



Pulsed Magnetic Fields Cause *hes1* and *hsa-Circ-0068530* Expression Changes in Gastric Cancer Cell Line and Human Normal Fibroblast Cell Line

Fereshteh Mansoury¹, Soheila Abdi ^{2,*}, Nahid Babaei¹, Maliheh Entezari ^{3,4}, Abbas Doosti ⁵ and Fatemeh Mashayekhi²

¹Department of Cell Biology and Genetics, Bushehr Branch, Islamic Azad University, Bushehr, Iran

²Department of Physics, Safadasht Branch, Islamic Azad University, Tehran, Iran

³Department of Genetics, Faculty of Advanced Science and Technology, Tehran Medical Sciences Branch, Islamic Azad University, Tehran, Iran

⁴Farhikhtegan Medical Convergence Sciences Research Center, Farhikhtegan Sciences, Islamic Azad University, Tehran, Iran

⁵Biotechnology Research Center, Shahrekord Branch, Islamic Azad University, Shahrekord, Iran

*Corresponding author: Department of Physics, Safadasht Branch, Islamic Azad University, Tehran, Iran. Email: soheilaabdi@safaiu.ac.ir

Received 2021 August 28; Revised 2021 September 26; Accepted 2021 October 02.

Abstract

Background: In recent years, the relationship between cancer cells and electromagnetic radiation has received much attention.

Objectives: The present study aimed to evaluate the effects of different intensities of electromagnetic fields on gastric cancer cell lines (AGS).

Methods: After preparing AGS and Hu02 (normal) cell lines, they were exposed to magnetic flux densities of 0.25, 0.5, 1, and 2 millitesla (mT) for 18 h. The cell viability was studied by the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay. The expression levels of *hes1* and *hsa-circ-0068530* RNAs were studied by the quantitative Real-time-PCR technique.

Results: The inhibition of gastric cancer cell line growth was observed under the influence of electromagnetic fields at different intensities. However, they did not affect the viability of normal cells. A sharp increase in the expression of *hes1* and *hsa-circ-0068530* genes was observed in normal cells exposed to 2 mT electromagnetic fields.

Conclusions: In general, it can be concluded that the effect of electromagnetic fields on gastric cancer cells depends on their intensity. Magnetic flux densities of 0.25 and 0.5 mT had anti-cancer effects and magnetic flux density of 2 mT showed carcinogenic effects.

Keywords: Gastric Cell Line (AGS), Magnetic Field, *hes1* Expression, *Hsa-circ-0068530* Expression

1. Background

In recent decades, the use of electricity has expanded significantly and become one of the hallmarks of advanced societies. Therefore, researchers have focused on the biological effects of electromagnetic fields. Several studies have shown a relationship between electromagnetic fields and multiple types of cancer (1, 2). Low-frequency electromagnetic fields can induce heat in tissues and cells. This increase in temperature can lead to cell death (3). It has also been reported that short-term exposure of pregnant rats to the electromagnetic field causes teratogenicity effects in the developing fetus (4). Some researchers believe that low-frequency electromagnetic radiation acts like ionizing waves and causes DNA damage by inducing mutations (5). Increasing the concentration of free radicals and changes in cell behavior following electromagnetic fields

irradiation can lead to DNA damage (6). Low-frequency electromagnetic fields can affect cell growth (7), morphology and cell shape (8), carcinogenicity (9), cell differentiation (10), and programmed cell death (11). Exposure to low-frequency electromagnetic fields could increase oxidative stress in chick embryos (12), cultured mammalian cells (13), and human erythrocytes (14). Gastric cancer is a multifactorial disease in which bacterial contamination, environmental factors, and host genetic agents play an important role in its progression (15). A review of statistics from the last 30 years shows that the incidence of gastric cancer in Iran is higher than the global average, and despite the decrease in the incidence of gastric cancer in the world, its incidence is increasing in Iran (16). Studies show that the onset and progression of cancer depend on several factors, including genetic background (17).

The *hes1* gene is a target gene for the Notch1 signaling pathway and plays an important role in preserving neural stem cells and intestinal precursor cells and determining cell fate and apoptosis (18). The Notch1 receptor is also one of the most frequent receptors in the Notch signaling pathway. The expression of Notch1-3 and hairy enhancer of split 1 (*hes1*) has been reported in the human gastric mucosa (19). As known, Notch1 controls the number and fate of intestinal stem cells by increasing *hes1* (20). Studies have also shown that Notch receptors and their ligands play an important role in some cancers (21). The expression of Notch1 and *hes1* is associated with cancer cell proliferation and angiogenesis, as reported that the expression of these genes is very high in cancer stem cells (CSCs) (22, 23).

A group of non-coding RNAs that have a circular structure is called circRNAs that have been identified for their role in regulating gene expression at the transcriptional level and, subsequently their sponge miRNA function (24). In recent years, the association of circRNAs with a variety of cancers has attracted much attention and has been cited as a biomarker for cancer diagnosis (25).

2. Objectives

This research evaluated the effect of extremely low frequency (ELF) magnetic flux densities (MFDs) of 0.25, 0.5, 1, and 2 mT on *hes1* and *hsa-Circ-0068530* expression levels.

3. Methods

3.1. Cell Culture

Two AGS cell lines and a HuO2 fibroblast cell line were purchased from the National Genetic Re-sources Center of Iran. Ham's F12 medium (Gibco, USA) was used for gastric cancer cell line culture, and Dulbecco's modified Eagle's medium (DMEM; Gibco, USA) was used for culture of normal HuO2 cells. A humidified incubator at $37 \pm 2^\circ\text{C}$ with 5% CO_2 was used to maintain each cell line (26).

3.2. Exposure System

Figure 1 shows the exposure system that included a solenoid cylinder explained in our previous study (27, 28). The cells were exposed to ELF magnetic flux densities of 0.25, 0.5, 1, and 2 mT for 18 h. The control and exposure cells were incubated in a constant condition of temperature, humidity, and CO_2 (26).

3.3. MTT Assay

The MTT assay was used to evaluate cell viability according to our previous study (26).

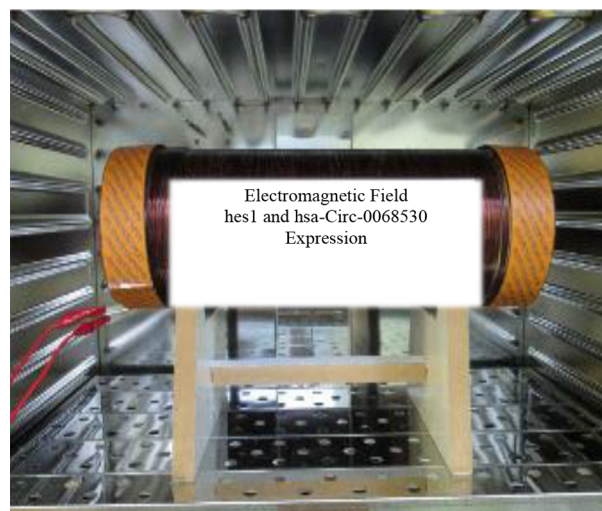


Figure 1. Magnetic field exposure setup

3.4. Quantitative Real-Time Polymerase Chain Reaction (qRT-PCR)

The Real-time PCR was used to measure expression changes of *hes1* and *hsa-Circ-0068530*. Total RNA was extracted by using TRIzol reagent (TRI Sigma-Aldrich) according to the manufacturer's instructions. Total extracted RNA concentration was qualified by measuring the absorbance at 260 nm. Then, cDNA was synthesized from the total extracted RNA using the cDNA Synthesis Kit (Biofact, Korea) with oligo (dT) or random primers. Finally, qRT-PCR was performed by Bioneer Exicycler™'s 96 Detection System. The primer sequences are listed in Table 1.

Table 1. Sequences of GAPDH, *hes1*, and *hsa-circ-0068530* Primers

Genes	Sequences
hsa-circ-0068530	
Forward	5'-GGAAATGACAGTGAAGCACCTCC-3'
Reverse	5'-GAAGCGGGTCACCTCGTTCATG-3'
hes1	
Forward	5'-GAGTGCATGAACGAGGTGAC-3'
Reverse	5'-GGTCATGGCATTGATCTGGG-3'
GAPDH	
Forward	5'-GCACCGTCAAGGCTGAGAAC-3'
Reverse	5'-GGATCTCGCTCCTGGAAGATG-3'

The $2^{-\Delta\Delta\text{CT}}$ method was applied to determine relative changes in gene expression in samples (26, 29). Each experiment consisted of six separated flasks of cells. After extraction of RNA and synthesis of cDNA, the qRT-PCR test was

done in duplicate.

3.5. Statistical Analysis

All statistical analyses were performed with SPSS 25.0 (IBM, SPSS, Chicago, USA). The values are expressed as mean \pm standard deviation (SD) from three independent experiments performed in duplicate. Statistical analyses were carried out using a two-independent-sample and Mann-Whitney U test. Bivariate correlations between variables were analyzed by the Spearman test. Differences in values were considered significant if P-value < 0.05.

4. Results

4.1. MTT Assay

The results showed a significant difference in the survival rate of the tumor and normal cells following exposure to different electromagnetic fields ($P < 0.05$). Electromagnetic fields decreased the survival rate of tumor cells while normal cells continued to multiply and their percentage increased. The inhibitory effect of tumor cell division was observed from an intensity of 0.5 to 2 mT (Figure 2). Thus, electromagnetic fields showed inhibitory effects on cell division in the gastric tumor cell line.

4.2. *hes1* Expression

The expression of the *hes1* gene in tumor cells was dependent on the intensities of electromagnetic fields, but in normal cells, no change in the expression of this gene was observed with increasing the intensity of the electromagnetic field. The expression of *hes1* in gastric cancer cells was downregulated at intensities of 0.25 and 0.5 mT compared to controls, but with an increase in the electromagnetic field to 2 mT, this gene was overexpressed (Figure 3).

4.3. *hsa-circ-0068530*

There was a significant difference in the expression of *hsa-circ-0068530* between normal and tumor cells exposed to different intensities of the electromagnetic field. The expression of this circRNA in tumor cells initially showed a decreasing trend when exposed to the intensities of 0.25, 0.5, and 1 mT, which overexpressed sharply with increasing the electromagnetic field to 2 mT (2.5 times compared to the control (Figure 4). In normal cells, the expression of *hsa-circ-0068530* decreased under exposure to electromagnetic fields in comparison with the control group, and the greatest decrease was seen in the magnetic flux density of 2 mT.

4.4. Correlation Analysis

A positive significant correlation ($r = 0.908$, 95% $P < 0.0001$) was observed between the expression levels of *hsa-circ-0068530* and *hes1* genes in the AGS cell lines exposed to electromagnetic fields (Figure 5A). But, there was no significant correlation between the expression levels of *hsa-circ-0068530* and *hes1* genes in normal cells exposed to electromagnetic fields ($P = 0.540$) (Figure 5B).

5. Discussion

In this study, AGS gastric cancer cell line and normal fibroblast cell line were exposed to ELF-MFDs of 0.25, 0.5, 1, and 2 mT. The results showed that normal cells continued to proliferate over time and their viability increased. However, the survival rate of tumor cells decreased following exposure to the electromagnetic fields. Therefore, it seems that the magnetic fields can damage gastric tumor cells and prevent them from growing and multiplying. Interaction of ELF-MF with living organisms can induce different biological effects that depend on the type, MFD, frequency, and time of exposure (30, 31).

In recent years, concerns have been raised about the strong electromagnetic fields of household appliances, and more importantly, high-pressure towers and cell phone waves, on human health. It seems that with the increasing intensity of electromagnetic fields, their biological effects on living systems increase (32). The production of liver cysts with fibrotic bands, severe obstructive hepatitis, and edema in chick embryos exposed to 50 Hz electromagnetic field (33), DNA damage due to oxidative stress (34), and slow cell division due to the inhibition of mitotic spindle formation (35) have been reported. In the present study, normal fibroblast cells following exposure to magnetic fields showed no reduction in viability, which is contrary to the findings of the above-mentioned studies. This can be attributed to the different magnetic intensities used in the present study. The inhibitory effect of magnetic fields on the growth and proliferation of gastric cancer cells can be attributed to the DNA fragmentation of cancer cells, inhibition of antioxidant enzymes, and reduced cell tolerance to oxidative stress. These events affect the cell signal transduction pathways and the expression of genes specific for the inflammatory response, cell growth, differentiation, and proliferation, and generally reduce cancer cell growth and inhibit its proliferation (36). Cancer and normal cells showed different cellular behaviors to interact with electromagnetic fields. It has been shown that electromagnetic fields induce an increase in free radicals in the environment. On the other hand, an increase in free radicals leads to oxidative stress, which is one of the causes of cell death (37). In normal cells, several detoxification

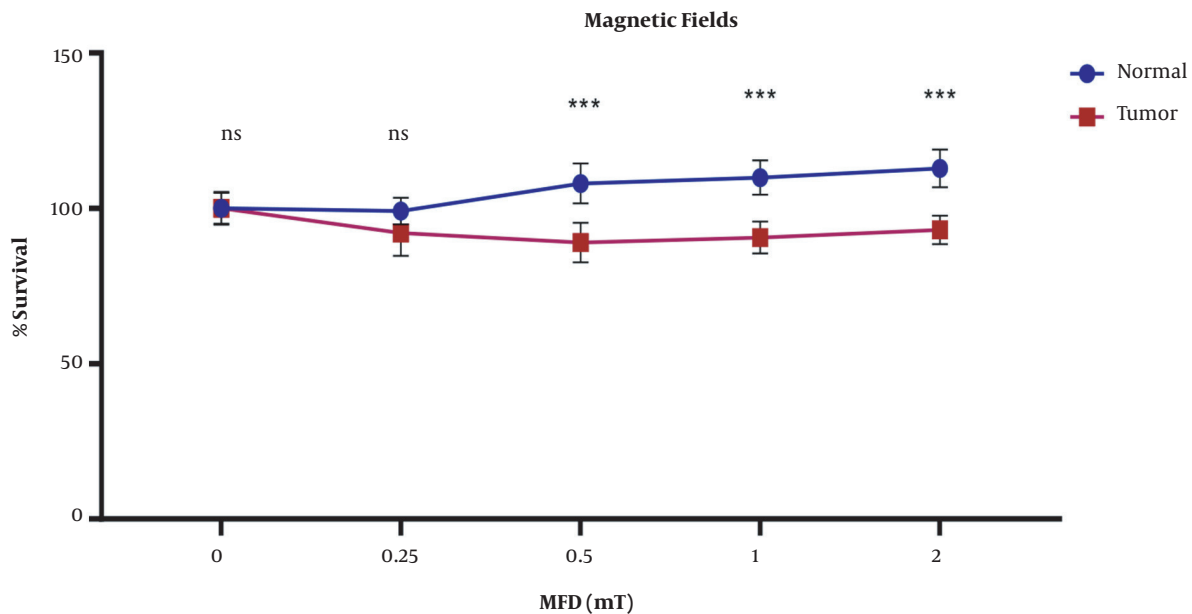


Figure 2. Effects of magnetic flux densities of 0.25 - 2 mT on cell survival of gastric cancer cell and normal cell lines (**P value < 0.001)

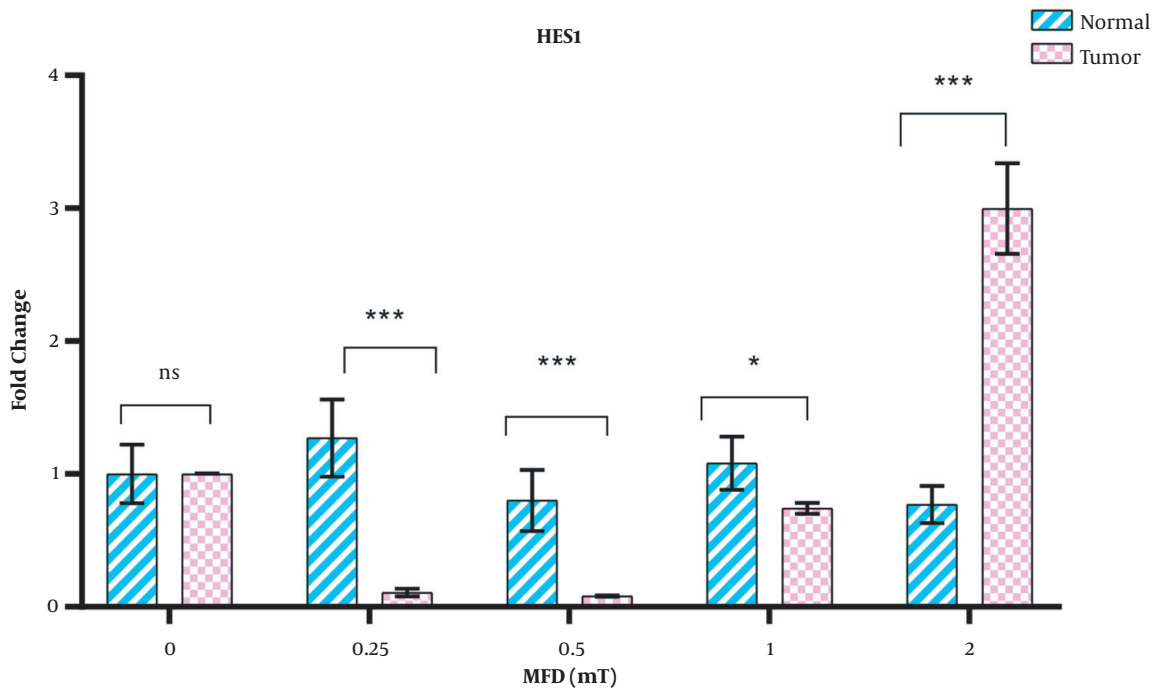


Figure 3. Effects of magnetic flux densities of 0.25 - 2 mT on hes1 gene expression in normal and gastric cancer lines

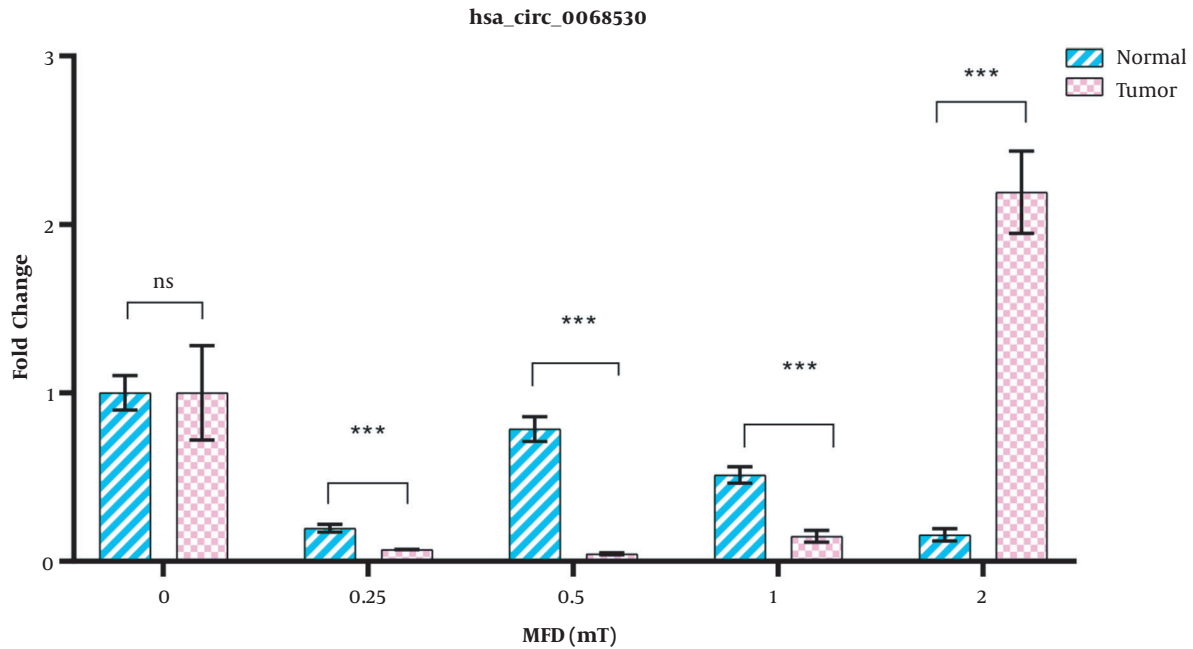


Figure 4. Effects of magnetic flux densities of 0.25 - 2 mT on hsa-circ-0068530 expression in normal and gastric cancer lines

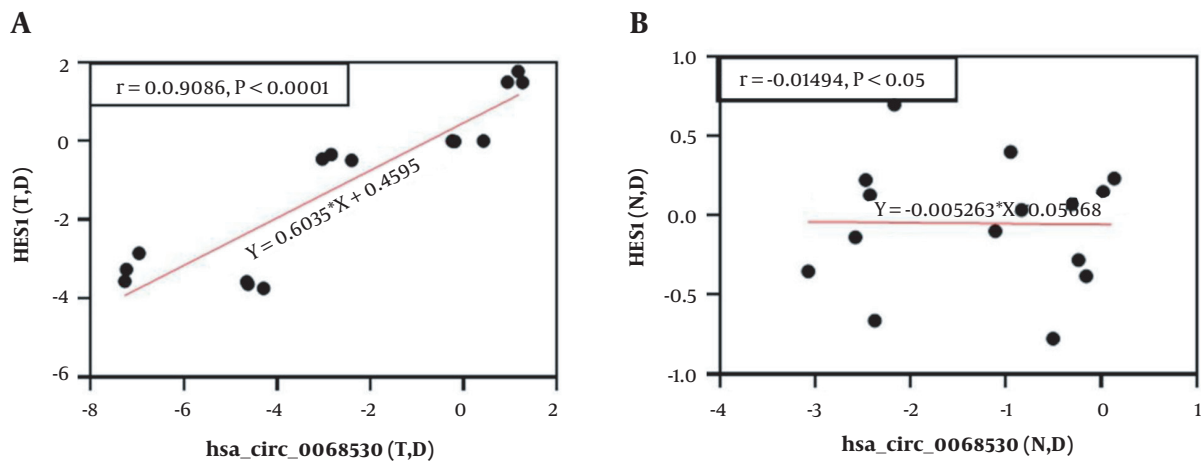


Figure 5. Correlation analysis between hest and hsa-circ-0068530 gene expression in A, gastric tumor cells; and B, normal cell lines exposed to magnetic flux densities of 0.25 - 2 mT

processes regulated through antioxidant enzymes balance the ROS levels. Therefore, the homeostasis of ROS is well sustained, which can contribute to the maintenance of redox balance in normal cells (38).

In the present study, significant differences in terms of hest expression were observed in gastric tumor cells exposed to different electromagnetic field intensities. Expression of this gene decreased at low intensities of elec-

tromagnetic field (0.25 and 0.5 mT) in tumor cells but increased sharply at higher intensities (2 mT). It seems that intensities above 2 mT of electromagnetic fields may lead to increased expression of this gene in tumor cells. Research has shown that hest is overexpressed in cancer stem cells. Therefore using down-regulation of hest1 by targeting therapy can reduce the number of CSCs (23). Hence, hest1 is an oncogene that can lead to the development of

gastric cancer (39). A decrease in *hes1* expression was observed at low intensities of 0.25 and 0.5 mT of magnetic fields. However, when the intensity of the electromagnetic field increased to 2 mT, a sharp increase in the expression of this gene was observed in gastric cancer cells, which could indicate the carcinogenic effect of electromagnetic fields at high intensities. Therefore, it seems that the electromagnetic field can also exert its anti-proliferative effect through changes in the Notch transduction pathway. However, more studies are needed in this area.

One of the most important molecular elements involved in regulating gene expression is circRNAs, which can originate in either exon or intron regions (40). In recent years, the role of circRNAs in causing cancer has received much attention, which has led to the identification of a large number of circRNAs involved in a variety of cancers (41). As known, circRNAs have multiple regions for binding to miRNAs and act as miRNA sponges. They play a role in various cancers (42). In the present study, a sharp increase in the expression of *hsa-circ-0068530* in a magnetic flux density of 2mT could indicate its carcinogenic role in gastric cancer. In normal cells, no significant difference was observed in the expression *hsa-circ-0068530* at different intensities of magnetic fields. In this study, the expression of *hsa-circ-0068530* in the cancer cell line was studied for the first time, so its importance must be confirmed in other studies. Correlation analysis was performed to find the relationship between the *hes1* gene and its circRNA *hsa-circ-0068530*, which showed a direct relationship between gene expression and its circRNA in tumor cells in electromagnetic fields. There was no correlation between *hes1* and *hsa-circ-0068530* gene expression in normal cells. Considering the effect of electromagnetic fields on reducing the expression of this circRNA in normal cells, it can be said that the electromagnetic field is safe for normal cells.

5.1. Conclusion

The inhibition of AGS gastric cancer cell growth under exposure to electromagnetic fields at intensities of 0.25 and 0.5 mT was observed, indicating the cytotoxic effect of these waves on the tumor. At intensities of 1 and 2 mT, the electromagnetic fields showed an increase in the expression of *hes1* and *hsa-circ-0068530* genes, which could indicate the carcinogenic effects of electromagnetic fields at high intensities.

Acknowledgments

The authors would like to thank the staff of the Cancer Institute of Iran and Razi Drug Research Center, particularly Ms. Neda Tekieh Maroof, who contributed to this work.

Footnotes

Authors' Contribution: S. A., study concept, design, and supervision; F. M., N. B., and M. E., analysis and interpretation of data; A. D., statistical analysis; F. M., administrative, technical, and material support.

Conflict of Interests: The authors declare that there is no conflict of interest.

Ethical Approval: IR.BPUMS.REC.1399.180

Funding/Support: The present study financially supported by Ms. Fereshteh Mansoury.

References

- Blackman CF. Can EMF exposure during development leave an imprint later in life? *Electromagn Biol Med.* 2006;**25**(4):217-25. doi: [10.1080/15368370601034086](https://doi.org/10.1080/15368370601034086). [PubMed: 17178582].
- Belpomme D, Irigaray P, Hardell L. Electromagnetic fields as cancer-causing agents. *Environ Res.* 2008;**107**(2):289-90. doi: [10.1016/j.envres.2008.01.017](https://doi.org/10.1016/j.envres.2008.01.017).
- Juutilainen J. Developmental effects of electromagnetic fields. *Bioelectromagnetics.* 2005;**Suppl 7**:S107-15. doi: [10.1002/bem.20125](https://doi.org/10.1002/bem.20125). [PubMed: 16037961].
- Yang MJ, Liu JY, Wang YF, Lang HY, Miao X, Zhang LY, et al. Effects of electromagnetic pulse on polydactyly of mouse fetuses. *Theriogenology.* 2013;**80**(1):18-23. doi: [10.1016/j.theriogenology.2013.03.004](https://doi.org/10.1016/j.theriogenology.2013.03.004). [PubMed: 23623167].
- Reese JA, Jostes RF, Frazier ME. Exposure of mammalian cells to 60-Hz magnetic or electric fields: Analysis for DNA single-strand breaks. *Bioelectromagnetics.* 1988;**9**(3):237-47. doi: [10.1002/bem.2250090305](https://doi.org/10.1002/bem.2250090305). [PubMed: 3178898].
- Barnes FS. Some engineering models for interactions of electric and magnetic fields with biological systems. *Bioelectromagnetics.* 1992;**Suppl 1**:67-85. doi: [10.1002/bem.2250130708](https://doi.org/10.1002/bem.2250130708). [PubMed: 1285723].
- Patruno A, Ferrone A, Costantini E, Franceschelli S, Pesce M, Speranza L, et al. Extremely low-frequency electromagnetic fields accelerates wound healing modulating MMP-9 and inflammatory cytokines. *Cell Prolif.* 2018;**51**(2). e12432. doi: [10.1111/cpr.12432](https://doi.org/10.1111/cpr.12432). [PubMed: 29357406]. [PubMed Central: PMC6528910].
- Koziorowska A, Romerowicz-Misielak M, Solek P, Koziorowski M. Extremely low frequency variable electromagnetic fields affect cancer and noncancerous cells in vitro differently: Preliminary study. *Electromagn Biol Med.* 2018;**37**(1):35-42. doi: [10.1080/15368378.2017.1408021](https://doi.org/10.1080/15368378.2017.1408021). [PubMed: 29513614].
- Kocaman A, Altun G, Kaplan AA, Deniz OG, Yurt KK, Kaplan S. Genotoxic and carcinogenic effects of non-ionizing electromagnetic fields. *Environ Res.* 2018;**163**:71-9. doi: [10.1016/j.envres.2018.01.034](https://doi.org/10.1016/j.envres.2018.01.034). [PubMed: 29427953].
- Bai W, Li M, Xu W, Zhang M. Comparison of effects of high- and low-frequency electromagnetic fields on proliferation and differentiation of neural stem cells. *Neurosci Lett.* 2021;**741**:135463. doi: [10.1016/j.neulet.2020.135463](https://doi.org/10.1016/j.neulet.2020.135463). [PubMed: 33129846].
- Yadmani S, Neamati A, Homayouni-Tabrizi M, Beyramabadi SA, Yadmani S, Gharib A, et al. Treatment of the breast cancer by using low frequency electromagnetic fields and Mn(II) complex of a Schiff base derived from the pyridoxal. *Breast.* 2018;**41**:1107-12. doi: [10.1016/j.breast.2018.07.001](https://doi.org/10.1016/j.breast.2018.07.001). [PubMed: 30025273].
- Siddiqi N, Al Nazwani N. Effects of electromagnetic field on the development of chick embryo: An in vivo study. In: Yeap KH, Hirasawa K, editors. *Electromagnetic fields and waves.* Norderstedt, Germany: Books on Demand; 2019. doi: [10.5772/intechopen.84704](https://doi.org/10.5772/intechopen.84704).

13. Kuzniar A, Laffeber C, Eppink B, Bezstarosti K, Dekkers D, Woelders H, et al. Semi-quantitative proteomics of mammalian cells upon short-term exposure to non-ionizing electromagnetic fields. *PLoS One*. 2017;**12**(2). e0170762. doi: [10.1371/journal.pone.0170762](https://doi.org/10.1371/journal.pone.0170762). [PubMed: [28234898](https://pubmed.ncbi.nlm.nih.gov/28234898/)]. [PubMed Central: [PMC5325209](https://pubmed.ncbi.nlm.nih.gov/PMC5325209/)].
14. Hosseinabadi MB, Khanjani N, Samaei SE, Nazarkhani F. Effect of long-term occupational exposure to extremely low-frequency electromagnetic fields on proinflammatory cytokine and hematological parameters. *Int J Radiat Biol*. 2019;**95**(11):1573-80. doi: [10.1080/09553002.2019.1642542](https://doi.org/10.1080/09553002.2019.1642542). [PubMed: [31329007](https://pubmed.ncbi.nlm.nih.gov/31329007/)].
15. Mareel M, Leroy A. Clinical, cellular, and molecular aspects of cancer invasion. *Physiol Rev*. 2003;**83**(2):337-76. doi: [10.1152/physrev.00024.2002](https://doi.org/10.1152/physrev.00024.2002). [PubMed: [12663862](https://pubmed.ncbi.nlm.nih.gov/12663862/)].
16. Khalighinejad N, Hariri H, Behnamfar O, Yousefi A, Momeni A. Adenoviral gene therapy in gastric cancer: A review. *World J Gastroenterol*. 2008;**14**(2):180-4. doi: [10.3748/wjg.14.180](https://doi.org/10.3748/wjg.14.180). [PubMed: [18186552](https://pubmed.ncbi.nlm.nih.gov/18186552/)]. [PubMed Central: [PMC2675111](https://pubmed.ncbi.nlm.nih.gov/PMC2675111/)].
17. Wang D, Sadee W. Searching for polymorphisms that affect gene expression and mRNA processing: example ABCB1 (MDR1). *AAPS J*. 2006;**8**(3):E515-20. doi: [10.1208/aapsj080361](https://doi.org/10.1208/aapsj080361). [PubMed: [17025270](https://pubmed.ncbi.nlm.nih.gov/17025270/)]. [PubMed Central: [PMC2761059](https://pubmed.ncbi.nlm.nih.gov/PMC2761059/)].
18. Brown DM, Lee HC, Liu S, Quick CM, Fernandes LM, Simmen FA, et al. Notch-1 signaling activation and progesterone receptor expression in ectopic lesions of women with endometriosis. *J Endocr Soc*. 2018;**2**(7):765-78. doi: [10.1210/je.2018-00007](https://doi.org/10.1210/je.2018-00007). [PubMed: [30151432](https://pubmed.ncbi.nlm.nih.gov/30151432/)]. [PubMed Central: [PMC6106104](https://pubmed.ncbi.nlm.nih.gov/PMC6106104/)].
19. Katoh M, Katoh M. Notch signaling in gastrointestinal tract (review). *Int J Oncol*. 2007;**30**(1):247-51. [PubMed: [17143535](https://pubmed.ncbi.nlm.nih.gov/17143535/)].
20. Kay SK, Harrington HA, Shepherd S, Brennan K, Dale T, Osborne JM, et al. The role of the Hesi crosstalk hub in Notch-Wnt interactions of the intestinal crypt. *PLoS Comput Biol*. 2017;**13**(2). e1005400. doi: [10.1371/journal.pcbi.1005400](https://doi.org/10.1371/journal.pcbi.1005400). [PubMed: [28245235](https://pubmed.ncbi.nlm.nih.gov/28245235/)]. [PubMed Central: [PMC5363986](https://pubmed.ncbi.nlm.nih.gov/PMC5363986/)].
21. Rani A, Greenlaw R, Smith RA, Galustian C. HES1 in immunity and cancer. *Cytokine Growth Factor Rev*. 2016;**30**:113-7. doi: [10.1016/j.cytogfr.2016.03.010](https://doi.org/10.1016/j.cytogfr.2016.03.010). [PubMed: [27066918](https://pubmed.ncbi.nlm.nih.gov/27066918/)].
22. Liu X, Yun F, Shi L, Li ZH, Luo NR, Jia YF. Roles of signaling pathways in the epithelial-mesenchymal transition in cancer. *Asian Pac J Cancer Prev*. 2015;**16**(15):6201-6. doi: [10.7314/apjcp.2015.16.15.6201](https://doi.org/10.7314/apjcp.2015.16.15.6201). [PubMed: [26434817](https://pubmed.ncbi.nlm.nih.gov/26434817/)].
23. Yan B, Liu L, Zhao Y, Xiu LJ, Sun DZ, Liu X, et al. Xiaotan Sanjie decoction attenuates tumor angiogenesis by manipulating Notch-1-regulated proliferation of gastric cancer stem-like cells. *World J Gastroenterol*. 2014;**20**(36):13105-18. doi: [10.3748/wjg.v20.i36.i3105](https://doi.org/10.3748/wjg.v20.i36.i3105). [PubMed: [25278704](https://pubmed.ncbi.nlm.nih.gov/25278704/)]. [PubMed Central: [PMC4177489](https://pubmed.ncbi.nlm.nih.gov/PMC4177489/)].
24. Li P, Chen H, Chen S, Mo X, Li T, Xiao B, et al. Circular RNA 0000096 affects cell growth and migration in gastric cancer. *Br J Cancer*. 2017;**116**(5):626-33. doi: [10.1038/bjc.2016.451](https://doi.org/10.1038/bjc.2016.451). [PubMed: [28081541](https://pubmed.ncbi.nlm.nih.gov/28081541/)]. [PubMed Central: [PMC5344286](https://pubmed.ncbi.nlm.nih.gov/PMC5344286/)].
25. Li P, Chen S, Chen H, Mo X, Li T, Shao Y, et al. Using circular RNA as a novel type of biomarker in the screening of gastric cancer. *Clin Chim Acta*. 2015;**444**:132-6. doi: [10.1016/j.cca.2015.02.018](https://doi.org/10.1016/j.cca.2015.02.018). [PubMed: [25689795](https://pubmed.ncbi.nlm.nih.gov/25689795/)].
26. Mansoury F, Babaei N, Abdi S, Entezari M, Doosti A. Evaluation of the PTEN and circRNA-CDRIAs gene expression changes in gastric cancer and normal cell lines following the exposure to weak and moderate 50 hz electromagnetic fields. *Int J Cancer Manag*. 2021;**14**(4). e11079. doi: [10.5812/ijcm.111079](https://doi.org/10.5812/ijcm.111079).
27. Abdi S, Dorranian D, Naderi GA, Razavi AE. Changes in physico-chemical characteristics of human low density lipoprotein nanoparticles by electromagnetic field exposure. *STUD U BABES-BOL CHE*. 2016;**61**(1):185-97.
28. Abdi S, Dorranian D, Razavi AE, Naderi GA, Boshtam M, Ghorannevis M. Evaluation of the effects of weak and moderate static magnetic fields on the characteristics of human low density lipoprotein in vitro. *Bioelectromagnetics*. 2013;**34**(5):397-404. doi: [10.1002/bem.21779](https://doi.org/10.1002/bem.21779). [PubMed: [23361580](https://pubmed.ncbi.nlm.nih.gov/23361580/)].
29. Mansoury F, Babaei N, Abdi S, Entezari M, Doosti A. Changes in NOTCH1 gene and its regulatory circRNA, hsa_circ_0005986 expression pattern in human gastric adenocarcinoma and human normal fibroblast cell line following the exposure to extremely low frequency magnetic field. *Electromagn Biol Med*. 2021;**40**(3):375-83. doi: [10.1080/15368378.2021.1891092](https://doi.org/10.1080/15368378.2021.1891092). [PubMed: [33620018](https://pubmed.ncbi.nlm.nih.gov/33620018/)].
30. Bahar M, Majd A, Abdi S. Effects of (ELF) extremely low frequency (50 Hz) AC and DC magnetic fields on lentil germination and seedlings growth. *Iran Phys J*. 2009;**3**(2):12-6.
31. Aalami Zavareh F, Abdi S, Entezari M. Up-regulation of miR-144 and miR-375 in the human gastric cancer cell line following the exposure to extremely low-frequency electromagnetic fields. *Int J Radiat Biol*. 2021;**97**(9):1324-32. doi: [10.1080/09553002.2021.1941376](https://doi.org/10.1080/09553002.2021.1941376). [PubMed: [34125651](https://pubmed.ncbi.nlm.nih.gov/34125651/)].
32. McNamee JP, Bellier PV, McLean JR, Marro L, Gajda GB, Thansandote A. DNA damage and apoptosis in the immature mouse cerebellum after acute exposure to a 1 mT, 60 Hz magnetic field. *Mutat Res*. 2002;**513**(1-2):121-33. doi: [10.1016/s1383-5718\(01\)00302-3](https://doi.org/10.1016/s1383-5718(01)00302-3). [PubMed: [11719097](https://pubmed.ncbi.nlm.nih.gov/11719097/)].
33. Lahijani MS, Tehrani DM, Sabouri E. Histopathological and ultrastructural studies on the effects of electromagnetic fields on the liver of preincubated white Leghorn chicken embryo. *Electromagn Biol Med*. 2009;**28**(4):391-413. doi: [10.3109/15368370903287689](https://doi.org/10.3109/15368370903287689). [PubMed: [20017630](https://pubmed.ncbi.nlm.nih.gov/20017630/)].
34. Calvente I, Fernandez MF, Villalba J, Olea N, Nunez MI. Exposure to electromagnetic fields (non-ionizing radiation) and its relationship with childhood leukemia: A systematic review. *Sci Total Environ*. 2010;**408**(16):3062-9. doi: [10.1016/j.scitotenv.2010.03.039](https://doi.org/10.1016/j.scitotenv.2010.03.039). [PubMed: [20451240](https://pubmed.ncbi.nlm.nih.gov/20451240/)].
35. Boorman GA, McCormick DL, Findlay JC, Hailey JR, Gauger JR, Johnson TR, et al. Chronic toxicity/oncogenicity evaluation of 60 Hz (power frequency) magnetic fields in F344/N rats. *Toxicol Pathol*. 1999;**27**(3):267-78. doi: [10.1177/019262339902700301](https://doi.org/10.1177/019262339902700301). [PubMed: [10356702](https://pubmed.ncbi.nlm.nih.gov/10356702/)].
36. Simko M. Induction of cell activation processes by low frequency electromagnetic fields. *ScientificWorldJournal*. 2004;**4** Suppl 2:4-22. doi: [10.1100/tsw.2004.174](https://doi.org/10.1100/tsw.2004.174). [PubMed: [15517098](https://pubmed.ncbi.nlm.nih.gov/15517098/)]. [PubMed Central: [PMC5956406](https://pubmed.ncbi.nlm.nih.gov/PMC5956406/)].
37. Dai X, Wang L, Deivasigamni A, Looi CY, Karthikeyan C, Trivedi P, et al. A novel benzimidazole derivative, MBIC inhibits tumor growth and promotes apoptosis via activation of ROS-dependent JNK signaling pathway in hepatocellular carcinoma. *Oncotarget*. 2017;**8**(8):12831-42. doi: [10.18632/oncotarget.14606](https://doi.org/10.18632/oncotarget.14606). [PubMed: [28086233](https://pubmed.ncbi.nlm.nih.gov/28086233/)]. [PubMed Central: [PMC5355059](https://pubmed.ncbi.nlm.nih.gov/PMC5355059/)].
38. Aggarwal V, Tuli HS, Varol A, Thakral F, Yerer MB, Sak K, et al. Role of reactive oxygen species in cancer progression: Molecular mechanisms and recent advancements. *Biomolecules*. 2019;**9**(11):735. [PubMed: [doi:10.3390/biom9110735](https://pubmed.ncbi.nlm.nih.gov/doi:10.3390/biom9110735/)].
39. Ying M, Wang S, Sang Y, Sun P, Lal B, Goodwin CR, et al. Regulation of glioblastoma stem cells by retinoic acid: Role for Notch pathway inhibition. *Oncogene*. 2011;**30**(31):3454-67. doi: [10.1038/onc.2011.58](https://doi.org/10.1038/onc.2011.58). [PubMed: [21383690](https://pubmed.ncbi.nlm.nih.gov/21383690/)].
40. Zhang Y, Zhang XO, Chen T, Xiang JF, Yin QF, Xing YH, et al. Circular intronic long noncoding RNAs. *Mol Cell*. 2013;**51**(6):792-806. doi: [10.1016/j.molcel.2013.08.017](https://doi.org/10.1016/j.molcel.2013.08.017). [PubMed: [24035497](https://pubmed.ncbi.nlm.nih.gov/24035497/)].
41. Kristensen LS, Hansen TB, Venø MT, Kjems J. Circular RNAs in cancer: Opportunities and challenges in the field. *Oncogene*. 2018;**37**(5):555-65. doi: [10.1038/onc.2017.361](https://doi.org/10.1038/onc.2017.361). [PubMed: [28991235](https://pubmed.ncbi.nlm.nih.gov/28991235/)]. [PubMed Central: [PMC5799710](https://pubmed.ncbi.nlm.nih.gov/PMC5799710/)].
42. Ren S, Lin P, Wang J, Yu H, Lv T, Sun L, et al. Circular RNAs: Promising molecular biomarkers of human aging-related diseases via functioning as an miRNA sponge. *Mol Ther Methods Clin Dev*. 2020;**18**:215-29. doi: [10.1016/j.omtm.2020.05.027](https://doi.org/10.1016/j.omtm.2020.05.027). [PubMed: [32637451](https://pubmed.ncbi.nlm.nih.gov/32637451/)]. [PubMed Central: [PMC7326721](https://pubmed.ncbi.nlm.nih.gov/PMC7326721/)].