



Associations of Uncoupling Protein 2 Ala55Val and Uncoupling Protein 3-55C/T Polymorphisms with Heart Rate Variability in Young Oarsmen-a Pilot Study

Andrey Aleksandrovich Melnikov^{1,*}, Artem Sergeevich Bobylev² and Zoya Semenovna Varfolomeeva¹

¹Department of Theory and Methods of Physical Education and Sport, Cherepovets State University, Cherepovets, Russia

²Physical Education Department, Yaroslavl State Pedagogical University Named After K.D. Ushinsky, Yaroslavl, Russia

*Corresponding author: 162600, Vologda Region, Cherepovets, Sovetsky pr-t., 25, aud. 22, Cherepovets, Russia. Email: melni974@yandex.ru

Received 2018 May 08; Revised 2019 February 04; Accepted 2019 March 14.

Abstract

Background: Human heredity is known to determine about 50% of heart rate variability (HRV) based on a twin study; however, genetic polymorphisms causing increased HRV in athletes are not known.

Objectives: This article is aimed at studying the associations of uncoupling protein 2 (*UCP2*) Ala55Val (rs660339) and *UCP3*-55C/T (rs1800849) polymorphisms with heart rate variability (HRV) in trained oarsmen.

Methods: HRV indices (standard deviations of NN intervals (SDNN), root mean square of successive differences (RMSSD), HF, LF, VLF), stroke index and cardiac index of the young oarsmen (age 17.6 ± 1.6 years old, $n = 23$) were determined by impedance cardiography method in the supine and standing positions. The maximum oxygen consumption (VO_{2max}) was examined with a rower ergometer by means of a gas analyzer. Polymorphisms of the *UCP2* Ala55Val and the *UCP3*-55T/C were genotyped by polymerase chain reaction (PCR) method and length analysis of restriction products.

Results: It was found that both polymorphisms were not associated with VO_{2max} . The *UCP2* Val/Val genotype compared to the combined variant (Ala/Ala + Ala/Val) containing the 55 Ala allele, as well as the *UCP3* T/T genotype compared to the combined variant (C/T + C/C) containing the -55C allele were associated with low HR and increased HRV: SDNN, RMSSD and HF vagal indices in the supine position. Moreover, the athletes with the *UCP3*-55T/T genotype had a pronounced increase in heart rate (ANOVA, $P < 0.001$) and a decrease in stroke index (ANOVA, $P = 0.005$) in response to active orthostasis compared to C/T and C/C genotypes.

Conclusions: The *UCP2* Val/Val and the *UCP3* T/T genotypes may be genetic markers of increased HRV in the highly trained athletes, suggesting an influence of these *UCP2*/*UCP3* polymorphisms on autonomic cardiovascular regulation.

Keywords: *UCP2* Ala55Val, *UCP3*-55T/S, Gene Polymorphisms, Heart Rate Variability, Athletes

1. Background

Heart rate variability (HRV) is a term used to describe the RR-interval oscillation, which is primarily due to both parasympathetic and sympathetic neural control of the heart. It is believed that HRV is a useful noninvasive indicator of cardiac autonomic function with important prognostic value (1).

In sports, an increase in HRV is associated with a high ability of the body to adapt to high physical stimuli (2). On the contrary, a decrease in HRV indices during growing physical exercises indicates incomplete recovery after intensive exercises and even overreaching state (3).

It has been shown that HRV of highly trained athletes is increased (4). Mechanisms of a HRV increase are not fully known. In addition to physical training effects, genetic pre-

disposition to increased HRV is highly important (5). Twin studies show that the inheritance of HRV indexes is about 46% - 57% (5). However, the genetic polymorphisms causing increased HRV remain completely unclear.

Knowledge of molecular genetic factors affecting HRV indices will help to explain consistently high or low HRV during frequent HRV monitoring, as well as to improve the determination of fitness.

Genetic polymorphisms of the uncoupling proteins 2 (*UCP2*) and 3 (*UCP3*) are of particular interest regarding genetic factors determining autonomic regulation of the athletes' hearts. These genes determine the synthesis and activity of uncoupling proteins of the appropriate type that can dissociate oxidation and phosphorylation in mitochondria of different tissues, that is, to change oxida-

tions to increase heat generation, thereby reducing the metabolic efficiency of mitochondria (6, 7). In addition, *UCP2* and *UCP3* proteins are implicated in the formation of reactive oxygen species (ROS) by the mitochondria during respiration, and lowering of the proton gradient across the inner mitochondrial membrane by UCPs results in a reduction in ROS generation (7). Furthermore, these proteins affect insulin sensitivity (8), leptin level (9) and angiotensin-converting enzyme expression (10). All these functions of *UCP2/3* may mediate the effects of these genetic polymorphisms on the autonomous regulation of the heart. Moreover, it was previously established that *UCP2* 45 bp I/D and *UCP3*-55C/T polymorphisms were associated with HRV and blood pressure in untrained men.

2. Objectives

The purpose of this study was to evaluate possible associations of the *UCP2* Ala55Val and *UCP3*-55T/C polymorphisms with HRV of the trained oarsmen. This was a pilot study of the role of the *UCP2/3* genes in the determination of HRV.

3. Methods

3.1. Subjects

Young healthy oarsmen ($n = 23$) who have been rowing regularly for more than 1.5 years in St. Petersburg sports clubs and participating in local and national competitions were invited to participate in the study. The age of the oarsmen was 17.6 ± 1.6 years old; their height: 180.2 ± 8.4 cm; body weight: 75.5 ± 10.4 kg, sports experience of rowing: 4.8 ± 2.9 years, VO_{2max} : 55.4 ± 10.4 mL/min/kg. In the last month before the examination, the athletes trained according to a similar training program so that their relative intensity of loads was about the same, which caused similar changes in the autonomous regulation of the heart. Athletes did not have any abnormalities in the health and regulation of the cardiovascular system. The examination was carried out at the end of the pre-competition period with high training loads.

The study conforms to the code of ethics of the World Medical Association Declaration of Helsinki and ethical standards in sport and exercise science research. All underage (age < 18 years, $n = 12$) athletes' parents and adult athletes gave their written consent after a detailed explanation about the study procedure and risks involved with the investigation.

3.2. Maximum Oxygen Consumption (VO_{2max}) Test

VO_{2max} was determined using an incremental test to exhaustion on a rowing ergometer Weba Sport Slider Kayak ergometer (Austria). The initial stage of the workload was 120 W and increased every two minutes by 75 W. The rest interval between the steps was 30 seconds. Oxygen consumption was recorded by means of a gas analyzer MetaMax 3B (Cortex, Leipzig, Germany). The heart rate was determined using a Polar RS 400sd heart rate monitor. Using the gas analyzer, O_2 and CO_2 contents were measured by the electrochemical and non-dispersive infrared sensors, respectively. The air flow was measured by a turbine transducer. Standard calibrations of gas sensors and transducer were performed before each test. Maximal oxygen consumption was recorded as the highest mean value recorded in the test observed over a 30s period in the final part of the workloads. The criteria used to confirm a maximal test were a decrease in power of more than 30 W from the target power; a respiratory exchange ratio greater than 1.1 and a final heart rate above 95% of the age-related maximum.

3.3. Genotyping

DNA analysis was performed in a specialized genetic laboratory. DNA was extracted from buccal cells using a DNK-sorb-A sorbent kit according to the manufacturer's instruction (Central Research Institute of Epidemiology, Moscow, Russia).

UCP2 Ala55Val (rs660339) and *UCP3*-55T/C (rs1800849) polymorphisms were genotyped by the polymerase chain reaction-restriction fragment length polymorphism method in accordance with studies (11). The sequences of the primers (The Scientific Production Company "LITECH". Moscow) to detect Ala55Val polymorphism were forward 5'-CTGGGAGTCTTGATGGTGCTAC-3' and reverse 5'-CACCGCGGTACTGGGCGTTG-3', those to detect -55C/T polymorphism were forward 5'-GAGCTATATAAAGCACCCCAAGT-3' and reverse 5'-TCTGCTGCTTCTGGCTTGGCACTGGTCTTATACACC-3'. The polymerase chain reactions were carried out as follows. Initial denaturation at 95°C for 5 minutes (1 cycle) followed by 30 cycles of denaturation at 95°C for 60 seconds, annealing at 63°C for 60 seconds, extension at 72°C for 60 seconds and a final extension at 72°C for 5 minutes. Amplicons (199 bp DNA fragments for *UCP2*; 189 bp DNA fragments for *UCP3*) were incubated (for *UCP2* at 37°C and for *UCP3* at 30°C during the night) together with restriction endonucleases (HindII for *UCP2*, SmaI for *UCP3* produced by "SibEnzyme". Novosibirsk, Russia) to identify single nucleotide replacements. Length analysis of restriction products was carried out by electrophoretic

separation of 8% polyacrylamide gel followed by staining with ethidium bromide and visualization in ultraviolet light with a transilluminator. Unrestricted fragments of length 199 bp corresponded to the *UCP2* Ala/Ala genotype, three fragments of length 199, 180, and 19 bp corresponded to the Ala/Val genotype and two fragments of length 180 and 19 bp - the Val/Val genotype. The *UCP3* T/T genotype corresponded to an unrestricted fragment 189 bp long, the CT genotype three fragments 189, 152, and 37 bp long, and the CC genotype two fragments 152 and 37 bp long.

3.4. Analysis of HRV, Cardiac (CI) and Stroke (SI) Indices

Cardiohemodynamic measurements were performed in standard comfort conditions of a single room. After 5 minutes of supine rest, a lead II ECG and tetrapolar rheocardiogram (the active electrodes were placed on the jugular fossa and the xiphoid process) using the impedance cardiography hardware-software complex "Reodin-504" (Medass, Moscow) were simultaneously recorded during spontaneous breathing for 5 minutes in the supine position and during 6 minutes of active orthostasis. The first minute of orthostasis was excluded from the recordings to remove transient processes. Heart rate (HR), SI (mL/m^2), CI ($\text{L}/\text{min}/\text{m}^2$) were calculated using ECG and rheocardiograms.

To estimate HRV, we defined time-domain indices: standard deviations of NN intervals (SDNN), root mean square of successive differences (RMSSD), and power spectral measures (based on the fast Fourier transformation analysis): high-frequency (0.15 - 0.4 Hz, HF, ms^2), low-frequency (0.04 - 0.14 Hz, LF, ms^2) and very low-frequency (less than 0.04 Hz, VLF, ms^2) powers of the RR-intervals oscillations. The normalized powers of HFnu, LFnu as well as LF/HF ratio were calculated (1). "Orto" was added to all studied indexes in order to mark the orthostatic position.

3.5. Statistics

The results are presented as the arithmetic mean \pm standard deviation (SD). The correspondence of the genotypes distribution to the Hardy-Weinberg equilibrium was determined by comparing the observed and expected frequencies by the chi-square test. The associations of HRV, CI, and SI with the studied polymorphisms were assessed by a one-way ANOVA with the Bonferroni criterion for post-hoc comparisons. Student's *t*-test was used for pair comparisons between the groups. Before the ANOVA analysis, HRV indices were converted by logarithm with a base of 10 (Lg).

4. Results

4.1. Distribution of the Athletes' Genotypes and Alleles

Genotype distributions of *UCP2* Ala55Val polymorphism ($\chi^2 = 2.27$, $P = 0.321$) and *UCP3*-55C/T polymorphism ($\chi^2 = 0.631$, $P = 0.729$) in all oarsmen ($n = 23$) were in Hardy-Weinberg equilibrium.

4.2. Associations of HRV and CI Indices with the *UCP2* Ala55Val Polymorphism

4.2.1. The Supine Position

The *UCP2* Ala55Val polymorphism was associated to HR ($P = 0.001$), SDNN ($P = 0.006$), RMSSD ($P = 0.001$), HF ($P = 0.009$), LF ($P = 0.004$), HFnu ($P = 0.047$), LF/HF ($P = 0.044$) and CI ($P = 0.027$), but an association with VO₂max was not significant ($P = 0.078$) (Table 1). Major differences were connected with a lower HR and higher indices of HRV in persons with the Val/Val genotype regarding to the Ala/Val and (Ala/Ala + Ala/Val) groups.

4.2.2. The Orthostatic Position

In the orthostatic position the *UCP2* Ala55Val polymorphism was associated with HR ($P = 0.022$), SDNN ($P = 0.006$), RMSSD ($P = 0.001$) and CI ($P = 0.029$) (Table 2). The athletes with the *UCP2* Val/Val genotype had decreased HR ($P = 0.022$), increased SDNN ($P = 0.060$) and RMSSD ($P = 0.020$), as well as low CI ($P = 0.007$) compared to the (Ala/Ala + Ala/Val) group. The *UCP2* Ala55Val polymorphism was not associated with the responses of HRV, CI, and SI in response to orthostasis (all $P > 0.1$ results not shown).

4.3. Associations of HRV and CI Indices with the *UCP3*-55T/C Polymorphism

4.3.1. The Supine Position

The *UCP3*-55T/C polymorphism was associated with HR ($P = 0.001$), RMSSD ($P = 0.006$), HF ($P = 0.032$), LF ($P = 0.053$), VLF ($P = 0.047$) and SI ($P = 0.085$), there was not a correlation with VO₂max (Table 3). The T/T genotype had lower HR ($P = 0.001$), and higher levels of SDNN ($P = 0.007$), RMSSD ($P = 0.002$), HF ($P = 0.008$), LF ($P = 0.014$), VLF ($P = 0.013$) compared to the (C/T + C/C) group. Comparison of the C/C genotype with the (C/T + T/T) group did not reveal any significant differences (all $P > 0.1$ results not shown).

4.3.2. The Orthostatic Position

In the standing position, the *UCP3*-55T/C polymorphism was associated only with LF ($P = 0.024$), and the indices CI_{orto} ($P = 0.055$), SI_{orto} ($P = 0.089$), SDNN_{orto} ($P = 0.063$) had near significant associations (Table 4). However, the *UCP3*-55T/C polymorphism showed associations with

Table 1. Heart Rate Variability (HRV), Stroke Index (SI) and Cardiac Index (CI) in the Supine Position in the Oarsmen with Different Genotypes of the *UCP2* Ala55Val Polymorphism^{a,b}

	<i>UCP2</i> Ala55Val Genotype				Post-Hoc		
	Ala/Ala (N=3)	Ala/Val (N=15)	Val/Val (N=5)	P Value 1 ^c	P Value 2 ^d	P Value 3 ^e	P Value 4 ^f
VO ₂ max, ml/min/kg	50.5 ± 8.4	53.4 ± 9.0	64.3 ± 11.7	0.078	1.00	0.185	0.116
Lg HR, Lg bpm	1.79 ± 0.07	1.84 ± 0.04	1.70 ± 0.07	0.001	0.448	0.058	0.001
	1.83 ± 0.05		1.70 ± 0.07	0.001			
Lg SDNN, Lg ms	1.89 ± 0.04	1.71 ± 0.17	2.00 ± 0.16	0.006	0.266	1.000	0.007
	1.74 ± 0.17		2.00 ± 0.16	0.006			
Lg RMSSD, Lg ms	1.78 ± 0.12	1.62 ± 0.15	2.01 ± 0.24	0.001	0.459	0.239	0.001
	1.65 ± 0.16		2.01 ± 0.24	0.001			
Lg HF, Lg ms ²	3.16 ± 0.22	3.03 ± 0.43	3.74 ± 0.33	0.009	1.000	0.181	0.008
	3.05 ± 0.40		3.74 ± 0.33	0.002			
Lg LF, Lg ms ²	3.52 ± 0.23	2.96 ± 0.34	3.49 ± 0.30	0.004	0.035	1.000	0.013
	3.05 ± 0.38		3.49 ± 0.30	0.028			
Lg VLF, Lg ms ²	2.86 ± 0.42	2.84 ± 0.48	3.32 ± 0.43	0.149	1.000	0.566	0.171
	2.84 ± 0.46		3.32 ± 0.43	0.048			
Lg LF/HF, Lg %	0.37 ± 0.32	-0.07 ± 0.32	-0.24 ± 0.26	0.044	0.114	0.043	0.890
	0.00 ± 0.36		-0.24 ± 0.26	0.168			
HFnu, %	31.5 ± 14.9	53.9 ± 17.0	62.8 ± 13.5	0.047	0.122	0.046	0.899
	50.2 ± 18.4		62.8 ± 13.5	0.170			
LFnu, %	68.5 ± 14.9	46.1 ± 17.0	37.2 ± 13.5	0.047	0.122	0.046	0.899
	49.8 ± 18.4		37.2 ± 13.5	0.170			
CI, l/min/m ²	3.52 ± 0.57	3.62 ± 0.75	2.60 ± 0.45	0.027	1.00	0.235	0.025
	3.60 ± 0.70		2.60 ± 0.45	0.007			
SI, ml/m ²	57 ± 12	53 ± 11	53 ± 13	0.819	1.00	1.00	1.00
	54 ± 11		53 ± 13	0.868			

Abbreviations: HR, heart rate; RMSSD, root mean square of successive differences; SDNN, standard deviations of NN intervals.

^aValues are expressed as mean ± SD.

^bLg, logarithm with a base of 10.

^cSignificance level for ANOVA.

^dAla/Ala vs. Ala/Val.

^eAla/Ala vs. Val/Val.

^fAla/Val vs. Val/Val.

HR response (mean ± SD: T/T = 59 ± 11%, C/T = 10 ± 15%, C/C = 24 ± 7%, ANOVA: P < 0.001; T/T vs. C/T P < 0.001; T/T vs. C/C P = 0.002; C/T vs. C/C P = 0.122) and SI response (ANOVA: P = 0.005; Figure 1) to active orthostasis.

5. Discussion

This study evaluated the associations of the *UCP2* Ala55Val and the *UCP3*-55C/T polymorphisms with HRV and cardiac index (CI) in supine and standing positions in the young oarsmen. Our main findings showed that higher HRV and lower HR was associated with *UCP2* Val/Val and

UCP3 T/T genotypes in a supine position and less significant in orthostasis.

It is believed that the *UCP2* Val/Val genotype is associated with low uncoupling activity of mitochondria, increased metabolic efficiency during rest and exercise, reduced fat oxidation and higher spontaneous physical activity (12, 13). Although we did not reveal a significant correlation of the polymorphism with aerobic capacity, however, there was a tendency for increased VO₂max (P = 0.078) in the Val/Val genotype, which is consistent with the works (11, 13, 14), which showed the positive effect of the Val allele on physical performance.

Table 2. Heart Rate Variability (HRV), Stroke Index (SI) and Cardiac Index (CI) in the Standing Position in the Oarsmen with Different Genotypes of the *UCP2* Ala55Val Polymorphism (Mean \pm SD)^{a,b}

	UCP2 Ala55Val Genotype				Post-Hoc		
	Ala/Ala (N = 3)	Ala/Val (N = 15)	Val/Val (N = 5)	P Value 1 ^c	P Value 2 ^d	P Value 3 ^e	P Value 4 ^f
Ig HRortho, Ig bpm	1.84 \pm 0.08	1.90 \pm 0.07	1.81 \pm 0.06	0.022	0.412	1.00	0.027
	1.89 \pm 0.07		1.81 \pm 0.06	0.019			
Ig SDNNortho, Ig ms	1.83 \pm 0.13	1.63 \pm 0.13	1.83 \pm 0.13	0.006	0.056	1.00	0.018
	1.66 \pm 0.15		1.83 \pm 0.13	0.030			
Ig HFortho, Ig ms²	2.83 \pm 0.52	2.73 \pm 0.49	3.03 \pm 0.30	0.474	1.00	1.00	0.685
	2.75 \pm 0.48		3.03 \pm 0.30	0.235			
Ig LFortho, Ig ms²	3.46 \pm 0.39	3.14 \pm 0.41	3.33 \pm 0.37	0.379	0.668	1.00	1.00
	3.19 \pm 0.41		3.33 \pm 0.37	0.513			
Ig VLFortho, Ig ms²	2.72 \pm 0.15	2.68 \pm 0.38	2.97 \pm 0.18	0.241	1.00	0.885	0.291
	2.69 \pm 0.34		2.97 \pm 0.18	0.089			
Ig LF/Hfortho, Ig %	0.63 \pm 0.59	0.41 \pm 0.37	0.30 \pm 0.47	0.570	1.00	0.888	1.00
	0.44 \pm 0.40		0.30 \pm 0.47	0.500			
Ig RMSSDortho, Ig ms	1.65 \pm 0.21	1.38 \pm 0.15	1.70 \pm 0.17	0.001	0.041	1.00	0.003
	1.42 \pm 0.19		1.70 \pm 0.17	0.008			
HFnu(ortho)	24.3 \pm 21.1	30.5 \pm 16.4	36.3 \pm 22.2	0.665	1.00	1.00	1.00
	29.5 \pm 16.7		36.3 \pm 22.2	0.462			
LFnu(ortho)	75.7 \pm 21.1	69.5 \pm 16.4	63.7 \pm 22.2	0.665	1.00	1.00	1.00
	70.5 \pm 16.7		63.7 \pm 22.2	0.462			
Clortho, L/min/m²	2.94 \pm 0.78	2.86 \pm 0.82	1.75 \pm 0.54	0.029	1.00	0.139	0.032
	2.87 \pm 0.79		1.75 \pm 0.54	0.008			
Slortho, mL/m²	43 \pm 15	36 \pm 10	28 \pm 11	0.156	0.814	0.187	0.523
	37 \pm 11		28 \pm 11	0.111			

Abbreviations: HR, heart rate; RMSSD, root mean square of successive differences; SDNN, standard deviations of NN intervals.

^aValues are expressed as mean \pm SD.

^bIg, logarithm with a base of 10.

^cSignificance level for ANOVA.

^dAla/Ala vs. Ala/Val.

^eAla/Ala vs. Val/Val.

^fAla/Val vs Val/Val.

The results of our study showed that the positive effect of the Val/Val genotype may be associated with the activation of cardiac vagus, because athletes with Val/Val genotype had higher HRV.

The mechanism of the associations is poorly understood, since reduced *UCP2* activity associated with Val/Val genotype is more likely correlated with increased ROS formation (7), and higher activity of the angiotensin-converting enzyme (10) which have sympathetic effects. It is known that higher ROS levels (15) and activity of the renin-angiotensin system (16) are associated with an increased sympathetic activity and decreased HRV. It is also shown (17) that *UCP2* 45 bp I/I genotype having low *UCP2* ac-

tivity was associated with lower HRV.

We suppose that in our study increased cardiac parasympathetic activities in the Val/Val genotype group may be due to higher insulin sensitivity (8) or lower leptin levels (9). In some studies were shown that both factors are associated to Val/Val genotype and increased HRV indices (18, 19). However, one cannot exclude the possibility that the associations of Val allele to higher HRV could be due to other polymorphisms that is in the linkage disequilibrium with Ala55Val polymorphism (20). Generally, physiologic mechanisms of increased parasympathetic activities in the Val/Val genotype carriers require further confirmation and research.

Table 3. Heart Rate Variability (HRV), Stroke Index (SI) and Cardiac Index (CI) in the Supine Position in the Oarsmen with Different Genotypes of the *UCP3-55T/C* Polymorphism^{a, b}

	UCP3-55T/C Genotype				Post-Hoc		
	TT (N = 3)	CT (N = 13)	CC (N = 7)	P Value 1 ^c	P Value 2 ^d	P Value 3 ^e	P Value 4 ^f
VO2max, ml/min/kg	63.5 ± 1.2	56.0 ± 12.3	50.7 ± 5.5	0.196	0.758	0.237	0.833
Lg HR, Lg bpm	1.66 ± 0.06	1.82 ± 0.05	1.81 ± 0.05	0.001	0.001	0.001	1.00
	1.66 ± 0.06	1.82 ± 0.05		0.001			
Lg RMSSD, Lg ms	2.08 ± 0.14	1.65 ± 0.014	1.73 ± 0.27	0.006	0.005	0.039	1.00
	2.08 ± 0.14	1.68 ± 0.19		0.002			
Lg SDNN, Lg ms	2.07 ± 0.13	1.75 ± 0.017	1.78 ± 0.19	0.027	0.025	0.070	1.00
	2.07 ± 0.13	1.76 ± 0.17		0.007			
Lg HF, Lg ms ²	3.85 ± 0.019	3.11 ± 0.43	3.08 ± 0.47	0.032	0.039	0.048	1.00
	3.85 ± 0.019	3.10 ± 0.43		0.008			
Lg LF, Lg ms ²	3.67 ± 0.06	3.06 ± 0.32	3.08 ± 0.049	0.053	0.056	0.101	1.00
	3.67 ± 0.06	3.07 ± 0.38		0.014			
Lg VLF, Lg ms ²	3.57 ± 0.51	2.88 ± 0.46	2.80 ± 0.35	0.047	0.069	0.058	1.00
	3.57 ± 0.51	2.85 ± 0.42		0.013			
Lg LF/HF, Lg %	-0.18 ± 0.15	-0.05 ± 0.28	0.01 ± 0.051	0.749	1.00	1.00	1.00
	-0.18 ± 0.15	-0.03 ± 0.37		0.488			
HFnu, %	60.3 ± 8.3	53.0 ± 15.3	49.7 ± 25.6	0.713	1.00	1.00	1.00
	60.3 ± 8.3	51.8 ± 18.9		0.460			
LFnu, %	39.7 ± 8.3	47.0 ± 15.3	50.3 ± 25.6	0.713	1.00	1.00	1.00
	39.7 ± 8.3	48.2 ± 18.9		0.460			
CI, l/min/m ²	2.86 ± 0.40	3.32 ± 0.72	3.72 ± 0.91	0.262	1.00	0.354	0.828
	2.86 ± 0.40	3.46 ± 0.79		0.221			
SI, ml/m ²	63 ± 10	49 ± 9	57 ± 13	0.085	0.161	1.00	0.338
	63 ± 10	52 ± 11		0.121			

Abbreviations: HR, heart rate; RMSSD, root mean square of successive differences; SDNN, standard deviations of NN intervals.

^aValues are expressed as mean ± SD.

^bLg, logarithm with a base of 10.

^cSignificance level of differences for ANOVA.

^dT/T vs. C/T.

^eT/T vs. C/C.

^fC/T vs. C/C.

Besides the *UCP2* Val/Val genotype, the *UCP3-55T/T* genotype was associated to lower HR and increased HRV in our athletes. In contrast to the *UCP2* Val/Val genotype, the *UCP3-55T/T* genotype contributes to the higher expression of *UCP3* generally in muscles disrupting oxidation and phosphorylation inducing decreased ATP synthesis and ROS formation (21).

A number of mechanisms can be proposed to explain increased HRV in athletes with the *UCP3* T/T genotype. Increased cardiac vagus tone in *UCP3* T/T athletes may be associated with low angiotensin 2 activity, increased insulin sensitivity or low ROS levels. Firstly, it was shown that angiotensin-converting enzyme activity (10), blood

glucose and insulin resistance (8) and presumably tissue ROS levels (7) may be reduced in carriers of the *UCP3* T/T genotype. Secondly, reduced angiotensin 2 levels (16), high insulin sensitivity (19) and low ROS levels (15), as a rule, are correlated with elevated HRV. Moreover, similar associations between HRV and *UCP3-55T/C* polymorphism have been established in the work (17) which showed increased parasympathetic HRV indices in carriers of the -55T allele compared to carriers of the *UCP3* C/C genotype. However, all these mechanisms are speculative and require further research.

Another feature of the *UCP3-55T/C* polymorphism is the association of the T/T genotype with an increased response

Table 4. Heart Rate Variability (HRV), Stroke Index (SI) and Cardiac Index (CI) in the Standing Position in the Oarsmen with Different Genotypes of the *UCP3-55T/C* Polymorphism^{a,b}

	<i>UCP3-55T/C</i> Genotype				Post-Hoc		
	TT (N = 3)	CT (N = 13)	CC (N = 7)	P Value 1 ^c	P Value 2 ^d	P Value 3 ^e	P Value 4 ^f
Ig HRortho, Ig bpm	1.86 ± 0.04	1.86 ± 0.09	1.90 ± 0.06	0.546	1.00	1.00	0.902
	1.86 ± 0.04	1.88 ± 0.08		0.737			
Ig RMSSDortho, Ig ms	1.56 ± 0.28	1.43 ± 0.18	1.55 ± 0.24	0.408	1.00	1.00	0.741
	1.56 ± 0.28	1.47 ± 0.21		0.512			
Ig SDNNortho, Ig ms	1.83 ± 0.16	1.63 ± 0.14	1.76 ± 0.15	0.063	0.134	1.00	0.245
	1.83 ± 0.16	1.68 ± 0.15		0.116			
Ig HFortho, Ig ms²	3.12 ± 0.33	2.78 ± 0.47	2.74 ± 0.48	0.474	0.806	0.744	1.00
	3.12 ± 0.33	2.76 ± 0.46		0.222			
Ig LFortho, Ig ms²	3.75 ± 0.09	3.08 ± 0.36	3.26 ± 0.38	0.024	0.022	0.166	0.858
	3.75 ± 0.09	3.14 ± 0.37		0.011			
Ig VLFortho, Ig ms²	3.00 ± 0.05	2.71 ± 0.40	2.71 ± 0.24	0.391	0.579	0.654	1.00
	3.00 ± 0.05	2.71 ± 0.34		0.166			
Ig LF/Hortho, Ig %	0.63 ± 0.24	0.30 ± 0.37	0.52 ± 0.50	0.318	0.636	1.00	0.760
	0.63 ± 0.24	0.38 ± 0.42		0.323			
Ig RMSSDortho, Ig ms	1.56 ± 0.28	1.43 ± 0.18	1.55 ± 0.24	0.408	1.00	1.00	0.741
	1.56 ± 0.28	1.47 ± 0.21		0.512			
HFnu(ortho)	19.8 ± 9.3	35.3 ± 17.7	27.7 ± 19.5	0.351	0.558	1.00	1.00
	19.8 ± 9.3	32.6 ± 18.2		0.251			
LFnu(ortho)	80.2 ± 9.3	64.7 ± 17.7	72.3 ± 19.5	0.351	0.558	1.00	1.00
	80.2 ± 9.3	67.4 ± 18.2		0.251			
Clortho, L/min/m²	1.68 ± 0.90	2.60 ± 0.65	3.09 ± 0.97	0.055	0.257	0.053	0.593
	1.68 ± 0.90	2.77 ± 0.79		0.041			
Slortho, mL/m²	23 ± 10	36 ± 9	39 ± 13	0.089	0.184	0.095	1.00
	23 ± 10	37 ± 10		0.034			

Abbreviations: HR, heart rate; RMSSD, root mean square of successive differences; SDNN, standard deviations of NN intervals.

^aValues are expressed as mean ± SD.

^bIg, logarithm with a base of 10.

^cSignificance level of differences for ANOVA.

^dT/T vs. C/T.

^eT/T vs. C/C.

^fC/T vs. C/C.

of HR and SI to orthostasis. This may indicate orthostatic intolerance in athletes with the T/T genotype. We cannot specify the effects of the *UCP3* T/T genotype, which may contribute to a decrease in orthostatic stability. However, taking into account the mechanisms of development of orthostatic intolerance among athletes (22), it can be assumed that the involvement of the *UCP3* T/T genotype in reducing ROS and angiotensin 2 levels, linked anti-inflammatory activity and other pleiotropic effects of this genotype may contribute to increased dilatation and compliance of the vascular system and myocardium (eccentric hypertrophy)

(23) and lead to a side orthostatic intolerance in athletes with the T/T genotype. Thus, we believe that the associations of the T/T genotype with higher SI and HR orthostatic responses may reflect the involvement of this genotype in increasing compliance and dilatation of the vascular wall and myocardium induced by physical exercises.

Despite the lack of associations between the *UCP2* and *UCP3* polymorphisms with VO₂max, our data are consistent with the works (11, 14) and indicate that *UCP2* 55Val and *UCP3-55T* alleles can positively affect athletes' adaptation to high physical exertion, because these alleles were asso-

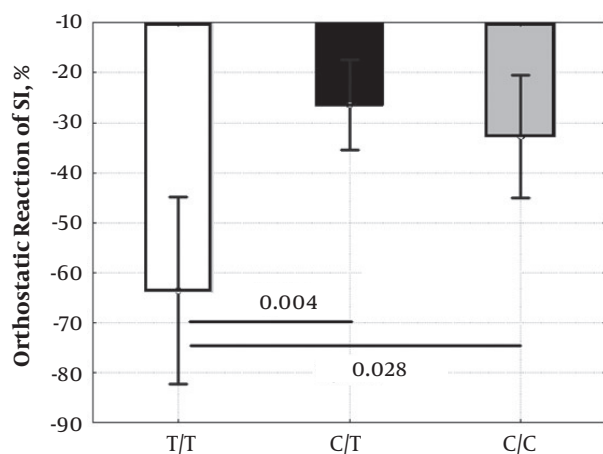


Figure 1. The association of the *UCP3*-55T/C polymorphism with SI reactions to active orthostasis. The SI reaction of the athletes with the T/T genotype was higher than that of the subjects with the C/T ($P=0.004$) and the C/C ($P=0.028$) genotypes (mean \pm D.I.).

ciated with higher cardiac vagus tone, which is a favorable marker of high athletic performance.

5.1. Limitations

Due to the small number of the athletes surveyed, especially with *UCP2* Ala/Ala and *UCP3* T/T genotypes, the statistical power of this study is moderate, so, at the current time it requires caution when generalizing the results obtained. In addition, further studies with a larger number of subjects are required to understand the associations between *UCP2* and *UCP3* polymorphisms and cardiac autonomic regulation.

5.2. Conclusions

The findings of our pilot study show that the athletes with the *UCP2* Val/Val and the *UCP3* T/T genotypes have increased HRV suggesting an influence of *UCP2*/*UCP3* polymorphisms on autonomic cardiovascular regulation. Consequently, these genotypes, often found in athletes and associated with high athletic and physical performance, can at least in part constitute a genetic basis that explains the increased HRV in highly trained athletes. However, the role of the *UCP2* Ala55Val and the *UCP3*-55T/C polymorphisms in determining HRV needs further clarification.

Acknowledgments

The authors are grateful to the Helix genetic analysis laboratory personnel (St. Petersburg) for the technical assistance that they provided in carrying out the genetic studies.

Footnotes

Authors' Contribution: Study concept and design: Andrey Aleksandrovich Melnikov and Artem Sergeevich Bobylev. Acquisition of data: Artem Sergeevich Bobylev. Analysis and interpretation of data: Andrey Aleksandrovich Melnikov, Artem Sergeevich Bobylev and Zoya Semenovna Varfolomeeva. Drafting of the manuscript: Andrey Aleksandrovich Melnikov and Zoya Semenovna Varfolomeeva. Critical revision of the manuscript for important intellectual content: Andrey Aleksandrovich Melnikov and Zoya Semenovna Varfolomeeva. Statistical analysis: Andrey Aleksandrovich Melnikov and Artem Sergeevich Bobylev. Administrative, technical, and material support: Artem Sergeevich Bobylev. Zoya Semenovna Varfolomeeva. Study supervision: Andrey Aleksandrovich Melnikov.

Conflict of Interests: The authors Andrey Aleksandrovich Melnikov, Artem Sergeevich Bobylev and Zoya Semenovna Varfolomeeva have no financial interests related to the material in the manuscript.

Ethical Approval: The study was approved by the Ethical Committee of the Cherepovets State University. None be declared.

Funding/Support: Genetic Laboratory Helix (St. Petersburg, Russia) provided genetic research support for this study.

References

- [No Authors Listed]. Heart rate variability: Standards of measurement, physiological interpretation and clinical use. Task force of the European Society of Cardiology and the North American Society of Pacing and Electrophysiology. *Circulation*. 1996;**93**(5):1043-65. [PubMed: 8598068].
- Bellenger CR, Fuller JT, Thomson RL, Davison K, Robertson EY, Buckley JD. Monitoring athletic training status through autonomic heart rate regulation: A systematic review and meta-analysis. *Sports Med*. 2016;**46**(10):1461-86. doi: 10.1007/s40279-016-0484-2. [PubMed: 26888648].
- Uusitalo AL, Uusitalo AJ, Rusko HK. Heart rate and blood pressure variability during heavy training and overtraining in the female athlete. *Int J Sports Med*. 2000;**21**(1):45-53. doi: 10.1055/s-2000-8853. [PubMed: 10683099].
- da Silva VP, de Oliveira NA, Silveira H, Mello RG, Deslandes AC. Heart rate variability indexes as a marker of chronic adaptation in athletes: A systematic review. *Ann Noninvasive Electrocardiol*. 2015;**20**(2):108-18. doi:10.1111/anec.12237. [PubMed: 25424360].
- Neijts M, Van Lien R, Kupper N, Boomsma D, Willemsen G, de Geus EJ. Heritability of cardiac vagal control in 24-h heart rate variability recordings: Influence of ceiling effects at low heart rates. *Psychophysiology*. 2014;**51**(10):1023-36. doi: 10.1111/psyp.12246. [PubMed: 24894483].
- Affourtit C, Crichton PG, Parker N, Brand MD. *Novel uncoupling proteins*. 287. Wiley Online Library; 2008. p. 70-91. doi: 10.1002/9780470725207.ch6.

7. Brand MD, Affourtit C, Esteves TC, Green K, Lambert AJ, Miwa S, et al. Mitochondrial superoxide: Production, biological effects, and activation of uncoupling proteins. *Free Radic Biol Med.* 2004;**37**(6):755–67. doi: [10.1016/j.freeradbiomed.2004.05.034](https://doi.org/10.1016/j.freeradbiomed.2004.05.034). [PubMed: [15304252](https://pubmed.ncbi.nlm.nih.gov/15304252/)].
8. Vimalleswaran KS, Radha V, Ghosh S, Majumder PP, Sathyanarayana Rao MR, Mohan V. Uncoupling protein 2 and 3 gene polymorphisms and their association with type 2 diabetes in asian indians. *Diabetes Technol Ther.* 2011;**13**(1):19–25. doi: [10.1089/dia.2010.0091](https://doi.org/10.1089/dia.2010.0091). [PubMed: [21175267](https://pubmed.ncbi.nlm.nih.gov/21175267/)].
9. Rance KA, Johnstone AM, Murison S, Duncan JS, Wood SG, Speakman JR. Plasma leptin levels are related to body composition, sex, insulin levels and the A55V polymorphism of the UCP2 gene. *Int J Obes (Lond).* 2007;**31**(8):1311–8. doi: [10.1038/sj.ijo.0803535](https://doi.org/10.1038/sj.ijo.0803535). [PubMed: [17342078](https://pubmed.ncbi.nlm.nih.gov/17342078/)].
10. Dhamrait SS, Maubaret C, Pedersen-Bjergaard U, Brull DJ, Gohlke P, Payne JR, et al. Mitochondrial uncoupling proteins regulate angiotensin-converting enzyme expression: Crosstalk between cellular and endocrine metabolic regulators suggested by RNA interference and genetic studies. *Inside Cell.* 2016;**1**(1):70–81. doi: [10.1002/icl3.1019](https://doi.org/10.1002/icl3.1019). [PubMed: [27347560](https://pubmed.ncbi.nlm.nih.gov/27347560/)]. [PubMed Central: [PMC4915277](https://pubmed.ncbi.nlm.nih.gov/PMC4915277/)].
11. Akhmetov I, Popov DV, Astratenkova IV, Druzhevskaya AM, Missina SS, Vinogradova OL, et al. [Using molecular genetic methods for prognosis of aerobic and anaerobic performance in athletes]. *Fiziol Cheloveka.* 2008;**34**(3):86–91. Russian. [PubMed: [18677952](https://pubmed.ncbi.nlm.nih.gov/18677952/)].
12. Astrup A, Toubro S, Dalgaard LT, Urhammer SA, Sorensen TI, Pedersen O. Impact of the v/v 55 polymorphism of the uncoupling protein 2 gene on 24-h energy expenditure and substrate oxidation. *Int J Obes Relat Metab Disord.* 1999;**23**(10):1030–4. [PubMed: [10557023](https://pubmed.ncbi.nlm.nih.gov/10557023/)].
13. Buemann B, Schierner B, Toubro S, Bibby BM, Sorensen T, Dalgaard L, et al. The association between the val/ala-55 polymorphism of the uncoupling protein 2 gene and exercise efficiency. *Int J Obes Relat Metab Disord.* 2001;**25**(4):467–71. [PubMed: [11319648](https://pubmed.ncbi.nlm.nih.gov/11319648/)].
14. Ahmetov I, Williams AG, Popov DV, Lyubaeva EV, Hakimullina AM, Fedotovskaya ON, et al. The combined impact of metabolic gene polymorphisms on elite endurance athlete status and related phenotypes. *Hum Genet.* 2009;**126**(6):751–61. doi: [10.1007/s00439-009-0728-4](https://doi.org/10.1007/s00439-009-0728-4). [PubMed: [19653005](https://pubmed.ncbi.nlm.nih.gov/19653005/)].
15. Danson EJ, Paterson DJ. Reactive oxygen species and autonomic regulation of cardiac excitability. *J Cardiovasc Electrophysiol.* 2006;**17** Suppl 1:S104–12. doi: [10.1111/j.1540-8167.2006.00391.x](https://doi.org/10.1111/j.1540-8167.2006.00391.x). [PubMed: [16686664](https://pubmed.ncbi.nlm.nih.gov/16686664/)].
16. Mann MC, Exner DV, Hemmelgarn BR, Turin TC, Sola DY, Ahmed SB. Impact of gender on the cardiac autonomic response to angiotensin II in healthy humans. *J Appl Physiol (1985).* 2012;**112**(6):1001–7. doi: [10.1152/jappphysiol.01207.2011](https://doi.org/10.1152/jappphysiol.01207.2011). [PubMed: [22223455](https://pubmed.ncbi.nlm.nih.gov/22223455/)].
17. Matsunaga T, Gu N, Yamazaki H, Tsuda M, Adachi T, Yasuda K, et al. Association of UCP2 and UCP3 polymorphisms with heart rate variability in Japanese men. *J Hypertens.* 2009;**27**(2):305–13. doi: [10.1097/HJH.0b013e32831ac967](https://doi.org/10.1097/HJH.0b013e32831ac967). [PubMed: [19155787](https://pubmed.ncbi.nlm.nih.gov/19155787/)].
18. Van De Wielle R, Michels N. Longitudinal associations of leptin and adiponectin with heart rate variability in children. *Front Physiol.* 2017;**8**:498. doi: [10.3389/fphys.2017.00498](https://doi.org/10.3389/fphys.2017.00498). [PubMed: [28747890](https://pubmed.ncbi.nlm.nih.gov/28747890/)]. [PubMed Central: [PMC5506193](https://pubmed.ncbi.nlm.nih.gov/PMC5506193/)].
19. Saito I, Maruyama K, Eguchi E, Kato T, Kawamura R, Takata Y, et al. Low heart rate variability and sympathetic dominance modifies the association between insulin resistance and metabolic syndrome- The toon health study. *Circ J.* 2017;**81**(10):1447–53. doi: [10.1253/circ.CJ-17-0192](https://doi.org/10.1253/circ.CJ-17-0192). [PubMed: [28566656](https://pubmed.ncbi.nlm.nih.gov/28566656/)].
20. Wang H, Chu WS, Lu T, Hasstedt SJ, Kern PA, Elbein SC. Uncoupling protein-2 polymorphisms in type 2 diabetes, obesity, and insulin secretion. *Am J Physiol Endocrinol Metab.* 2004;**286**(1):E1–7. doi: [10.1152/ajpendo.00231.2003](https://doi.org/10.1152/ajpendo.00231.2003). [PubMed: [12915397](https://pubmed.ncbi.nlm.nih.gov/12915397/)].
21. Schrauwen P, Xia J, Walder K, Snitker S, Ravussin E. A novel polymorphism in the proximal UCP3 promoter region: Effect on skeletal muscle UCP3 mRNA expression and obesity in male non-diabetic Pima Indians. *Int J Obes Relat Metab Disord.* 1999;**23**(12):1242–5. [PubMed: [10643679](https://pubmed.ncbi.nlm.nih.gov/10643679/)].
22. Levine BD, Lane LD, Buckley JC, Friedman DB, Blomqvist CG. Left ventricular pressure-volume and Frank-Starling relations in endurance athletes. Implications for orthostatic tolerance and exercise performance. *Circulation.* 1991;**84**(3):1016–23. [PubMed: [1884438](https://pubmed.ncbi.nlm.nih.gov/1884438/)].
23. Lang H, Xiang Y, Ai Z, You Z, Jin X, Wan Y, et al. UCP3 ablation exacerbates high-salt induced cardiac hypertrophy and cardiac dysfunction. *Cell Physiol Biochem.* 2018;**46**(4):1683–92. doi: [10.1159/000489244](https://doi.org/10.1159/000489244). [PubMed: [29694982](https://pubmed.ncbi.nlm.nih.gov/29694982/)].