
- [63] Itoh M, Umegaki Arao N, Guo Z, Liu L, Higgins CA, Christiano AM. Generation of 3D skin equivalents fully reconstituted from human Induced Pluripotent Stem Cells (iPSCs). *PLoS One*. 2013; 8(10):e77673. [DOI:10.1371/journal.pone.0077673] [PMID] [PMCID]
- [64] Sebastiano V, Zhen HH, Haddad B, Bashkurova E, Melo SP, Wang P, et al. Human COL7A1-corrected induced pluripotent stem cells for the treatment of recessive dystrophic epidermolysis bullosa. *Science Translational Medicine*. 2014; 6(264):264ra163. [DOI:10.1126/scitranslmed.3009540] [PMID] [PMCID]
- [65] Martin P, Leibovich SJ. Inflammatory cells during wound repair: The good, the bad and the ugly. *Trends in Cell Biology*. 2005; 15(11):599-607. [DOI:10.1016/j.tcb.2005.09.002] [PMID]
- [66] Takahashi K, Yamanaka S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell*. 2006; 126(4):663-76. [DOI:10.1016/j.cell.2006.07.024] [PMID]
- [67] Odorico JS, Kaufman DS, Thomson JA. Multilineage differentiation from human embryonic stem cell lines. *Stem Cells*. 2001; 19(3):193-204. [DOI:10.1634/stemcells.19-3-193] [PMID]
- [68] Wu DC, Boyd AS, Wood KJ. Embryonic stem cell transplantation: Potential applicability in cell replacement therapy and regenerative medicine. *Frontiers in Bioscience*. 2007; 12(8-12):4525-35. [DOI:10.2741/2407]
- [69] Galderisi U, Giordano A. The gap between the physiological and therapeutic roles of mesenchymal stem cells. *Medicinal Research Reviews*. 2014; 34(5):1100-26. [DOI:10.1002/med.21322] [PMID]
- [70] Kirby GT, Mills SJ, Cowin AJ, Smith LE. Stem cells for cutaneous wound healing. *Biomedical Research International*. 2015; 2015:285869. [DOI:10.1155/2015/285869] [PMID] [PMCID]
- [71] Wu Y, Wang J, Scott PG, Tredget EE. Bone marrow-derived stem cells in wound healing: A review. *Wound Repair and Regeneration*. 2007; 15:S18-26.
- [72] Salem HK, Thiemermann C. Mesenchymal stromal cells: Current understanding and clinical status. *Stem Cells*. 2010; 28(3):585-96. [PMID]
- [73] Taylor DA, Zenovich AG. Cardiovascular cell therapy and endogenous repair. *Diabetes, Obesity & Metabolism*. 2008; 10:5-15. [DOI:10.1111/j.1463-1326.2008.00937.x] [PMID] [PMCID]
- [74] Singer AJ, Clark RA. Cutaneous wound healing. *The New England Journal of Medicine*. 1999; 341(10):738-46. [DOI:10.1056/NEJM1999023411006] [PMID]
- [75] Gnecci M, Zhang Z, Ni A, Dzau VJ. Paracrine mechanisms in adult stem cell signaling and therapy. *Circulation Research*. 2008; 103(11):1204-19. [DOI:10.1161/CIRCRESAHA.108.176826] [PMID] [PMCID]
- [76] Maxson S, Lopez EA, Yoo D, Danilkovitch-Miagkova A, Leroux MA. Concise review: Role of mesenchymal stem cells in wound repair. *Stem Cells Translational Medicine*. 2012; 1(2):142-9. [DOI:10.5966/sctm.2011-0018] [PMID] [PMCID]
- [77] Lu D, Chen B, Liang Z, Deng W, Jiang Y, Li S, et al. Comparison of bone marrow mesenchymal stem cells with bone mar-

- row-derived mononuclear cells for treatment of diabetic critical limb ischemia and foot ulcer: A double-blind, randomized, controlled trial. *Diabetes Research and Clinical Practice*. 2011; 92(1):26-36. [DOI:10.1016/j.diabres.2010.12.010] [PMID]
- [78] Chunmeng S, Tianmin C, Yongping S, Xinze R, Yue M, Jifu Q, et al. Effects of dermal multipotent cell transplantation on skin wound healing. *The Journal of Surgical Research*. 2004; 121(1):13-19. [DOI:10.1016/j.jss.2004.04.008] [PMID]
- [79] Martin P. Wound healing-aiming for perfect skin regeneration. *Science*. 1997; 276(5309):75-81. [DOI:10.1126/science.276.5309.75] [PMID]
- [80] Mori L, Bellini A, Stacey MA, Schmidt M, Mattoli S. Fibrocytes contribute to the myofibroblast population in wounded skin and originate from the bone marrow. *Experimental Cell Research*. 2005; 304(1):81-90. [DOI:10.1016/j.yexcr.2004.11.011] [PMID]
- [81] Harris RG, Herzog EL, Bruscia EM, Grove JE, Van Arnem JS, Krause DS. Lack of a fusion requirement for development of bone marrow-derived epithelia. *Science*. 2004; 305(5680):90-93. [DOI:10.1126/science.1098925] [PMID]
- [82] Borue X, Lee S, Grove J, Herzog EL, Harris R, Diflo T, et al. Bone marrow-derived cells contribute to epithelial engraftment during wound healing. *The American Journal of Pathology*. 2004; 165(5):1767-72. [DOI:10.1016/S0002-9440(10)63431-1]
- [83] Alvarez Viejo M, Menendez Menendez Y, Otero Hernandez J. CD271 as a marker to identify mesenchymal stem cells from diverse sources before culture. *World Journal of Stem Cells*. 2015; 7(2):470-6. [DOI:10.4252/wjsc.v7.i2.470] [PMID] [PMCID]
- [84] Zuk PA, Zhu M, Ashjian P, De Ugarte DA, Huang JI, Mizuno H, et al. Human adipose tissue is a source of multipotent stem cells. *Molecular Biology of The Cell*. 2002; 13(12):4279-95. [DOI:10.1091/mbc.e02-02-0105] [PMID] [PMCID]
- [85] Kilroy GE, Foster SJ, Wu X, Ruiz J, Sherwood S, Heifetz A, et al. Cytokine profile of human adipose-derived stem cells: expression of angiogenic, hematopoietic, and pro-inflammatory factors. *Journal of Cellular Physiology*. 2007; 212(3):702-9. [DOI:10.1002/jcp.21068] [PMID]
- [86] Caruana G, Bertozzi N, Boschi E, Pio Grieco M, Grignaffini E, Rapisio E. Role of adipose-derived stem cells in chronic cutaneous wound healing. *Annali Italiani di Chirurgia*. 2015; 86(1):1-4. [PMID]
- [87] Wallner C, Abraham S, Wagner JM, Harati K, Ismer B, Kessler L, et al. Local application of isogenic adipose-derived stem cells restores bone healing capacity in a type 2 diabetes model. *Stem Cells Translational Medicine*. 2016; 5(6):836-44. [DOI:10.5966/sctm.2015-0158] [PMID] [PMCID]
- [88] Talaie Khozani T, Heidari F, Esmailpour T, Vojdani Z, Mostafavi Pour Z, Rohani L. Cardiomyocyte marker expression in mouse embryonic fibroblasts by cell-free cardiomyocyte extract and epigenetic manipulation. *Iranian Journal of Medical Sciences*. 2014; 39(2 Suppl):203-12. [PMID] [PMCID]
- [89] Joshi CV, Enver T. Plasticity revisited. *Current Opinion in Cell Biology*. 2002; 14(6):749-55. [DOI:10.1016/S0955-0674(02)00392-7]
- [90] King A, Balaji S, Keswani SG, Crombleholme TM. The Role of Stem Cells in Wound Angiogenesis. *Advances in Wound Care*. 2014; 3(10):614-25. [DOI:10.1089/wound.2013.0497] [PMID] [PMCID]
- [91] Ronfard V, Rives JM, Neveux Y, Carsin H, Barrandon Y. Long-term regeneration of human epidermis on third degree burns transplanted with autologous cultured epithelium grown on a fibrin matrix. *Transplantation*. 2000; 70(11):1588-98. [DOI:10.1097/00007890-200012150-00009] [PMID]
- [92] Mistriotis P, Andreadis ST. Hair follicle: A novel source of multipotent stem cells for tissue engineering and regenerative medicine. *Reviews*. 2013; 19(4):265-78. [DOI:10.1089/ten.teb.2012.0422] [PMID] [PMCID]
- [93] Jahoda CA, Whitehouse J, Reynolds AJ, Hole N. Hair follicle dermal cells differentiate into adipogenic and osteogenic lineages. *Experimental Dermatology*. 2003; 12(6):849-59. [DOI:10.1111/j.0906-6705.2003.00161.x] [PMID]
- [94] Reynolds AJ, Lawrence C, Cserhalmi Friedman PB, Christiano AM, Jahoda CA. Trans-gender induction of hair follicles. *Nature*. 1999; 402(6757):33-34. [DOI:10.1038/46938] [PMID]
- [95] Shi C, Zhu Y, Su Y, Cheng T. Stem cells and their applications in skin-cell therapy. *Trends in Biotechnology*. 2006; 24(1):48-52. [DOI:10.1016/j.tibtech.2005.11.003] [PMID]
- [96] Christoph T, Muller Rover S, Audring H, Tobin DJ, Hermes B, Cotsarelis G, et al. The human hair follicle immune system: Cellular composition and immune privilege. *The British Journal of Dermatology*. 2000; 142(5):862-73. [DOI:10.1046/j.1365-2133.2000.03464.x] [PMID]
- [97] Najafzadeh N, Sagha M, Heydari Tajaddod S, Golmohammadi MG, Massahi Oskoui N, Deldadeh Moghaddam M. In vitro neural differentiation of CD34 stem cell populations in hair follicles by three different neural induction protocols. *In vitro Cellular & Developmental Biology- Animal*. 2015; 51(2):192-203.
- [98] Jaks V, Kasper M, Toftgard R. The hair follicle-a stem cell zoo. *Experimental Cell Research*. 2010; 316(8):1422-28. [DOI:10.1016/j.yexcr.2010.03.014] [PMID]
- [99] Heidari F, Yari A, Rasoolijazi H, Soleimani M, Dehpoor A, Sajedi N, et al. Bulge hair follicle stem cells accelerate cutaneous wound healing in rats. *Wounds: A Compendium of Clinical Research And Practice*. 2016; 28(4):132-41. [PMID]
- [100] Hoffman RM. Gene and stem cell therapy of the hair follicle. *Methods in Molecular Biology*. 2005; 289:437-48. [PMID]
- [101] Heng BC, Cao T, Liu H, Phan TT. Directing stem cells into the keratinocyte lineage in vitro. *Experimental Dermatology*. 2005; 14(1):1-16. [DOI:10.1111/j.0906-6705.2005.00262.x] [PMID]
- [102] Ghoroghi FM, Hejazian LB, Esmailzade B, Dodel M, Roudbari M, Nobakht M. Evaluation of the effect of NT-3 and biodegradable Poly-L-lactic acid nanofiber scaffolds on differentiation of rat hair follicle stem cells into neural cells in vitro. *Journal of Molecular Neuroscience*. 2013; 51(2):318-27.
- [103] Yari A, Teimourian S, Amidi F, Bakhtiyari M, Heidari F, Sajedi N, et al. The role of biodegradable engineered random polycaprolactone nanofiber scaffolds seeded with nestin-positive hair follicle stem cells for tissue engineering. *Advanced Biomedical Research*. 2016; 5:22. [DOI:10.4103/2277-9175.175911] [PMID] [PMCID]
- [104] Gilanchi S, Esmailzade B, Eidi A, Barati M, Mehrabi S, Ghoroghi FM, et al. Neuronal differentiation of rat hair follicle stem cells: The involvement of the neuroprotective factor Seldin-1 (DHCR24). *Iranian Biomedical Journal*. 2014; 18(3):136-42. [PMID] [PMCID]

- [105] Hoffman RM. Nestin-expressing hair follicle-accessible pluripotent stem cells for nerve and spinal cord repair. *Cells, Tissues, Organs*. 2014; 200(1):42-7. [DOI:10.1159/000366098] [PMID]
- [106] Amoh Y, Katsuoka K, Hoffman RM. The advantages of hair follicle pluripotent stem cells over embryonic stem cells and induced pluripotent stem cells for regenerative medicine. *Journal of Dermatological Science*. 2010; 60(3):131-37. [DOI:10.1016/j.jdermsci.2010.09.007] [PMID]
- [107] Ito M, Liu Y, Yang Z, Nguyen J, Liang F, Morris RJ, et al. Stem cells in the hair follicle bulge contribute to wound repair but not to homeostasis of the epidermis. *Nature Medicine*. 2005; 11(12):1351-4. [DOI:10.1038/nm1328] [PMID]
- [108] Langton AK, Herrick SE, Headon DJ. An extended epidermal response heals cutaneous wounds in the absence of a hair follicle stem cell contribution. *The Journal of Investigative Dermatology*. 2008; 128(5):1311-8. [DOI:10.1038/sj.jid.5701178] [PMID]
- [109] Page ME, Lombard P, Ng F, Gottgens B, Jensen KB. The epidermis comprises autonomous compartments maintained by distinct stem cell populations. *Cell Stem Cell*. 2013; 13(4):471-82. [DOI:10.1016/j.stem.2013.07.010] [PMID] [PMCID]
- [110] Brownell I, Guevara E, Bai CB, Loomis CA, Joyner AL. Nerve-derived sonic hedgehog defines a niche for hair follicle stem cells capable of becoming epidermal stem cells. *Cell Stem Cell*. 2011; 8(5):552-65. [DOI:10.1016/j.stem.2011.02.021] [PMID] [PMCID]
- [111] Mascre G, Dekoninck S, Drogat B, Youssef KK, Brohée S, Sotiropoulou PA, et al. Distinct contribution of stem and progenitor cells to epidermal maintenance. *Nature*. 2012; 489(7415):257-62. [DOI:10.1038/nature11393] [PMID]
- [112] Lowell S, Jones P, Le Roux I, Dunne J, Watt FM. Stimulation of human epidermal differentiation by delta-notch signalling at the boundaries of stem-cell clusters. *Current Biology*. 2000; 10(9):491-500. [DOI:10.1016/S0960-9822(00)00451-6]
- [113] Guo L, Yu QC, Fuchs E. Targeting expression of keratinocyte growth factor to keratinocytes elicits striking changes in epithelial differentiation in transgenic mice. *The EMBO Journal*. 1993; 12(3):973-86. [DOI:10.1002/j.1460-2075.1993.tb05738.x] [PMID] [PMCID]
- [114] Esmailzade B, Nobakht M, Joghataei MT, Roshandel NR, Rasouli H, Kuchaksaraei AS, et al. Delivery of Epidermal Neural Crest Stem Cells (EPI-NCSC) to hippocamp in Alzheimer's disease rat model. *Iranian Biomedical Journal*. 2012; 16(1):1-9. [PMID] [PMCID]
- [115] Plikus MV, Gay DL, Treffeisen E, Wang A, Supapannachart RJ, Cotsarelis G. Epithelial stem cells and implications for wound repair. *Seminars in Cell & Developmental Biology*. 2012; 23(9):946-53. [DOI:10.1016/j.semcdb.2012.10.001] [PMID] [PMCID]
- [116] Blanpain C, L, Geoghegan A, Polak L, Fuchs E. Self-renewal, multipotency, and the existence of two cell populations within an epithelial stem cell niche. *Cell*. 2004; 118:635-48. [DOI:10.1016/j.cell.2004.08.012] [PMID]
- [117] Ohyama TA, Tock CL, Radonovich MF, Pise Masison CA, Hopping SB, Brady JN, et al. Characterization and isolation of stem cell-enriched human hair follicle bulge cells. *The Journal of Clinical Investigation*. 2006; 116(1):249-60. [DOI:10.1172/JCI26043] [PMID] [PMCID]
- [118] Ohyama M, Vogel JC, Amagai M. Gene ontology analysis of human hair follicle bulge molecular signature. *Journal of Dermatological Science*. 2007; 45(2):147-50. [DOI:10.1016/j.jdermsci.2006.09.009] [PMID]
- [119] Veijouyeh SJ, Mashayekhi F, Yari A, Heidari F, Sajedi N, Ghoroghi FM, et al. In vitro induction effect of 1, 25 (OH) 2D3 on differentiation of hair follicle stem cell into keratinocyte. *Biomedical Journal*. 2017; 40(1):31-8.
- [120] Jensen KB, Collins CA, Nascimento E, Tan DW, Frye M, Itami S, et al. Lrig1 expression defines a distinct multipotent stem cell population in mammalian epidermis. *Cell Stem Cell*. 2009; 4(5):427-39. [DOI:10.1016/j.stem.2009.04.014] [PMID] [PMCID]
- [121] Snippert HJ, Haegerbarth A, Kasper M, Jaks V, van Es JH, Barker N, et al. Lgr6 marks stem cells in the hair follicle that generate all cell lineages of the skin. *Science*. 2010; 327(5971):1385-9. [DOI:10.1126/science.1184733] [PMID]
- [122] Horsley V, O'Carroll D, Toozé R, Ohinata Y, Saitou M, Obukhanych T, et al. Blimp1 defines a progenitor population that governs cellular input to the sebaceous gland. *Cell*. 2006; 126(3):597-609. [DOI:10.1016/j.cell.2006.06.048] [PMID] [PMCID]
- [123] Veijouyeh SJ, Abazar Y, Heidari F, Sajedi N, Moghani FG, Nobakht M. Bulge region as a putative hair follicle stem cells niche: A brief review. *Iranian Journal of Public Health*. 2017; 46(9):1167-75. [PMID] [PMCID]