

## Review Article



# Prospects of Saffron and its Derivatives in Alzheimer's Disease

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## Abstract

Alzheimer's disease (AD) is a progressive neurodegenerative disorder and the most common form of dementia in the old age population, making it a worldwide concern. Unfortunately, few drugs have been presented for treatment of mild and moderate AD. To meet this need, more effective anti-AD agents are emerging. Accumulating evidence supports the beneficial roles of natural-based products in brain function, neurotransmission, neurogenesis, synaptogenesis, and the prevention of amyloid fibrillation and neuronal injury. Several *in vitro*, preclinical, and clinical studies suggest that saffron (its bioactive compounds) is a potential nutraceutical with antioxidant, radical scavenging, anti-inflammatory, hypolipidemic, hypotensive, neuroendocrine, and neuroprotective effects. It has also been proposed that saffron may delay the onset of AD, prevent its progression or help to attenuate the symptoms of the disease. Therefore, we performed a comprehensive search on this plant and its derivatives for AD treatment. Saffron and its active constituents interfere with AD by improving learning behavior, spatial memory, and cognitive function; protecting against neuronal loss; inhibiting beta-amyloid aggregation and neurotoxicity; preventing senile plaques and neurofibrillary tangle (NFT) formation; suppressing the acetylcholinesterase (AChE) activity; and reducing neuroinflammation. Given conclusive scientific findings, saffron and its derivatives might counter neurodegenerative diseases through multiple pathways. Further clinical trials are expected to confirm the neuroprotective properties of this herb and also to translate such findings to improve patients' outcomes.

**Keywords:** Acetylcholinesterase inhibitors, Amyloid beta, Apolipoprotein E, Neurofibrillary tangles, Saffron, Alzheimer's disease

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## Introduction

Alzheimer's disease (AD) is the main etiology of memory loss, prompting irreversible progressive impairments in cognition and memory.<sup>1</sup> Both the incidence and prevalence of AD are increasing worldwide. It is estimated that in 2050, one out of 85 individuals will suffer from AD.<sup>2</sup> Thus far, several mechanisms have been proposed for AD induction or progression. The cholinergic hypothesis refers to reduction of acetylcholine (ACh) in the central cortex of the brain – an area that involves functional skills.<sup>3</sup> Hence, acetylcholinesterase (AChE) inhibitors like donepezil have been found effective in improving mild and moderate to severe AD symptoms.<sup>3</sup> The amyloid cascade hypothesis (ACH) suggests a lack of balance between production and clearance of amyloid-beta (A $\beta$ ), which leads to nerve cell dysfunction and death.<sup>4</sup> On the other

hand, intracellular neurofibrillary tangles (NFTs) block neurotransmitters and cause neuronal cell death.<sup>5</sup> Tau oligomers are accumulated in  $\beta$ -sheet conformation and produce NFTs.<sup>6</sup> Also, high concentration of A $\beta$  triggers NFT formation, and accumulation of NFTs in neurons lead to cell death.<sup>7</sup> Another hypothesis points to the fact that the  $\epsilon 4$  and  $\epsilon 3$  carriers of the apolipoprotein E gene (*APOE*) are more prone to AD; notwithstanding, the  $\epsilon 4$  allele is the main genetic risk factor for late-onset AD. Apolipoprotein E (ApoE) is known to regulate lipid and protein homeostasis in the brain.<sup>8–10</sup> In addition to ApoE, several other genes have also been implicated in AD such as polymorphisms in sortilin related receptor 1 (SORL1), clusterin, complement component receptor 1 (CCR1), Cluster of differentiation 2 associated protein (CD2AP), Cluster of differentiation 33 (CD33), Ephrin type-A

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receptor 1 (EPHA1), and Membrane spanning 4-domains A6E (MS4A4/MS4A6E) genes.<sup>11</sup>

Oxidative stress and free radicals are effective factors for behavior and memory impairments in age-related neurodegenerative disease.<sup>12</sup> Genetic factors,<sup>13</sup> neuroinflammation,<sup>14</sup> type 2 diabetes, environmental factors, stroke, and diet<sup>15,16</sup> have also been proven to be involved in AD onset and/or progression, although aging is still the main risk factor for AD.<sup>17</sup>

Considering the shortcomings of current treatments for late-onset AD and regarding the positive impact of plant species in AD treatment (i.e. *Melissa officinalis*, *Nigella sativa*, *Boswellia* spp. and *Cinnamon* spp.) natural products are considered as high priorities for treatment of neurodegenerative diseases.<sup>18</sup> Saffron is one of the spices extracted from stigmas of the Persian herb *Crocus sativus* L. and is usually used in cooking<sup>19</sup> and contains four main constituents; safranal, crocin, crocetin, and picrocrocin.

Traditionally, saffron is used for gingival sedation, catarrhal healing, expectoration, improving appetite and digestion, nerve sedation and anticonvulsant, improving sweating, and as antispasmodic.<sup>20,21</sup> During the past two decades, various clinical and experimental studies have revealed that saffron and its bioactive constituents have therapeutic functions as anticonvulsant, anti-hypertensive, anti-spasmodic,<sup>22</sup> cardioprotective, anti-atherosclerotic,<sup>23</sup> anticancer,<sup>24</sup> antidiabetic,<sup>25</sup> antioxidant, antiparasitic,<sup>26</sup> anti-inflammatory, analgesic,<sup>27</sup> and immunomodulators.<sup>28</sup> They might be involved in modulation of smooth muscles,<sup>29</sup> gastrointestinal,<sup>30</sup> respiratory,<sup>31</sup> and reproductive<sup>32</sup> systems. Up to now, the majority of pharmacological experiments on saffron and/or its components have focused on their probable significant effects on CNS. The antidepressant,<sup>33</sup> anticonvulsant,<sup>34</sup> and anti-anxiety properties of saffron were reported, while it may improve memory impairments, tremor,<sup>35</sup> opioid withdrawal syndrome,<sup>36</sup> and was shown to have a broad spectrum of protective effects on the CNS. The stigmas, corms and phytochemicals of *Crocus sativus* could improve neuronal impairments,<sup>37,38</sup> such as Parkinson's disease, by reduction of dopamine in the substantia nigra,<sup>39</sup> suppression of neurotoxicity by diminishing oxidative damage,<sup>40,41</sup> repression of neuroinflammation due to increased intraocular pressure in order to avert retinal ganglion cell death in patients with glaucoma,<sup>42</sup> and improvement of neurodegenerative retinal diseases<sup>43</sup> and visual function in age-related macular degeneration patients.<sup>44</sup> *In vivo*, administration of saffron improved memory impairment induced by ethanol, aluminum (Al), morphine, ketamine, and arsenic.<sup>41,45-48</sup> Extracts of saffron stigma have been presented to have antioxidant and anti-amyloidogenic functions and also inhibited A $\beta$  aggregation and deposition.<sup>21,49</sup>

### Active Constituents of Saffron

Phytochemical analysis showed that saffron contains nearly

150 volatile and some nonvolatile compounds, of which only a few have already been identified. Apocarotenoid glycosides (i.e. crocin); picrocrocin; volatile oil (i.e. safranal); carotenoids; lycopene; alpha-, beta-, and gamma-carotene; fatty oil and starch are the main constituents of this plant.<sup>50</sup> Crocin, and crocetin belong to carotenoids, while picrocrocin and safranal are monoterpene aldehydes. Crocin is a glucosyl ester of crocetin and the compound responsible for the red color of *Crocus sativus*. However, picrocrocin, a glycoside of safranal, provides the unpleasant taste of *Crocus sativus*. Safranal is the main component of saffron and is associated with its aroma.<sup>48</sup> Crocin and safranal isomers have bioactive properties for better absorption in the intestinal lumen.<sup>51-53</sup> It was shown that saffron hydrolyzes to trans-crocetin by intestinal enzymes immediately after the entrance to the lumen and absorbed through the intestinal wall by passive diffusion.<sup>54,55</sup> Crocin is not absorbed orally, after a single dose or repeated doses, but its oral administration produces a higher level of crocetin in comparison with intravenous administration.<sup>56</sup> However, crocin is highly detected in the intestinal tract following oral administration. Crocin can hydrolyze to crocetin when used orally, then the absorbed crocetin is partially metabolized to mono- and di-glucuronide conjugates.<sup>55,57</sup> Crocetin's affinity for binding to albumin is low, which facilitates its transmission to different tissues and helps to cross the blood brain barrier via transcellular diffusion more easily.<sup>54,57,58</sup> According to a recent study in 2019, fast intestinal absorption of saffron extracts leads to a higher serum level of crocetin compared with intravenous administration.<sup>59</sup> The active components of saffron have been shown to possess antidepressant and antitumor effects, while they are able to neutralize free radicals and reduce inflammation.<sup>20,21</sup> Taken together, saffron might be a candidate for research on neurodegenerative diseases. Therefore, this review provides an overview of recently published clinical, preclinical, and experimental studies on therapeutic approaches using saffron and its derivatives for different aspects of AD.

### Amyloid- $\beta$ , a Key Molecule in AD

There are three distinct types of amyloid beta including very short oligomers, A $\beta$  derived diffusible ligands, and protofibrils. Amyloid precursor protein (APP) is produced in the brain and is a major source of neurotoxic A $\beta$ .<sup>60</sup> In detail,  $\beta$ -site APP cleaving enzyme 1 (BACE1), the main  $\beta$ -secretase in the brain, facilitates APP conversion to C<sub>99</sub>.<sup>61,62</sup> Later, A $\beta$  is generated from C<sub>99</sub> by activity of  $\gamma$ -secretase. The  $\gamma$ -secretase function is regulated by presenilin 1 and 2 (PSEN1, 2), and any mutation in these proteins leads to excessive production of A $\beta$ , initiating the early onset of AD.<sup>63</sup> In normal physiological states, there is an equilibrium between the production and clearance of A $\beta$  in the brain,<sup>64</sup> and any disturbance in A $\beta$  elimination or its overproduction will result in AD.<sup>62</sup> Interestingly,

low amounts of A $\beta$  propitiously contribute to neural development<sup>65</sup> and can restrain lipoprotein oxidation in cerebrospinal fluid (CSF).<sup>66</sup> In addition, at low concentrations, A $\beta$  was shown to have neuronal protective effects,<sup>67</sup> whereas high levels of A $\beta$  lead to neuronal dysfunction by disrupting synaptic function and inducing neurotoxicity through free radical formation. Free radicals are accumulated in cerebral vessels, initiating a condition called cerebral amyloid angiopathy (CAA). CAA is a situation in which the amyloid proteins are placed through cerebral blood vessel walls,<sup>68</sup> which happens abundantly in AD.<sup>64,69-71</sup> Besides, accumulation of A $\beta$  disrupts Ca homeostasis in cells and induces excitotoxicity.<sup>72</sup>

Deposition of A $\beta$  also triggers an inflammatory condition through the nuclear factor kappa-light-chain-enhancer of activated B cell (NF- $\kappa$ B) signaling pathway, and activation of microglial cells disrupts central nervous system (CNS) homeostasis in the chronic state.<sup>73</sup> The soluble form of A $\beta$  is attributed to production of imperative proteins related to memory function (i.e. dendritic spines<sup>74</sup>). Soluble A $\beta$  was also shown to have a significant role in AD induction and progression, and its levels rise in the brain,<sup>75</sup> blood, and the CSF<sup>76</sup> of AD patients.<sup>77</sup> Therefore, accumulation or formation of A $\beta$  plaque is an assessment factor for AD diagnosis<sup>67,78</sup> however, there is no known relation between severity of disease and the insoluble form of A $\beta$  or the plaque numbers.<sup>78</sup>

### Tau and Neurofibrillary Tangles in AD

Tau is a protein involved in assembling of tubulin into microtubules<sup>79,80</sup> able to interact with cytoskeletal proteins actin and spectrin. In physiological conditions, neurons are responsible for tau production; nonetheless, in certain pathologic situations, it is also generated by glial cells. Typically, tau proteins are expressed in the CNS; however, the footprints of their mRNAs were also detected in other tissues.<sup>81</sup> It seems that tauopathy leads to neural death and NFT formation, which was also correlated with neuronal disturbance and severity of AD.<sup>82,83</sup> Hyperphosphorylated tau is the main reason behind its neurotoxic properties and also participates in NFT production as a core component.<sup>84,85</sup>

In addition, accumulation of tau is correlated with various degenerative disorders such as AD, progressive supranuclear palsy, argyrophilic grain disease, Pick's disease, Parkinson-dementia complex of Guam, and corticobasal degeneration.<sup>86</sup> Thus far, NFTs and A $\beta$  are the most important components of AD pathology. Indeed, AD-type NFTs are mostly observed in the brain of old individuals even when there are no A $\beta$  plaques. People with NFTs in the brain share comparable symptoms like AD. Regarding resemblance of Primary age related tauopathy (PART) and AD symptoms, by some definitions, PART is considered as a pre-AD factor or a subtype of AD.<sup>87</sup> Despite common features of PART and AD, it has been demonstrated that

PART possesses limited effects on memory and cognition compared to AD.<sup>88</sup>

### Apolipoprotein E and AD

*APOE* is a gene encoding ApoE with 299 amino acids, mainly existing in astrocytes. ApoE regulates lipid homeostasis by modulating lipid transport between different cells and by the action of ApoE receptors in the brain.<sup>9</sup> Liver and macrophages produce ApoE in peripheral tissue and it plays an important role in cholesterol metabolism. ApoE-4 is reported as a risk factor for various disorders such as atherosclerosis, coronary artery disease, peripheral artery disease, type 2 diabetes, and stroke; such diseases are also correlated with AD onset.<sup>89-92</sup>

There are three major alleles of the *APOE* gene including  $\epsilon$ 2,  $\epsilon$ 3, and  $\epsilon$ 4<sup>10</sup> displaying contradictory effects on AD likely due to the difference in amino acid residues 112 and 158.<sup>91</sup> People carrying  $\epsilon$ 4 are more prone to AD than those carrying  $\epsilon$ 3, by far, especially the  $\epsilon$ 4 homozygotes. In contrast,  $\epsilon$ 2 was shown to reduce AD risk.<sup>93-95</sup> According to various genome-wide studies,  $\epsilon$ 4 is the key genetic risk factor for AD.<sup>96,97</sup> Conversely, some studies refute this statement because some people carrying the  $\epsilon$ 4 allele never experience AD. However, they are susceptible to AD twenty times more than others.<sup>98</sup> It was shown that females and ApoE-4 positive individuals can weakly regulate the interaction between microglia cells and amyloid plaques, leading to greater risk of AD.<sup>98</sup> In addition, it was reported that aging and  $\epsilon$ 4 allele synergistically increase the risk of AD.<sup>8,99</sup>

The findings of human and animal studies demonstrated that ApoEs mediate APP and regulate A $\beta$  aggregation and clearance ( $\epsilon$ 4 >  $\epsilon$ 3 >  $\epsilon$ 2) via triggering a non-canonical mitogen-activated protein kinase (MAPK) signaling pathway.<sup>8,100-102</sup> It was stated that absence of ApoE leads to elimination of fibrillar A $\beta$  deposition in the *APOE* gene knockout mouse model.<sup>103</sup> ApoE-4 carriers have more senile plaques and experience CAA more frequently than non-carriers,<sup>104-106</sup> enhancing the risk of AD.<sup>107</sup> It was indicated that the presence of the  $\epsilon$ 4 allele exacerbated the consequences of sedentary lifestyle and aerobic exercise on cognition in individuals who carry  $\epsilon$ 4 in comparison with those not carrying this allele.<sup>108-110</sup> Smoking tobacco,<sup>111-113</sup> mild to moderate alcohol consumption,<sup>114</sup> and diets rich in high saturated fats<sup>113</sup> have also been shown to be responsible for higher risk of AD in  $\epsilon$ 4 carriers.

### Anti-oxidant Effect of Saffron

The antioxidant properties of *Crocus sativus* and its constituents were associated with their activities against the oxidative enzymes; glutathione (GSH), glutathione peroxidase (GPx), superoxide dismutase (SOD),<sup>23</sup> catalase (CAT),<sup>115</sup> glutathione reductase (GR), and glutathione-S-transferase (GST).<sup>116</sup> Thus, saffron and its bioactive compounds can modulate oxidative stress in

cellular organelles and molecules, providing an effective mechanism against neurodegenerative disorders such as AD. It was demonstrated that stressed animals have higher amount of malondialdehyde (MDA), as well as higher activities of GR, GPx, and SOD enzymes in the brain, liver and kidneys, with lower total antioxidant capacity, compared with non-stressed animals.<sup>117</sup> In stressed groups, the corticosterone level was raised, confirming the point that glucocorticoids are involved in chronic-stress-induced oxidative damages, neuronal damage, and impairment of antioxidant defense.<sup>118,119</sup> Chronically elevated glucocorticoids caused neurogenesis blockade, hippocampal volume loss, and atrophy of dendrites in hippocampal CA3 pyramidal neurons.<sup>120,121</sup> Clinically, these changes lead to stress-mediated impairments in spatial learning and memory. Treatment with *Crocus sativus* extract and crocins improved such damages in the stressed group compared with the control group through enhancement of cellular antioxidant and detoxifying pathways.<sup>122</sup>

It was shown that the antioxidant components of saffron, such as crocins, crocetin, safranal, and flavonoids have synergic anti-oxidative effects, as the saffron extract is more efficient than each component alone.<sup>123</sup> Therefore, saffron and its bioactive compounds suppress oxidative and neuronal damages, and can thus alleviate cognitive deficits. Several studies demonstrated that streptozotocin (STZ) induces brain glucose deprivation and oxidative stress in animal models. Reduced cerebral glucose uptake and energy metabolism results in severe and progressive memory loss and poor learning ability due to deficiency in hippocampal choline acetyltransferase content.<sup>12,124</sup> Glucose hypo-metabolism and impaired insulin signaling were implicated in early onset and persistent complications in AD. The behavioral alterations of STZ-lesioned rats were attributed to increased MDA, as well as reduced GSH, total thiol, and GPx activity in the brain.<sup>125</sup>

Crocins were shown to improve cognitive performance, restore GPx activity, reduce lipid peroxidation and MDA pool, and replenish total thiol content in STZ-injected mice.<sup>125</sup> *Striatum* was chosen for injection due to the fact that it is particularly susceptible to oxidative stress damage due to increasing endogenous levels of antioxidants. Cerebral hypoperfusion leads to excessive reactive oxygen species (ROS) generation, which overwhelms the brain's antioxidant machinery, especially in the cortex and hippocampus.<sup>126,127</sup> *Crocus sativus* extract and crocins improved chronic cerebral hypoperfusion-induced cognitive impairments in mice by means of their anti-oxidative properties. In mice with cerebral ischemia-reperfusion injury, safranal treatment significantly restored the hippocampal antioxidant capacity and total-SH content.<sup>128</sup> Moreover, safranal elevated MDA levels in a dose-dependent style in the rat hippocampus in one animal study which was performed on male NMRI rats

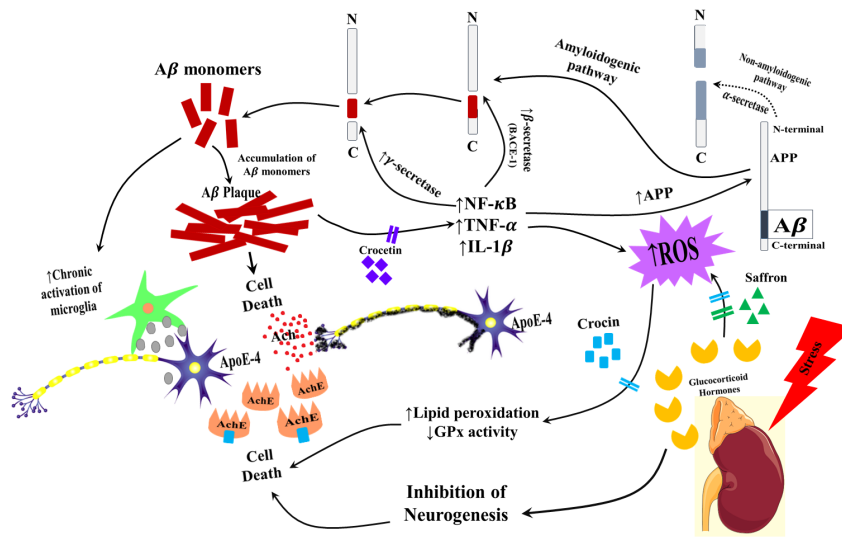
and transient global cerebral ischemia model was induced using the four-vessel-occlusion method for 20 min.<sup>23</sup> *In vitro* experiments on neuronally differentiated PC12 cells demonstrated that stress stimuli (i.e. serum/glucose deprivation, hypoxia), triggers cellular oxidative stress events like decline in intercellular levels of GSH and SOD activity.<sup>23,128</sup> Crocin treatment in PC12 cells attenuated lipid peroxidation and preserved neuron morphology. These effects were correlated to restoration of the activity and expression of SOD, GR,  $\gamma$ -glutamyl-cysteinyl synthase ( $\gamma$ -GCS), and the GSH pool. As mentioned, acrolein activated MAPK/ERK signaling pathway in rat cerebral cortex, as verified by phosphorylation of upstream kinases ERK1/2, c-JNK and p-38, resulting in reduced GSH and an enhancement of MDA content, A $\beta$  deposition, and tau phosphorylation. Co-administration of crocin modulated MAPK signaling pathways, limited MDA pool, reduced A $\beta$  level and tau phosphorylation, and therefore, prevented neuron apoptosis (Figure 1).<sup>129,130</sup>

### Inhibition of AChE Activity and Saffron

It was proven that there is a significant correlation between cholinergic deficiency and cognitive impairments in AD pathogenesis, depending on ACh level in the brain.<sup>131</sup> Cholinergic pathways encompass the medial forebrain cholinergic nuclei and distribute to the hippocampus, amygdala, and neocortex. AChE hydrolyzes ACh to choline and the acetyl group. AChE inhibitors (AChEIs) prevent this breakdown in the brain.<sup>132</sup> However, increased ACh precursors such as choline and lecithin are not useful, but AChEIs have been found to be significantly effective in improving cognitive impairments. Tacrine, donepezil, and rivastigmine are approved AChEI drugs for AD treatment,<sup>133-135</sup> while there are many natural products that can act similarly.<sup>135</sup> Crocins have been shown to inhibit AChE by enhancing ACh levels in synapses and ameliorating cognitive symptoms.<sup>128,136</sup> In a 22-week, double-blind controlled trial, participants with mild to moderate AD randomly consumed either a 30 mg/d capsule of saffron or 10 mg/day of donepezil. Data showed the AChE ratio was comparable for both groups, demonstrating that saffron displayed the same therapeutic effect on cognitive function as donepezil. Besides, patients consuming saffron experienced less vomiting, slightly more dry mouth, and hypomania (Figure 1).<sup>5</sup>

### Inhibition of A $\beta$ Aggregation by Saffron

It was reported that trans-crocetin decreased A $\beta$ 42 aggregation *in vitro* and increased the level of a key A $\beta$ 42 degrading enzyme: the A $\beta$ 42-degrading lysosomal protease cathepsin B (CatB). These data indicate CatB involvement in the degradation pathway of A $\beta$ 42 in AD. Additionally, the compound modulated the intracellular level of CatB, suggesting a potential mechanism by which the degradation ability of A $\beta$ 42 could be retrieved. Studies



**Figure 1.** Inhibitory Effect of Saffron and/or its Derivatives in AD. Amyloid precursor protein (APP) is the main source of amyloid  $\beta$  plaques in the brain. Crocetin lowered the APP level by decreasing the inflammatory cytokines (IL1 $\beta$ , IL18, INF $\gamma$ , and TNF $\alpha$ ). Also, safranal and crocin inhibited the ROS formation resulted from increased stress. Crocin has been shown to inhibit acetylcholine esterase (AChE) and enhances the acetylcholine (ACh) in the neuronal synapses which ameliorates the cognitive symptoms.

also revealed that trans-crocetin has a positive effect on A $\beta$ 42 clearance and verified its neuroprotective effects on A $\beta$ 42-induced toxicity in hippocampal-derived cells, resulting in reduced cellular apoptosis.<sup>7,137</sup> However, it has been shown that crocetin affects multiple signaling pathways involved in neurodegenerative diseases such as extracellular signal-regulated kinase 1/2 (ERK-1/2) and caspases.<sup>129,137</sup> In the rat cerebral cortex, crocin alleviated tau phosphorylation by suppression of ERK and c-Jun N-terminal kinases (JNK) in acrolein induced oxidative stress and amyloid toxicity, showing that modulation of MAPK expression may be a mechanism underlying the crocin neuroprotective characteristic.<sup>138</sup> Supporting this notion, ERK was shown to mediate A $\beta$ -induced tau phosphorylation.<sup>139</sup> In organotypic hippocampal slice cultures, both crocin and crocetin attenuated LPS-induced hippocampal cell death by decreasing nitric oxide (NO) release from activated microglia, highlighting their neuroprotection abilities. It was also demonstrated that crocin inhibited the A $\beta$ 42 formation and aggregation *in vitro*.<sup>140,141</sup> In the *in vitro* neuronal membrane bioreactor model, concomitant administration of crocin and A $\beta$ -peptide repressed apoptosis and ROS production dose dependently.<sup>142</sup> In the *in vivo* model of AD, the compound inhibited A $\beta$ -induced apoptosis through modulating the Bax/Bcl-2 ratio and cleaved caspase-3.<sup>143</sup> In the same manner, crocin prevented neuronal cell death caused by both internal and external apoptotic stimuli in tumor necrosis factor (TNF)- $\alpha$  treated pheochromocytoma PC12 cells via suppression of Bcl-Xs, LICE, and release of cytochrome c from mitochondria.<sup>144</sup> In another study, pretreatment with safranal reduced A $\beta$ 42 induced cell toxicity and apoptosis via MAPK and phosphoinositide

3-kinases (PI3K) pathways in PC12 cells.<sup>137</sup> In an A $\beta$ -induced rat model, application of safranal (0.025, 0.1, and 0.2 mL/kg) for a week improved cognition deficits, and reduced CA1 neuronal loss and the hippocampal levels of MDA, ROS, protein carbonyl, interleukin 1 $\beta$  (IL-1 $\beta$ ), IL-6, TNF- $\alpha$ , Nf- $\kappa$ B, apoptotic biomarkers and DNA fragmentation, glial fibrillary acidic protein (GFAP), myeloperoxidase (MPO), and AChE activity, while enhancing the SOD activity and mitochondrial membrane potential (Figure 1).<sup>145</sup>

**Inhibition of A $\beta$ -Induced Inflammation by Saffron**

High concentrations of neuroinflammatory cytokines were observed in AD brains. Aggregation of A $\beta$  plaques in the brain leads to enhanced neuroinflammatory cytokine levels such as IL-1 $\beta$ , IL-18, interferon- $\gamma$  (IFN- $\gamma$ ), and TNF- $\alpha$ . This enhancement has been correlated with overproduction of APP in glial cells and upregulation of  $\beta$ - and  $\gamma$ - secretaseenzymes, which split APP and produce A $\beta$ .<sup>146,147</sup> Animal studies pointed out that crocetin treatment lowered inflammation, prevented A $\beta$  toxicity and reduced A $\beta$  accumulation by enhancing tightness of the blood brain barrier (BBB), attenuating the increase of NF- $\kappa$ B p65 subunit and P53 in AD mice hippocampus. As a result, nitric oxide synthase (iNOS) production increased whereas proinflammatory cytokines such as IL-1 $\beta$ , IL-18, IFN- $\gamma$ , and TNF- $\alpha$  diminished (Figure 1).<sup>148</sup>

**Inhibition of tau Phosphorylation by Saffron**

It was shown that corcin inhibited the beta-structure/ random coil ratio of tau protein under fibril state and the aggregation of 1N/4R human tau protein in PC12 cells, which was correlated with its chemical structure. It was

proposed that carbonyl groups of crocin could interact with lysine residues of tau, leading to disruption of fibril formation.<sup>149</sup> As mentioned, in the rat cerebral cortex, crocin suppressed acrolein induced tau phosphorylation through modulation of ERK and JNK pathways.<sup>138</sup> Organophosphorus pesticides are accounted as important risk factors of AD. Mohammadzadeh et al reported that crocin (10, 20 and 40 mg/kg) improved spatial memory deficits in rats through inhibition of postsynaptic density protein 93 (PSD93) gene expression and tau phosphorylation. Besides, crocin significantly alleviated both oxidative and inflammatory parameters such as MDA, TNF- $\alpha$  and IL-6 levels, while increasing GSH in the hippocampus. The compound also reduced the plasma AChE activity and malathion-induced apoptosis in the hippocampus cells.<sup>150</sup>

### Saffron and ApoE Related Approaches

Transcriptions of ApoE and ABCA1 are regulated via the linkage of peroxisome proliferator activated receptor  $\gamma$  (PPAR $\gamma$ ) and liver X receptor (LXRs) to the retinoid X receptor (RXR).<sup>151</sup> It was shown that deletion of ABCA1 led to an increase in A $\beta$  deposition in the murine brain, especially in ApoE-4 carriers, signifying its function in A $\beta$  clearance.<sup>152</sup> ABCA1 regulates the ApoE lipidation by means of cholesterol efflux to ApoE and adjusts the ApoE level. On the other hand, binding of ApoE to A $\beta$  changes the conformation of A $\beta$  and increases its clearance.<sup>153</sup> In the Batarseh study, administration of saffron extract enhanced ABCA1 and PPAR $\gamma$  expression in murine brain, which led to A $\beta$  degradation and deposition by modulating the BBB clearance and upregulation of ApoE-dependent A $\beta$  clearance pathway (Figure 1).<sup>148</sup>

### Clinical Trials on Saffron and AD

The mechanism of action of saffron in AD treatment and clinical trials is still under investigation. As previously mentioned, saffron showed similar efficacy as donepezil on patients with mild to moderate AD after 22 weeks by exploiting clinical assessment methods; the Alzheimer's Disease Assessment Scale-cognitive subscale (ADAS-cog) and Clinical Dementia Rating Scale-Sums of Boxes (CDR-SB).<sup>154</sup> Memantine is another approved drug for AD treatment, known to block the glutamatergic N-Methyl-D-aspartic acid (NMDA) receptors and their mediated excitotoxicity in the brain. Administration of saffron at low dose (30 mg/kg) resulted in the same outcomes as Memantine in moderate to severe AD patients. The Severe Cognitive Impairment Rating Scale (SCIRS) scores for both groups indicated no significant difference in the baseline and the final outcomes of the therapy.<sup>155</sup> Furthermore, the results of 16 weeks of saffron therapy versus placebo in individuals with mild to moderate AD were in line with the aforementioned clinical trials. Indeed, saffron significantly prevented cognitive impairments compared to placebo,

highlighting the hypothesis that saffron is beneficial to people suffering from AD and memory deterioration.<sup>156</sup> Administration of saffron with standardized herbal medicine formula named sailuotong, containing *Panax ginseng* and *Ginkgo biloba*, showed potential effectiveness in improving working memory in comparison with placebo in healthy adults.<sup>157</sup> Concomitantly, application of saffron at a dose of 125 mg/d for a year enhanced cognitive function compared with the control group, suggesting that saffron may be an alternative medicine for AD drugs.<sup>158</sup> In a single blind randomized trial, 17 patients diagnosed with amnesic and multi domain mild cognitive impairment (aMCI<sub>md</sub>) were treated with saffron over a year. Neuropsychological assessment included a battery of psychometric tests assessing mood, activities of daily life, behavior, magnetic resonance imaging (MRI) 3T, and general cognitive function, while some patients were assessed via 256-channel electroencephalogram (HD-EEG). The findings of the study showed that saffron improved the MMSE scores, while amending the MRI, EEG, and event-related potential (ERP) in latency of P300 domain, suggesting that saffron may be a choice for MCI therapy (Table 1).<sup>158</sup>

### In Vivo Interventions of Saffron and AD

Regarding the potential role of oxidative stress in pathogenesis of AD and other neurodegenerative diseases, the efficacy of saffron extract was investigated in BALB/c mice hippocampus cells with neuronal damage induced by D-galactose and sodium nitrite (NaNO<sub>2</sub>). While D-galactose increased free radicals and NaNO<sub>2</sub> caused hypoxia, saffron inhibited the neurotoxicity resulting from their actions. This investigation suggested that in addition to anti-oxidative actions, saffron can also increase cerebral blood flow.<sup>159</sup> Administration of crocin in ddY mice after brain infarction induced by occlusion of a middle cerebral artery led to significant reduction of the infarcted area via passing the BBB. Interestingly, crocin was effective in a dose ten-fold less than  $\alpha$ -tocopherol.<sup>160</sup> Similarly, 8 mg/kg crocetin reversed memory derangement in the vascular dementia model in rats including cortical and hippocampal hypoperfusion through permanent occlusion of common carotids, which has been confirmed in histopathological analysis.<sup>161</sup> In accordance, Hosseinzadeh et al<sup>162</sup> found that crocin (25 mg/kg) and saffron (250 mg/kg) attenuated memory deficits via decreasing oxidative stress in Wistar rats.

Zheng et al reported that following cerebral ischemia in C57BL/6J mice, pre-treatment with crocin and saffron inhibited oxidative stress parameters such as MDA and NO, while it enhanced the GPx, SOD, and iNOS activities. In addition, other oxidative markers, phosphorylation of ERK1/2, and the expression of membrane G protein-coupled receptor kinase 2 (GRK2) was reduced. The structure of cortical microvascular endothelial cells was

preserved by crocin.<sup>163</sup>

In a similar manner, pre-treatment with crocin and saffron in rats modulated the CAT and Na-K ATPase activities as well as aspartate and glutamate levels.<sup>40</sup> Increased lipid peroxidation is known as a marker of oxidative stress and IP exposure of saffron extract in Wistar rats led to lipid peroxidation reduction and amelioration of mitochondrial function in synaptosomal fractions, which were predisposed to the neurotoxin mitochondrial toxin 3-nitropropionic (3-NPA).<sup>126</sup> IP and intrahippocampal administration of crocin significantly improved the indicators of spatial memory. In Wistar rats, application of crocin reduced the Bax/Bcl-2 ratio and apoptosis, while the ratio of autophagy markers Beclin-1 and LC3-II/LC3-I remained unchanged.<sup>143</sup>

Moreover, administration of crocin improved sporadic AD induced by STZ in Wistar rats. According to the result of the Passive Avoidance Test and Maze Task Performance, memory and learning deficits were attenuated in the crocin group.<sup>164-166</sup> From a molecular viewpoint, crocin decreased MDA levels while elevating the total thiol level and the GPx activity in contrast to STZ.<sup>165</sup> Likewise, pretreatment with NCSs (combination of *Nardostachys jatamansi*, crocetin and selenium) in Wistar rats attenuated STZ-elicited oxidative stress by reducing thiobarbituric acid reactive substance level and increasing the glutathione, GPx, GST,

and CAT activities, resulting in better performance in passive avoidance test and Morris water maze. Notably, this study mentioned that a multi-substance approach can be more potent than singular therapy.<sup>167</sup> Pretreatment of Wistar rats with saffron extract or crocin for 21 days before predisposition to chronic stress showed a significant neuroprotective effect on the hippocampus and an escalation in anti-oxidative stress markers,<sup>168,169</sup> as well as the mRNA expressions of CAT and SOD.<sup>170</sup> In another study, pre-treatment with a low dose of saffron prevented learning deficits induced by scopolamine in Wistar rats whereas post-treatment with saffron extract significantly retrieved data storage and recognition memory.<sup>171,172</sup> These data are against with findings by Zhang et al.<sup>173</sup>

It has been reported that saffron extract modified morphine-induced memory deficits in mice,<sup>47</sup> which is in line with the study by Haghhighizad et al which indicated the efficacy of saffron extract on improving morphine-induced spatial learning and memory deficit in rats. Other investigations achieved the same results in ethanol-induced memory deficits in Std-ddY mice.<sup>174</sup>

Moreover, administration of 15-30 mg/kg crocin in Wistar rats reduced ketamine (non-competitive NMDA receptor antagonist) induced memory impairments using the novel object recognition task.<sup>48</sup> Saffron extract attenuated the acetaldehyde-induced inhibition of

**Table 1.** Saffron and its Derivatives; Clinical Interventions in AD (Human Study)

Study Design	Study Assessment	Intervention		Number of Patients		Treatment Duration	Outcomes	Adverse Effects	Ref.
		Case	Control	Case	Control				
Mild to moderate AD	MMSEADAS-cog, CDR-SB	SE (15 mg twice/day), oral	donepezil (5 mg twice/day)	n = 24	n = 23	22 weeks	Effective as donepezil	Dizziness, dry mouth, fatigue, hypomania, nausea (adverse effects were similar in both treatment & control groups, except vomiting)	154
Mild cognitive impairment	MMSE	SE (125 mg/d), oral	-	n = 17	n = 18	12 months	Improvement of cognitive dysfunction	-	158
Healthy adults	Cognitive test scores, oddball task -ERP	Sailuotong ( <i>Panax ginseng</i> , <i>Ginkgo biloba</i> & <i>Crocus sativus</i> ) (120 mg/d)	Placebo	n = 8	n = 8	1 week	Increase of Sailuotong in alphabetic working memory & visual working memory	-	157
Mild to moderate AD	MMSEADAS-cog, CDR-SB	SE (15 mg twice/day)	Placebo	n = 22	n = 20	16 weeks	Improvement of cognitive function	Dizziness, dry mouth, fatigue, hypomania, nausea (adverse effects were similar in both treatment & control groups)	156
Moderate to severe AD	MMSE	SE (30 mg/d)	Memantin (20 mg/d)	n = 30	n = 30	12 months	Effective as Memantin in reducing cognitive decline	Nausea, vomiting, dry mouth, fatigue, dizziness, confusion, agitation, sedation (adverse effects were similar in both treatment & control groups)	155

SE, saffron extract; MMSE, Mini-mental state examination; ADAS-cog, Alzheimer's disease assessment scale-cognitive subscale; CDR-SB, Clinical dementia rating scale-sums of boxes; ERP, event-related potential; MRI, magnetic resonance imaging.

hippocampal long-term potentiation in Wistar rats.<sup>175</sup>

For *in vivo* studies, one of the best proficient models of AD can be imitated by chronic administration of aluminum due to the same neurotoxic pathological changes in the brain.<sup>176-179</sup> Al accumulation in the brain leads to oxidative stress in the hippocampus and the cerebral cortex including lipid peroxidation, and deterioration of endogenous antioxidant enzymes, protein kinases, and Na<sup>+</sup>-K<sup>+</sup> ATPase in the cell membrane.<sup>180-182</sup> It was shown that short-term co-administration of saffron extract with Al alleviated the oxidative stress markers and the monoamine oxidase activity; however, there was no effect on cognitive function and memory capacity in BALB/c mice.<sup>41</sup> Oral administration of 100 mg/kg saffron extract reversed the arsenic neurotoxicity while it promoted cognitive and memory functions. This was accompanied by decreasing glutamate and aspartate levels in cortical and hippocampal areas in Wistar rats.<sup>46</sup>

Various parts of the human body are affected by aging which ultimately results in dementia and progressive brain dysfunction. Oxidative stress in lipids, proteins and nucleic acids<sup>183-185</sup> along with poor performance of the cholinergic system due to reduced AChE activity in different parts of the cerebrum<sup>49</sup> and synaptic plasma membranes<sup>186</sup> is the basis for the main hypothesis for memory impairment in aged humans and rodents. In the study by Papandrou et al, crocetin decreased lipid peroxidation and caspase 3 activity in both adult and aged mice although the AChE activity was reduced in only adult BALB/c mice, emphasizing the greater role of oxidative stress in cognitive dysfunction compared to the cholinergic system (Table 2).<sup>136</sup>

### **In Vitro Interventions of Saffron and AD**

*In vitro* administration of safranal, crocetin, and dimethylcrocetin inhibited and preserved the SOD activity which were exposed to serum/glucose deprivation. Moreover, crocin suppressed the caspase-8 (an initiator caspase) activity and increased the survival time of neuronal cells. It was indicated that the anti-oxidative ability of crocin was more than  $\alpha$ -tocopherol (a form of vitamin E) at the same dosage.<sup>188</sup> Comparison of different saffron carotenoids revealed that 10  $\mu$ M crocin is more potent than tricrocetin and dicrocetin in terms of reducing the GSH and caspase 3 activities in PC12 cells.<sup>160</sup>

Saffron and crocetin showed neuroprotective effects on H<sub>2</sub>O<sub>2</sub> induced toxicity in human neuroblastoma SH-SY5Y cells by diminishing ROS products and caspase 3 activity.<sup>136</sup> Pretreatment of PC12 with 10-50  $\mu$ g crocin in the neurotoxic state, induced by acrylamide, reinforced the neuroprotective effect of this compound. Indeed, crocin suppressed intracellular ROS production and apoptosis in these cells.<sup>189</sup> The same neuroprotective action of crocin was recorded in PC12 cells toxicated by either glucose or high levels of ROS.<sup>190</sup> In crocin

pre-treated neurotoxic PC12 cells, the ratio of Bax/Bcl-2 decreased due to the apoptosis inhibitory effect of this compound.<sup>189,191,192</sup> Crocin downregulated TNF- $\alpha$  receptor activity in PC12 cells (mainly through the suppression of Bcl-2 mRNA expression) and increased caspase 3 activity. Besides, crocin prevented intracellular ROS formation elicited by daunorubicin.<sup>144</sup> In another study, 10  $\mu$ M of crocin significantly restored ethanol induced NMDA receptor dysfunction and improved memory impairment in hippocampal slices of male Wistar rats.<sup>45</sup> Crocin suppressed 1-methyl-4-phenylpyridinium-induced endoplasmic reticulum stress and mitochondrial dysfunction in PC12 cells.<sup>193</sup> It is well established that microglial cells play pivotal roles in CNS homeostasis, but chronic activation of microglial cells predisposes neuronal cells to the inflammatory state by producing inflammatory cytokines including IL6, IL1 $\beta$ , TNF- $\alpha$ , and NF- $\kappa$ B transcriptional activity as well as NO release. It was shown that saffron extract repressed the expressions of these elements in BV2 mouse brain microglial cells.<sup>194</sup> Overall, the neuroprotective features of crocin are mainly attributed to reduction of pro-inflammatory cytokines and neurotoxic factors (Table 3).<sup>187,194</sup>

### **Safety**

#### **Animal Studies**

Considering the worldwide application of saffron, monitoring the probable adverse effects of this plant and its bioactive components seems necessary. Acute oral application of saffron in mice and rats was shown to be safe. Following IP administration in mice, the 50% lethal dose (LD<sub>50</sub>) for saffron was reported as 1.6 g/kg, while for oral intake, LD<sub>50</sub> was 4120  $\pm$  556 mg/kg.<sup>195</sup> Administration of 3 g/kg crocin (IP and PO) for two days did not cause any mortality in mice; therefore, it was deduced that crocin is the safest substance of saffron.<sup>196</sup> Safranal exhibited LD<sub>50</sub> values of 0.75 mL/kg and 3.5 mL/kg for IP and oral administration in male Wistar rats, respectively.<sup>196</sup> In rats, sub-acute IP exposure to saffron ethanolic extract decreased body weight, red blood cell (RBC) count, hemoglobin (Hb), and hematocrit (Hct). Conversely in a dose-dependent manner, white blood cells (WBC), alanine aminotransferase (ALT), aspartate aminotransferase (AST) enzymes, serum urea, uric acid, and creatinine (Cr) levels increased. Pathological findings represented some mild to moderate liver and renal damage.<sup>197</sup>

Evaluation of saffron regarding spermatogenesis index in rats showed oral administration of 200 mg/kg saffron for 28 days reduced tubular differentiation index, spermatogenesis index, and repopulation index.<sup>198</sup> Another *in vivo* study demonstrated that crocin (90 mg/kg) for 21 days increased the low-density lipoprotein (LDL) level, while decreasing alkaline phosphatase (ALP) and albumin levels, without serious injuries in main organs even after



Table 2. Saffron and its derivatives; in vivo interventions in AD (Animal Study)

Animal Type	Disease Model (AD)	Intervention		Number of Animals		Treatment Duration	Outcomes	Adverse Effects	Ref.
		Case	Control	Case	Control				
BALB/c mice	D-galactose (90 & 120 mg/kg), i.p	SE (30 mg/kg/d), i.p	Water, i.p	1. amnesic treatment (n = 10) 2. amnesic prophylaxis (n = 10)	1. amnesic control (n = 10) 2. normal control (n = 10)	15 days	Improvement of learning memory impairment in amnesic induced groups	-	159
Wistar rats	3-NPA (20 mg/kg/d), i.p	SE (1 mg/kg/d), i.p	Saline, i.p	n = 6	n = 6	5 days	SE improves mitochondrial function via reduction in LP	-	126
Wistar rats	STZ, i.c.v	Crocin (15 & 30 mg/kg/d), i.p.	Vehicle	n = 15	n = 15	One day pre-surgery & continued for 3 weeks	↓ Memory deficits at higher dose	↑ Weight loss (crocin > crocin+STZ)	164
BALB/c mice	Aluminum chloride (50 mg/kg/d), oral	SE (60 mg/kg/d), i.p	Water, oral	n = 10	n = 10	5 weeks	No changes in cognitive function: ↓ MAO & AChE activity, ↓ LP & GSH	-	41
BALB/c mice	-	SE (60 mg/kg/d), i.p	Saline, i.p	Aged n = 8 Adult n = 8	Aged n = 8 Adult n = 8	7 days	Improvement of learning & memory impairment, ↓ LP, ↑ total brain antioxidant activity, ↓ caspase-3 activity in both aged and adult groups	-	136
Mice	Morphin (5 mg/kg), s.c	SE (50, 150, 450 mg/kg), i.p	Saline, i.p	n = 8	n = 8	3 days	Improvement of memory impairment	-	47
Wistar rats	Arsenic (100 mg/kg), oral	SE (100 mg/kg), gavage needle	-	n = 6	n = 6	-	Improvement of learning ability, ↓ Glutamate & aspartate levels	-	46
Wistar rats	Amyloid β (100 ng/μL), i.p & i.h	Crocin (150, 300, 600 nm), i.p & i.h	-	-	-	-	Crocin ↑ Spatial memory, ↓ brain death	-	143
Wistar rats	Ketamine (3-25 mg/kg), i.p	Crocin (15, 30 & 50 mg/kg), i.p	Vehicle	n = 8	n = 8	3 days	Revision of memory deficits at 50 mg/kg	-	48
Wistar rats	STZ (3 mg/kg on day 1 & 3), i.c.v	Crocin (100 mg/kg), oral	Vehicle	-	-	21 days	Improvement of cognitive function, ↓ MDA, ↑ total thiol content & GPx activity	-	165
Wistar rats	STZ	Combination of Nardosachys jatamansi extract (200 mg/kg), crocetin (25 mg/kg) & selenium (0.05 mg/kg), oral	Saline, oral	-	-	15 days	↓ Cognitive dysfunction	-	167

Table 2. Continued

Animal Type	Disease Model (AD)	Intervention		Number of Animals		Treatment Duration	Outcomes	Adverse Effects	Ref.
		Case	Control	Case	Control				
Wistar rats	Ethidium bromide (3 µL), i.h	SE (5–10 µg), i.h	Saline	n = 8	n = 8	1 week	Improvement of spatial learning & memory improvement, restoration of antioxidant status to the normal levels in hippocampus	-	212
Rats	Chronic cerebral hypoperfusion	crocetin (8 mg/kg), i.p	Control	-	-	-	Prevention of neuropathological alterations in hippocampus, improvement of spatial learning memory	-	161
Wistar rats	BeCl <sub>2</sub> (86 mg/kg), oral	crocin (200 mg/kg), i.p	-	n = 8	n = 8	7 days	↓ Oxidative stress, ↑ mRNA expression of SOD & catalase	-	170
Wistar rats	Scopolamine (0.2 mg/kg), i.p	crocins (15 and 30 mg/kg), i.p	-	n = 10	n = 10	2 days	Crocins (15 mg/kg) ↓ Memory impairment & ↑ recognition memory	-	171
Wistar rats	scopolamine (0.2 mg/kg), i.p	SE (250 mg/kg), oral	-	-	-	-	↓ Hippocampal LTP	-	175
Std-ddY mice	Ethanol (10 ml/kg), po	Crocin (50 to 200 mg/kg), p.o	-	-	-	2 days	↓ Learning behavior impairments & memory retrieval deficits	-	174
C57BL/6 mice	Carotid occlusion-reperfusion	Crocin (5, 10, 20 mg/kg)	-	n = 10	n = 10	21 days	Neuroprotective effect, ↓ GRK2 translocation from the cytosol to the membrane, ↓ ERK1/2 phosphorylation, ↓ expression of MMP-9 in cortical microvessels	-	163
Std-ddY mice	Scopolamin (0.5 mg/kg), i.p & ethanol (10 mg/kg), oral	SE, oral	-	-	-	Single dose	↓ Memory impairment	-	173

Table 2. Continued

Animal Type	Disease Model (AD)	Intervention Case		Number of Animals		Treatment Duration	Outcomes	Adverse Effects	Ref.
		Case	Control	Case	Control				
Wistar rats	(60 mg/kg), STZ i.p	Crocin (7.5, 15, 30, 60 mg/kg), i.p	Saline, i.p	n = 6	n = 6	30 days	Improvement of learning & memory impairments	-	166
Wistar rats	Scopolamin (0.75 mg/kg), s.c	SE (10,30,60 mg/kg), i.p	Vehicle, i.p/s.c	n = 10	n = 10	-	↓ Memory impairment	-	172
Wistar rats	Chronic stress	SE (30 mg/kg) Crocin (15, 30 mg/kg), s.c	Vehicle s.c	n = 10	n = 10	21 days	Prevention of learning impairments & memory deficits; ↓ oxidative stress	-	169
Wistar rats	Cerebral ischemia	SE (50, 100, 250 mg/kg), i.p Crocin (5, 10, 25 mg/kg), i.p	Saline	n = 7	n = 14	-	Improvement of spatial cognitive abilities	-	162
Wistar rats	Chronic stress	SE or crocin (30 mg/kg), i.p	Saline, i.p	n = 6	n = 6	21 days	Prevention of brain oxidative damage, ↓ LP & MDA, ↑ GPx, SOD, GR	-	168
ddy mice	Middle cerebral artery obstruction	crocin (10 mg/kg), i.v.	-	-	-	1 day	↓ Infarcted area via passing BBB	-	160
Wistar rats	Amyloid β (5 µg/µL),	Safranal (0.025, 0.1, 0.2 ml/kg/day), p.o	-	n = 11	n = 11	1 week	↓ CA1 neuronal loss, the hippocampal MDA, ROS, protein carbonyl, interleukin 1β (IL-1β), IL-6, TNF-α, NF-κB, apoptotic biomarkers & DNA fragmentation, glial fibrillary acidic protein (GFAP), myeloperoxidase (MPO), AChE activity, ↑ SOD & mitochondrial membrane potential	-	145
Wistar rats	Malathion (100 mg/kg/dl), i.p	Crocin (10, 20, 40 mg/kg), i.v	Saline, i.p	n = 6	n = 6	14 days	↓ PSD93, tau phosphorylation, MDA, TNF-α, IL-6, plasma AChE activity & malathion-induced apoptosis in hippocampus cells, ↑ GSH	-	150

LPO, Lipid peroxidation; GSH, Glutathione; AChE, Acetylcholine esterase; GPx, Glutathione peroxidase; SOD, Superoxide dismutase; IL, Interleukin; MDA, Malondialdehyde; PSD93, Postsynaptic density protein 93; GFAP, Glial fibrillary acidic protein; TNF-α, Tumor necrosis factor; NF-κB, Nuclear factor kappa-light-chain-enhancer of activated B cell (NF-κB); BCR, Beryllium chloride; LTP, Long-term potentiation; GR, Glutathione reductase; BBB, Blood brain barrier; GRK2, G protein-coupled receptor kinase 2; ERK1/2, Extracellular signal-regulated kinase1/2; i.p. Intraperitoneal; i.c.v. Intracerebroventricular; s.c. Subcutaneous; i.h. Intrahippocampal; p.o. per os  
↓: decrease, ↑: increase

**Table 3.** Saffron and its Derivatives; *In Vitro* Interventions in AD

Ref.	Cell Type	Study Model of AD	Intervention		Number of Cells		Treatment Duration	Results
			Case	Control	Case	Control		
187	-	Kinetic analysis of AChE	Crocin, crocetin, dimethylcrocetin, safranal	Control & galanthamine	-	-	-	AChE inhibition activity; Safranal > crocetin > dimethylcrocetin
49	-	Ferric-reducing antioxidant power & Trolox-equivalent antioxidant capacity	SE (50:50 water & methanol)	-	-	-	-	↑ Cognitive function via antioxidant & antiamyloidogenic activity, ↑ cognitive function
45	Hippocampal slices of male Wistar rats	Ethanol	Crocin (10 µM)	-	-	-	-	Revision of the inhibitory effect of ethanol on NMDA receptor-mediated responses
193	PC12	1-methyl-4-phenylpyridinium (MPP+)	Crocin	-	-	-	-	↓ MPP+-induced ER stress & cell injury
189	PC12	acrylamide (5 mM)	Crocin (10–50 mM)	-	-	-	-	↓ Apoptosis, ↓ Intracellular ROS formation
194	BV2 mouse microglial cells; hippocampal slice cultures organotypic of rats	LPS	Crocin, crocetin	-	-	-	-	↓ Cell death, ↑ anti-oxidative & anti-inflammatory effects
160	PC12	Serum-free & hypoxic induced cell-death	Crocin (10 µM)	-	-	-	-	↓ infarcted areas
188	PC12	Serum/glucose deprivation	Crocin (10 mM)	-	-	-	-	↓ LP content, ↑ SOD activity, protected neuron's morphology
136	Neuroblastoma SH-SY55 human cells	H2O2	SE & crocetin (1–125 µmol)	-	-	-	-	↓ Cell death, ↓ caspase 3 activity & ROS formation
144	PC12	TNF-α & daunorubicin	Crocin (1–10 µM)	-	-	-	-	↓ Cell death, ↓ both internal & external apoptotic stimuli
190	PC12	High glucose (4.5, 13.5, & 27 mg/mL)	SE (5 & 25 µg/mL), crocetin (10, 50 µM)	-	-	-	4 days	↓ Cell death, ↓ ROS production & glucose toxicity
141	-	Na <sub>2</sub> HPO <sub>4</sub> NaCl (100 mM)	Crocin (15.4 µM)	-	-	-	-	↓ Amyloid fibril content of Aβ; inhibits Aβ aggregation
140	-	Na <sub>2</sub> HPO <sub>4</sub> NaCl (100 mM)	Crocin (15 µg/mL)	-	-	-	-	↓ Aβ40 average fibril length, ↓ formation of Aβ fibril formation
213	PC12	-	Crocin (10 µg/mL)	-	-	-	-	↓ Tau protein fibrillation

SE, Saffron extract; MPP+, 1-methyl-4-phenylpyridinium; LPS, Lipopolysaccharide; LP, Lipid peroxidation; SOD, Superoxide dismutase; ROS, Reactive oxygen species  
↓, decrease; ↑, increase

exposure to 180 mg/kg of crocin.<sup>196</sup> This was in line with results of a study by Taheri et al since IP administration of crocin at concentrations of 50, 100 and 200 mg/kg once a week for four weeks caused no elevation in Cr, ALT, AST, ALP, uric acid and urea levels in rats. Pathological examination revealed no significant hepatic toxicity.<sup>199</sup>

Three weeks of safranal oral administration (0.1, 0.25, 0.5 mL/kg) in rats, led to reduction of triglyceride, cholesterol, ALP, RBC count, platelet count, Hb, and Hct, while the level of blood urea nitrogen (BUN) increased. However, no pathological lesion in organs (liver, spleen and heart) or toxicity effect on the cellular and humoral immune system were detected.<sup>200</sup> Oral administration of 4000 and 5000 mg/kg saffron in BALB/c mice for 5 weeks demonstrated that sub-chronic exposure to saffron decreased the RBC and WBC counts and increased BUN and Cr, indicating renal dysfunction.<sup>201</sup> Worth mentioning, usually in animal studies, saffron is used at high doses although it exhibited protective effects in lower doses.<sup>202</sup>

### Human Studies

Like other plant extracts, several side effects were reported for saffron such as nausea, vomiting, anxiety, headache, dizziness, epistaxis, bloody diarrhea, and numbness. It was assumed that at doses of 12-20 g, saffron can be fatal.<sup>203</sup>

In a clinical study on healthy volunteers, standing systolic blood pressure and mean arterial pressure were reduced by receiving saffron tablets (400 mg); however, there was no change at a dose of 200 mg.<sup>204</sup> In another study, hematologic factors and the coagulation system were not disturbed by saffron tablets (200 and 400 mg).<sup>205</sup> Safety of crocin was investigated in a double-blind, placebo-controlled trial in which healthy volunteers received crocin tablets (20 mg) for a month. Crocin tablets decreased amylase, partial thromboplastin time, and the WBC count, demonstrating that crocin was relatively safe.<sup>206</sup> Pregnant women with fetuses at gestational ages between the first and twentieth weeks were susceptible to abortion if they received saffron at high doses.<sup>207</sup> In addition, uterine contractions induced by saffron have been suggested as a mechanism for abortion.<sup>204,208,209</sup> At the beginning of the active phase of labor, administration of saffron capsules (250 mg) reduced mean anxiety score and mean fatigue score,<sup>210</sup> while saffron capsules in the active phase of labor reduced pain. The infant and mother did not show any toxicity in the saffron group compared with controls.<sup>211</sup>

Based on the findings of animal studies (LD<sub>50</sub> values), crocin might be the safest component of saffron, and no significant damage has been mentioned for this compound at pharmacological dosage. At high concentrations, saffron and its constituents showed some developmental toxicity on animal infants. Exposure to high levels of saffron was shown to increase miscarriage rates in pregnant women, suggesting avoidance of high doses during pregnancy.<sup>202</sup>

### Conclusion and Future Perspectives

Statistics confirm that AD remains a global growing health concern. A wide range of natural and synthetic molecules have been studied for their ability to either prevent or counteract AD initiation, progression, and complications. The findings of this study indicate that saffron and/or its components target various regulatory molecules involved in AD. Regarding its pleotropic effects on the nervous system, including anti-amyloid, anti-AChE, anti-inflammatory, and anti-oxidant features, along with its inhibitory effect on tau hyper-phosphorylation, and upregulation of ApoE activity, it seems that saffron could find its niche in AD therapy with substantial potential as a therapeutic nutraceutical with the advantage of low toxicity and easy accessibility. Further studies, particularly clinical trials, are now required to determine whether saffron and its bioactive phytochemicals may be suitable for AD or other neurodegenerative disorders. Other clinical trials are warranted to examine the safety and efficacy of various doses of the plant and improved formulations with better pharmacokinetics and bioavailability are needed. Several reports have raised questions about the safety and efficacy of saffron or its derivatives, especially at high doses, whereas some studies have shown no adverse effects. It is suggested that the mode of administration and the duration of saffron therapy are also critical factors that can significantly affect the efficacy of AD treatment. Since saffron is a part of daily diets in many Asian countries and seems non-toxic, it is obligatory to investigate whether dietary supplementation with saffron may be a beneficial preventive or slowing nutritional strategy for neurological disorders.<sup>212</sup>

### Authors' Contribution

NZ, BP, NMR and NAL: collection and/or assembly of data and interpretation, manuscript writing; SM, VJ, AHA and SA: provision of study material, conception and design, and final approval of manuscript. All the authors have read and approved the manuscript.

### Conflict of Interest Disclosures

None.

### Ethical Statement

Not applicable.

### References

- Bhardwaj D, Mitra C, Narasimhulu CA, Riad A, Doomra M, Parthasarathy S. Alzheimer's Disease—Current Status and Future Directions. *J Med Food*. 2017;20(12):1141-51. doi: 10.1089/jmf.2017.0093.
- Brookmeyer R, Johnson E, Ziegler-Graham K, Arrighi HM. Forecasting the global burden of Alzheimer's disease. *Alzheimers Dement*. 2007;3(3):186-91. doi: 10.1016/j.jalz.2007.04.381.
- Tsuno N. Donepezil in the treatment of patients with Alzheimer's disease. *Expert Rev Neurother*. 2009;9(5):591-8. doi: 10.1586/ern.09.23.
- Mohamed T, Shakeri A, Rao PP. Amyloid cascade in Alzheimer's disease: recent advances in medicinal chemistry. *Eur J Med Chem*. 2016;113:258-72. doi: 10.1016/j.ejmech.2016.02.049.

5. Adalier N, Parker H. Vitamin E, turmeric and saffron in treatment of Alzheimer's disease. *Antioxidants (Basel)*. 2016;5(4):40. doi: 10.3390/antiox5040040.
6. Swerdlow RH, Burns JM, Khan SM. The Alzheimer's disease mitochondrial cascade hypothesis: progress and perspectives. *Biochim Biophys Acta*. 2014;1842(8):1219-31. doi: 10.1016/j.bbadis.2013.09.010.
7. Barage SH, Sonawane KD. Amyloid cascade hypothesis: Pathogenesis and therapeutic strategies in Alzheimer's disease. *Neuropeptides*. 2015;52:1-18. doi: 10.1016/j.npep.2015.06.008.
8. Reiman EM, Chen K, Liu X, Bandy D, Yu M, Lee W, et al. Fibrillar amyloid- $\beta$  burden in cognitively normal people at 3 levels of genetic risk for Alzheimer's disease. *Proc Natl Acad Sci U S A*. 2009;106(16):6820-5. doi: 10.1073/pnas.0900345106.
9. Mahley RW, Weisgraber KH, Huang Y. Apolipoprotein E4: a causative factor and therapeutic target in neuropathology, including Alzheimer's disease. *Proc Natl Acad Sci U S A*. 2006;103(15):5644-51. doi: 10.1073/pnas.0600549103.
10. Nussbaum RL, McInnes RR, Willard HF. *Thompson & Thompson genetics in medicine e-book*. Elsevier Health Sciences; 2015.
11. Bird TD. Alzheimer disease overview. *GeneReviews* [Internet]: University of Washington, Seattle; 2018.
12. Naghizadeh B, Mansouri MT, Ghorbanzadeh B. Protective effects of crocin against streptozotocin-induced oxidative damage in rat striatum. *Acta Med Iran*. 2014;52(2):101-5.
13. Karch CM, Cruchaga C, Goate AM. Alzheimer's disease genetics: from the bench to the clinic. *Neuron*. 2014;83(1):11-26. doi: 10.1016/j.neuron.2014.05.041.
14. Heneka MT, Carson MJ, El Khoury J, Landreth GE, Brosseron F, Feinstein DL, et al. Neuroinflammation in Alzheimer's disease. *Lancet Neurol*. 2015;14(4):388-405. doi: 10.1016/S1474-4422(15)70016-5.
15. Chin-Chan M, Navarro-Yepes J, Quintanilla-Vega B. Environmental pollutants as risk factors for neurodegenerative disorders: Alzheimer and Parkinson diseases. *Front Cell Neurosci*. 2015;9:124. doi: 10.3389/fncel.2015.00124.
16. Arnold SE, Arvanitakis Z, Macauley-Rambach SL, Koenig AM, Wang HY, Ahima RS, et al. Brain insulin resistance in type 2 diabetes and Alzheimer disease: concepts and conundrums. *Nat Rev Neurol*. 2018;14(3):168-181. doi: 10.1038/nrneuro.2017.185.
17. Qiu C, Fratiglioni L. Aging without dementia is achievable: current evidence from epidemiological research. *J Alzheimers Dis*. 2018;62(3):933-942. doi: 10.3233/JAD-171037.
18. Ernst E. Herbal medicines—they are popular, but are they also safe? *Eur J Clin Pharmacol*. 2006;62(1):1-2. doi: 10.1007/s00228-005-0070-2.
19. José Bagur M, Alonso Salinas GL, Jiménez-Monreal AM, Chaouqi S, Llorens S, Martínez-Tomé M, et al. Saffron: An old medicinal plant and a potential novel functional food. *Molecules*. 2017;23(1):30. doi: 10.3390/molecules23010030.
20. Akhondzadeh S, Abbasi SH. Herbal medicine in the treatment of Alzheimer's disease. *Am J Alzheimers Dis Other Dement*. 2006;21(2):113-8. doi: 10.1177/153331750602100211.
21. Schmidt M, Betti G, Hensel A. Saffron in phytotherapy: pharmacology and clinical uses. *Wien Med Wochenschr*. 2007;157(13-14):315-9. doi: 10.1007/s10354-007-0428-4.
22. Fatehi M, Rashidabady T, Fatehi-Hassanabad Z. Effects of *Crocus sativus* petals' extract on rat blood pressure and on responses induced by electrical field stimulation in the rat isolated vas deferens and guinea-pig ileum. *J Ethnopharmacol*. 2003;84(2-3):199-203. doi: 10.1016/s0378-8741(02)00299-4.
23. Sachdeva J, Tanwar V, Golechha M, Siddiqui KM, Nag TC, Ray R, et al. *Crocus sativus* L.(saffron) attenuates isoproterenol-induced myocardial injury via preserving cardiac functions and strengthening antioxidant defense system. *Exp Toxicol Pathol*. 2012;64(6):557-64. doi: 10.1016/j.etp.2010.11.013.
24. Khorasanchi Z, Shafiee M, Kermanshahi F, Khazaei M, Ryzhikov M, Parizadeh MR, et al. *Crocus sativus* a natural food coloring and flavoring has potent anti-tumor properties. *Phytomedicine*. 2018;43:21-27. doi: 10.1016/j.phymed.2018.03.041
25. Yaribeygi H, Zare V, Butler AE, Barreto GE, Sahebkar A. Antidiabetic potential of saffron and its active constituents. *J Cell Physiol*. 2019;234(6):8610-8617. doi: 10.1002/jcp.27843
26. Wali AF, Alchamat HAA, Hariri HK, Hariri BK, Menzes GA, Zehra U, et al. Antioxidant, Antimicrobial, Antidiabetic and Cytotoxic Activity of *Crocus sativus* L. Petals. *Applied Sciences*. 2020;10(4):1519. doi: 10.3390/app10041519
27. Moallem SA, Hariri AT, Mahmoudi M, Hosseinzadeh H. Effect of aqueous extract of *Crocus sativus* L.(saffron) stigma against subacute effect of diazinon on specific biomarkers in rats. *Toxicol Ind Health*. 2014;30(2):141-146. doi: 10.1177/0748233712452609
28. Bani S, Pandey A, Agnihotri VK, Pathania V, Singh B. Selective Th2 upregulation by *Crocus sativus*: a nutraceutical spice. *Evid Based Complement Alternat Med*. 2011;2011. doi: 10.1155/2011/639862
29. Bayrami G, Boskabady M. The potential effect of the extract of *Crocus sativus* and safranal on the total and differential white blood cells of ovalbumin-sensitized guinea pigs. *Res Pharm Sci*. 2012;7(4):249.
30. Halataei BaS, Khosravi M, Arbabian S, Sahraei H, Golmanesh L, Zardooz H, et al. Saffron (*Crocus sativus*) aqueous extract and its constituent crocin reduces stress-induced anorexia in mice. *Phytother Res*. 2011;25(12):1833-8. doi: 10.1002/ptr.3495
31. Byrami G, Boskabady MH, Jalali S, Farkhondeh T. The effect of the extract of *Crocus sativus* on tracheal responsiveness and plasma levels of IL-4, IFN- $\gamma$ , total NO and nitrite in ovalbumin sensitized Guinea-pigs. *J Ethnopharmacol*. 2013;147(2):530-5. doi: 10.1016/j.jep.2013.03.014
32. Mohammadzadeh-Moghadam H, Nazari SM, Shamsa A, Kamalinejad M, Esmaeeli H, Asadpour AA, et al. Effects of a topical saffron (*Crocus sativus* L) gel on erectile dysfunction in diabetics: A randomized, parallel-group, double-blind, placebo-controlled trial. *J Evid Based Complementary Altern Med*. 2015;20(4):283-286. doi: 10.1177/2156587215583756
33. Lopresti AL, Smith SJ, Hood SD, Drummond PD. Efficacy of a standardised saffron extract (affron®) as an add-on to antidepressant medication for the treatment of persistent depressive symptoms in adults: A randomised, double-blind, placebo-controlled study. *J Psychopharmacol*. 2019;33(11):1415-27. doi: 10.1177/0269881119867703
34. Hosseinzadeh H, Talebzadeh F. Anticonvulsant evaluation of safranal and crocin from *Crocus sativus* in mice. *Fitoterapia*. 2005;76(7-8):722-4. doi: 10.1016/j.fitote.2005.07.008
35. Amin B, Malekzadeh M, Heidari MR, Hosseinzadeh H. Effect of *Crocus sativus* extracts and its active constituent safranal on the harmaline-induced tremor in mice. *Iran J Basic Med Sci*. 2015;18(5):449.
36. Hosseinzadeh H, Jahanian Z. Effect of *Crocus sativus* L.(saffron) stigma and its constituents, crocin and safranal, on morphine withdrawal syndrome in mice. *Phytother Res*. 2010;24(5):726-30. doi: 10.1002/ptr.3011
37. Hosseinzadeh H, Khosravan V. Anticonvulsant effect of aqueous and ethanolic extracts of *Crocus sativus* L stigmas in mice. *BMC Pharmacol*. 2002. doi: 10.1186/1471-2210-2-7
38. Sadeghnia HR, Kamkar M, Assadpour E, Boroushaki MT, Ghorbani A. Protective effect of safranal, a constituent of *Crocus sativus*, on quinolinic acid-induced oxidative damage in rat hippocampus. *Iran J Basic Med Sci*. 2013;16(1):73.
39. Ahmad AS, Ansari MA, Ahmad M, Saleem S, Yousuf S, Hoda

- MN, et al. Neuroprotection by crocetin in a hemi-parkinsonian rat model. *Pharmacol Biochem Behav.* 2005;81(4):805-13. doi: 10.1016/j.pbb.2005.06.007
40. Saleem S, Ahmad M, Ahmad AS, Yousuf S, Ansari MA, Khan MB, et al. Effect of Saffron (*Crocus sativus*) on neurobehavioral and neurochemical changes in cerebral ischemia in rats. *J Med Food.* 2006;9(2):246-53. doi: 10.1089/jmf.2006.9.246
  41. Linardaki ZI, Orkoulas MG, Kokkosis AG, Lamari FN, Margarity M. Investigation of the neuroprotective action of saffron (*Crocus sativus* L.) in aluminum-exposed adult mice through behavioral and neurobiochemical assessment. *Food Chem Toxicol.* 2013;52:163-70. doi: 10.1016/j.fct.2012.11.016
  42. Fernández-Albarral JA, Ramírez AI, de Hoz R, López-Villarín N, Salobar-García E, López-Cuenca I, et al. Neuroprotective and Anti-Inflammatory Effects of a Hydrophilic Saffron Extract in a Model of Glaucoma. *Int J Mol Sci.* 2019;20(17):4110. doi: 10.3390/ijms20174110
  43. Fernández-Albarral JA, de Hoz R, Ramírez AI, López-Villarín N, Salobar-García E, López-Cuenca I, et al. Beneficial effects of saffron (*Crocus sativus* L.) in ocular pathologies, particularly neurodegenerative retinal diseases. *Neural Regen Res.* 2020;15(8):1408-16. doi: 10.4103/1673-5374.274325
  44. Broadhead GK, Grigg JR, McCluskey P, Hong T, Schlub TE, Chang AA. Saffron therapy for the treatment of mild/moderate age-related macular degeneration: a randomised clinical trial. *Graefes Arch Clin Exp Ophthalmol.* 2019;257(1):31-40. doi: 10.1007/s00417-018-4163-x
  45. Abe K, Sugiura M, Shoyama Y, Saito H. Crocin antagonizes ethanol inhibition of NMDA receptor-mediated responses in rat hippocampal neurons. *Brain Res.* 1998;787(1):132-8. doi: 10.1016/s0006-8993(97)01505-9
  46. Sreenu G, Banala RR, Reddy KP. Saffron extract's protective effects against arsenic induced excitotoxicity and learning disabilities in male Wistar rats. *Int J Bioassay.* 2015;4:4223-9. doi:10.21746/IJBIO.2015.08.0012
  47. Naghibi SM, Hosseini M, Khani F, Rahimi M, Vafae F, Rakhshandeh H, et al. Effect of aqueous extract of *Crocus sativus* L. on morphine-induced memory impairment. *Adv Pharmacol Sci.* 2012;2012. doi: 10.1155/2012/494367
  48. Georgiadou G, Grivas V, Tarantilis PA, Pitsikas N. Crocins, the active constituents of *Crocus sativus* L., counteracted ketamine-induced behavioural deficits in rats. *Psychopharmacology (Berl).* 2014;231(4):717-26. doi: 10.1007/s00213-013-3293-4
  49. Papandreou MA, Kanakis CD, Polissiou MG, Efthimiopoulos S, Cordopatis P, Margarity M, et al. Inhibitory activity on amyloid- $\beta$  aggregation and antioxidant properties of *Crocus sativus* stigmas extract and its crocin constituents. *J Agric Food Chem.* 2006;54(23):8762-8. doi: 10.1021/jf061932a
  50. Al-Snafi AE. The pharmacology of *Crocus sativus*-A review. *IOSR Journal of Pharmacy.* 2016;6(6):8-38.
  51. Bukhari SI, Manzoor M, Dhar M. A comprehensive review of the pharmacological potential of *Crocus sativus* and its bioactive apocarotenoids. *Biomed Pharmacother.* 2018;98:733-45. doi: 10.1016/j.biopha.2017.12.090
  52. Mykhailenko O, Kovalyov V, Goryacha O, Ivanauskas L, Georgiyants V. Biologically active compounds and pharmacological activities of species of the genus *Crocus*: A review. *Phytochemistry.* 2019;162:56-89. doi: 10.1016/j.phytochem.2019.02.004
  53. Heitmar R, Brown J, Kyrou I. Saffron (*Crocus sativus* L.) in ocular diseases: A narrative review of the existing evidence from clinical studies. *Nutrients.* 2019;11(3):649. doi: 10.3390/nu11030649
  54. Lautenschläger M, Sendker J, Hüwel S, Galla HJ, Brandt S, Düfer M, et al. Intestinal formation of trans-crocetin from saffron extract (*Crocus sativus* L.) and in vitro permeation through intestinal and blood brain barrier. *Phytomedicine.* 2015;22(1):36-44. doi: 10.1016/j.phymed.2014.10.009
  55. Xi L, Qian Z, Du P, Fu J. Pharmacokinetic properties of crocin (crocetin digentiobiose ester) following oral administration in rats. *Phytomedicine.* 2007;14(9):633-636. doi: 10.1016/j.phymed.2006.11.028
  56. Yang N, Sun RB, Chen XL, Zhen L, Ge C, Zhao YQ, et al. In vitro assessment of the glucose-lowering effects of berberubine-9-O- $\beta$ -D-glucuronide, an active metabolite of berberubine. *Acta Pharmacol Sin.* 2017;38(3):351-61. doi: 10.1038/aps.2016.120
  57. Asai A, Nakano T, Takahashi M, Nagao A. Orally administered crocetin and crocins are absorbed into blood plasma as crocetin and its glucuronide conjugates in mice. *J Agric Food Chem.* 2005;53(18):7302-6. doi: 10.1021/jf0509355
  58. Hosseini A, Razavi BM, Hosseinzadeh H. Pharmacokinetic properties of saffron and its active components. *Eur J Drug Metab Pharmacokinet.* 2018;43(4):383-90. doi: 10.1007/s13318-017-0449-3
  59. Christodoulou E, Grafakou ME, Skaltsa E, Kadoglou N, Kostomitsopoulos N, Valsami G. Preparation, chemical characterization and determination of crocetin's pharmacokinetics after oral and intravenous administration of saffron (*Crocus sativus* L.) aqueous extract to C57/BL 6j mice. *J Pharm Pharmacol.* 2019;71(5):753-64. doi: 10.1111/jphp.13055
  60. O'Brien RJ, Wong PC. Amyloid precursor protein processing and Alzheimer's disease. *Annu Rev Neurosci.* 2011;34:185-204. doi: 10.1146/annurev-neuro-061010-113613
  61. Harkany T, Abraham I, Timmerman W, Laskay G, Tóth B, Sasvári M, et al.  $\beta$ -Amyloid neurotoxicity is mediated by a glutamate-triggered excitotoxic cascade in rat nucleus basalis. *Eur J Neurosci.* 2000;12(8):2735-45. doi: 10.1046/j.1460-9568.2000.00164.x
  62. Qiu C, Kivipelto M, von Strauss E. Epidemiology of Alzheimer's disease: occurrence, determinants, and strategies toward intervention. *Dialogues Clin Neurosci.* 2009;11(2):111. doi: 10.31887/DCNS.2009.11.2/cqiu
  63. Kirkitadze MD, Kowalska A. Molecular mechanisms initiating amyloid beta-fibril formation in Alzheimer's disease. *Acta Biochim Pol.* 2005;52(2):417-423.
  64. Shankar GM, Walsh DM. Alzheimer's disease: synaptic dysfunction and A $\beta$ . *Mol Neurodegener.* 2009;4(1):48. doi: 10.1186/1750-1326-4-48
  65. Sadigh-Eteghad S, Talebi M, Farhoudi M, Golzari SE, Sabermarouf B, Mahmoudi J. Beta-amyloid exhibits antagonistic effects on alpha 7 nicotinic acetylcholine receptors in orchestrated manner. *Journal of Medical Hypotheses and Ideas.* 2014;8(2):49-52. doi: 10.1016/j.jmhi.2014.01.001
  66. Kontush A, Berndt C, Weber W, Akopyan V, Arlt S, Schippling S, et al. Amyloid- $\beta$  is an antioxidant for lipoproteins in cerebrospinal fluid and plasma. *Free Radic Biol Med.* 2001;30(1):119-28. doi: 10.1016/s0891-5849(00)00458-5
  67. Nunomura A, Castellani RJ, Lee H-g, Moreira PI, Zhu X, Perry G et al. Neuropathology in Alzheimer's disease: awaking from a hundred-year-old dream. *Sci Aging Knowledge Environ.* 2006;8:e10. doi: 10.1126/sageke.2006.8.pe10
  68. Vinters HV. Cerebral amyloid angiopathy. A critical review. *Stroke.* 1987;18(2):311-24. doi: 10.1161/01.str.18.2.311
  69. Ellis R, Olichney JM, Thal L, Mirra SS, Morris JC, Beekly D et al. Cerebral amyloid angiopathy in the brains of patients with Alzheimer's disease: the CERAD experience, Part XV. *Neurology.* 1996;46(6):1592-6. doi: 10.1212/wnl.46.6.1592
  70. Attems J, Yamaguchi H, Saido TC, Thal DR. Capillary CAA and perivascular A $\beta$ -deposition: two distinct features of Alzheimer's disease pathology. *J Neurol Sci.* 2010;299(1-2):155-62. doi: 10.1016/j.jns.2010.08.030
  71. Kihara T, Shimohama S. Alzheimer's disease and acetylcholine receptors. *Acta Neurobiol Exp (Wars).* 2004;64(1):99-106.
  72. Sadigh-Eteghad S, Sabermarouf B, Majdi A, Talebi M,

- Farhoudi M, Mahmoudi J. Amyloid-beta: a crucial factor in Alzheimer's disease. *Med Princ Pract.* 2015;24(1):1-10. doi: 10.1159/000369101
73. Ridolfi E, Barone C, Scarpini E, Galimberti D. The role of the innate immune system in Alzheimer's disease and frontotemporal lobar degeneration: an eye on microglia. *Clin Dev Immunol.* 2013;2013. doi: 10.1155/2013/939786
  74. Balducci C, Beeg M, Stravalaci M, Bastone A, Sclip A, Biasini E, et al. Synthetic amyloid- $\beta$  oligomers impair long-term memory independently of cellular prion protein *Proc Natl Acad Sci U S A.* 2010;107(5):2295-2300. doi: 10.1073/pnas.0911829107
  75. Barry AE, Klyubin I, Mc Donald JM, Mably AJ, Farrell MA, Scott M, et al. Alzheimer's disease brain-derived amyloid- $\beta$ -mediated inhibition of LTP in vivo is prevented by immunotargeting cellular prion protein. *J Neurosci.* 2011;31(20):7259-63. doi: 10.1523/JNEUROSCI.6500-10.2011
  76. Shen CL, Fitzgerald MC, Murphy RM. Effect of acid predissolution on fibril size and fibril flexibility of synthetic beta-amyloid peptide. *Biophys J.* 1994;67(3):1238-46. doi:10.1016/S0006-3495(94)80593-4
  77. Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science.* 2002;297(5580):353-6. doi: 10.1126/science.1072994.
  78. Kittelberger KA, Piazza F, Tesco G, Reijmers LG. Natural amyloid-beta oligomers acutely impair the formation of a contextual fear memory in mice. *PLoS One.* 2012;7(1):e29940. doi: 10.1371/journal.pone.0029940.
  79. Davidowitz EJ, Chatterjee I, Moe JG. Targeting tau oligomers for therapeutic development for Alzheimer's disease and tauopathies. *Biotechnology.* 2008;4:47-64.
  80. Götz J, Probst A, Spillantini MG, Schäfer T, Jakes R, Bürki K, et al. Somatodendritic localization and hyperphosphorylation of tau protein in transgenic mice expressing the longest human brain tau isoform. *EMBO J.* 1995;14(7):1304-1313.
  81. Buée L, Bussi re T, Bu e-Scherrer V, Delacourte A, Hof PR. Tau protein isoforms, phosphorylation and role in neurodegenerative disorders. *Brain Res Brain Res Rev.* 2000;33(1):95-130. doi: 10.1016/S0165-0173(00)00019-9
  82. Brion J-P. Neurofibrillary tangles and Alzheimer's disease. *Eur Neurol.* 1998;40(3):130-40. doi: 10.1159/000007969
  83. Kimura T, Yamashita S, Fukuda T, Park JM, Murayama M, Mizoroki T et al. Hyperphosphorylated tau in parahippocampal cortex impairs place learning in aged mice expressing wild-type human tau. *EMBO J.* 2007;26(24):5143-52. doi: 10.1038/sj.emboj.7601917
  84. Kimura T, Whitcomb DJ, Jo J, Regan P, Piers T, Heo S, et al. Microtubule-associated protein tau is essential for long-term depression in the hippocampus. *Philos Trans R Soc Lond B Biol Sci.* 2014;369(1633):20130144. doi: 10.1098/rstb.2013.0144
  85. Mohorko N, Bresjanac M. Tau protein and human tauopathies: an overview. *Slovenian Medical Journal.* 2008;77.
  86. Goedert M, Spillantini MG. A century of Alzheimer's disease. *Science.* 2006;314(5800):777-81. doi: 10.1126/science.1132814.
  87. Braak H, Del Tredici K. Where, when, and in what form does sporadic Alzheimer's disease begin? *Curr Opin Neurol.* 2012;25(6):708-14. doi: 10.1097/WCO.0b013e32835a3432
  88. Santa-Maria I, Haggiagi A, Liu X, Wasserscheid J, Nelson PT, Dewar K, et al. The MAPT H1 haplotype is associated with tangle-predominant dementia. *Acta neuropathol.* 2012;124(5):693-704. doi: 10.1007/s00401-012-1017-1
  89. Haan MN, Shemanski L, Jagust WJ, Manolio TA, Kuller L. The role of apoE epsilon4 in modulating effects of other risk factors for cognitive decline in elderly persons. *Neuroimage.* 2018;172:118-29. doi: 10.1016/j.neuroimage.2017.12.027.
  90. Lahoz C, Schaefer EJ, Cupples LA, Wilson PW, Levy D, Osgood D, et al. Apolipoprotein E genotype and cardiovascular disease in the Framingham Heart Study. *Atherosclerosis.* 2001;154(3):529-37. doi: 10.1016/S0021-9150(00)00570-0
  91. Mahley RW, Rall SC Jr. Apolipoprotein E: far more than a lipid transport protein. *Annu Rev Genomics Hum Genet.* 2000;1(1):507-37. doi: 10.1146/annurev.genom.1.1.507
  92. Peila R, Rodriguez BL, Launer LJ. Type 2 diabetes, APOE gene, and the risk for dementia and related pathologies: The Honolulu-Asia Aging Study. *Diabetes.* 2002;51(4):1256-62. doi: 10.2337/diabetes.51.4.1256
  93. Corder EH, Saunders AM, Strittmatter WJ, Schmechel DE, Gaskell PC, Small GW, et al. Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science.* 1993;261(5123):921-3. doi: 10.1126/science.8346443.
  94. Farrer LA, Cupples LA, Haines JL, Hyman B, Kukull WA, Mayeux R, et al. Effects of age, sex, and ethnicity on the association between apolipoprotein E genotype and Alzheimer disease: a meta-analysis. *JAMA.* 1997;278(16):1349-56.
  95. Talbot C, Lendon C, Craddock N, Shears S, Morris JC, Goate A. Protection against Alzheimer's disease with apoE epsilon2. *Lancet.* 1994;1432-2. doi: 10.1016/S0140-6736(94)92557-7
  96. Harold D, Abraham R, Hollingworth P, Sims R, Gerrish A, Hamshere ML, et al. Erratum: Genome-wide association study identifies variants at CLU and PICALM associated with Alzheimer's disease (*Nature Genetics*)(2009) 41 (1088-1093). *Nat Genet.* 2009;41(10). doi: 10.1038/ng.440
  97. Lambert JC, Heath S, Even G, Campion D, Sleegers K, Hiltunen M, et al. Genome-wide association study identifies variants at CLU and CR1 associated with Alzheimer's disease. *Nat Genet.* 2009;41(10):1094. doi: 10.1038/ng.440
  98. Hauser PS, Ryan RO. Impact of apolipoprotein E on Alzheimer's disease. *Curr Alzheimer Res.* 2013;10(8):809-17. doi: 10.2174/15672050113109990156
  99. Glorioso CA, Pfenning AR, Lee SS, Bennett DA, Sibille EL, Kellis M, et al. Rate of brain aging and APOE  $\epsilon$ 4 are synergistic risk factors for Alzheimer's disease. *Life Sci Alliance.* 2019;2(3):e201900303. doi: 10.26508/lsa.201900303.
  100. Bales KR, Liu F, Wu S, Lin S, Koger D, DeLong C, et al. Human APOE isoform-dependent effects on brain  $\beta$ -amyloid levels in PDAPP transgenic mice. *J Neurosci.* 2009;29(21):6771-9. doi: 10.1523/JNEUROSCI.0887-09.2009
  101. Castellano JM, Kim J, Stewart FR, Jiang H, DeMattos RB, Patterson BW, et al. Human apoE isoforms differentially regulate brain amyloid- $\beta$  peptide clearance. *Sci Transl Med.* 2011;3(89):89ra57. doi: 10.1126/scitranslmed.3002156
  102. Huang Y-WA, Zhou B, Wernig M, S dnhof TC. ApoE2, ApoE3, and ApoE4 differentially stimulate APP transcription and A $\beta$  secretion. *Cell.* 2017;168(3):427-441.e421. doi: 10.1016/j.cell.2016.12.044
  103. Bales KR, Verina T, Dodel RC, Du Y, Altstiel L, Bender M et al. Lack of apolipoprotein E dramatically reduces amyloid  $\beta$ -peptide deposition. *Nat Genet.* 1997;17(3):263-4. doi: 10.1038/ng1197-263
  104. Kok E, Haikonen S, Luoto T, Huhtala H, Goebeler S, Haapasalo H, et al. Apolipoprotein E-dependent accumulation of Alzheimer disease-related lesions begins in middle age. *Ann Neurol.* 2009;65(6):650-7. doi: 10.1002/ana.21696
  105. Polvikoski T, Sulkava R, Haltia M, Kainulainen K, Vuorio A, Verkkoniemi A et al. Apolipoprotein E, dementia, and cortical deposition of  $\beta$ -amyloid protein. *N Engl J Med.* 1995;333(19):1242-8. doi: 10.1073/pnas.90.20.9649
  106. Schmechel D, Saunders A, Strittmatter W, Crain BJ, Hulette CM, Joo SH, et al. Increased amyloid beta-peptide deposition in cerebral cortex as a consequence of apolipoprotein E genotype in late-onset Alzheimer disease. *Proc Natl Acad Sci U S A.* 1993;90(20):9649-53. doi: 10.1073/pnas.90.20.9649
  107. Berlau DJ, Corrada MM, Head E, Kawas CH. APOE  $\epsilon$ 2 is associated with intact cognition but increased Alzheimer



- pathology in the oldest old. *Neurology*. 2009;72(9):829-34. doi: 10.1212/01.wnl.0000343853.00346.a4
108. Etnier JL, Caselli RJ, Reiman EM, Alexander GE, Sibley BA, Tessier D, et al. Cognitive performance in older women relative to ApoE-ε4 genotype and aerobic fitness. *Med Sci Sports Exerc*. 2007;39(1):199-207. doi: 10.1249/01.mss.0000239399.85955.5e
  109. Brown BM, Peiffer J, Taddei K, Lui JK, Laws SM, Gupta VB, et al. Physical activity and amyloid-β plasma and brain levels: results from the Australian Imaging, Biomarkers and Lifestyle Study of Ageing. *Mol Psychiatry*. 2013;18(8):875-881. doi: 10.1038/mp.2012.107
  110. Head D, Bugg JM, Goate AM, Fagan AM, Mintun MA, Benzinger J, et al. Exercise engagement as a moderator of the effects of APOE genotype on amyloid deposition. *Arch Neurol*. 2012;69(5):636-43. doi: 10.1001/archneurol.2011.845
  111. Berkowitz C, Mosconi L, Rahman A, Scheyer O, Hristov H, Isaacson RS. Clinical application of APOE in Alzheimer's prevention: a precision medicine approach. *J Prev Alzheimers Dis*. 2018;5(4):245-52. doi: 10.14283/jpad.2018.35
  112. Durazzo TC, Mattsson N, Weiner MW, Initiative AsDN. Interaction of cigarette smoking history with apoe genotype and age on amyloid level, glucose metabolism, and neurocognition in cognitively normal elders. *Nicotine Tob Res*. 2015;18(2):204-11. doi: 10.1093/ntr/ntv075
  113. Kivipelto M, Rovio S, Ngandu T, Kåreholt I, Eskelinen M, Winblad B, et al. Apolipoprotein E ε4 magnifies lifestyle risks for dementia: a population-based study. *J Cell Mol Med*. 2008;12(6b):2762-71. doi: 10.1111/j.1582-4934.2008.00296.x
  114. Downer B, Zanjani F, Fardo DW. The relationship between midlife and late life alcohol consumption, APOE ε4 and the decline in learning and memory among older adults. *Alcohol Alcohol*. 2014;49(1):17-22. doi: 10.1093/alcac/agt144
  115. Premkumar K, Abraham SK, Santhiya S, Ramesh A. Protective effects of saffron (*Crocus sativus* Linn.) on genotoxins-induced oxidative stress in Swiss albino mice. *Phytother Res*. 2003;17(6):614-17. doi: 10.1002/ptr.1209
  116. Soeda S, Aritake K, Urade Y, Sato H, Shoyama Y. Neuroprotective activities of saffron and crocin. *The Benefits of Natural Products for Neurodegenerative Diseases*: Springer; 2016:275-92. doi:10.1007/978-3-319-28383-8\_14
  117. Batish D, Singh H, Setia N, Kaur S, Kohli R. 2-Benzoxazolinone (BOA) induced oxidative stress, lipid peroxidation and changes in some antioxidant enzyme activities in mung bean (*Phaseolus aureus*). *Plant Physiol Biochem*. 2006;44(11-12):819-27. doi: 10.1016/j.plaphy.2006.10.014
  118. Bodnoff SR, Humphreys AG, Lehman JC, Diamond DM, Rose GM, Meaney MJ. Enduring effects of chronic corticosterone treatment on spatial learning, synaptic plasticity, and hippocampal neuropathology in young and mid-aged rats. *J Neurosci*. 1995;15(1):61-9. doi: 10.1523/JNEUROSCI.15-01-00061.1995
  119. Zafir A, Banu N. Modulation of in vivo oxidative status by exogenous corticosterone and restraint stress in rats. *Stress*. 2009;12(2):167-177. doi: 10.1080/10253890802234168
  120. Galea L, McEwen B, Tanapat P, Deak T, Spencer R, Dhabhar F. Sex differences in dendritic atrophy of CA3 pyramidal neurons in response to chronic restraint stress. *Neuroscience*. 1997;81(3):689-97. doi: 10.1016/s0306-4522(97)00233-9
  121. Magariños A, Orchinik M, McEwen BS. Morphological changes in the hippocampal CA3 region induced by non-invasive glucocorticoid administration: a paradox. *Brain Res*. 1998;809(2):314-8. doi: 10.1016/s0006-8993(98)00882-8
  122. Asdaq SMB, Inamdar MN. Potential of *Crocus sativus* (saffron) and its constituent, crocin, as hypolipidemic and antioxidant in rats. *Appl Biochem Biotechnol*. 2010;162(2):358-72. doi: 10.1007/s12010-009-8740-7
  123. Kanakis C, Tarantilis P, Pappas C, Bariyanga J, Tajmir-Riahi H, Polissiou M. An overview of structural features of DNA and RNA complexes with saffron compounds: Models and antioxidant activity. *J Photochem Photobiol B*. 2009;95(3):204-12. doi: 10.1016/j.jphotobiol.2009.03.006
  124. Das SK, Prusty A, Samantaray D, Hasan M, Jena S, Patra JK, et al. Effect of *Xylocarpus granatum* Bark Extract on Amelioration of Hyperglycaemia and Oxidative Stress Associated Complications in STZ-Induced Diabetic Mice. *Evid Based Complement Alternat Med*. 2019;2019:8493190. doi: 10.1155/2019/8493190.
  125. Aydin Dilsiz S, Bacanlı M, Anlar H, Çal T, Arı N, Ündeğer Bucurgat Ü, et al. Preventive role of Pycnogenol (R) against the hyperglycemia-induced oxidative stress and DNA damage in diabetic rats. *Food Chem Toxicol*. 2019 Feb;124:54-63. doi: 10.1016/j.fct.2018.11.038.
  126. Del-Angel D, Martínez N, Cruz M, Urrutia E, Riverón-Negrete L, Abdullaev F. Saffron extract ameliorates oxidative damage and mitochondrial dysfunction in the rat brain. Paper presented at: II International Symposium on Saffron Biology and Technology 7392006. doi: 10.17660/ActaHortic.2007.739.47
  127. Sh A, El-Azime A, NH S, N A E. Efficacy of aqueous extract of saffron (*Crocus sativus* L.) in modulating radiation-induced brain and eye retina damage in rats. *The Egyptian Journal of Hospital Medicine*. 2014;54(1):101-8. doi: 10.12816/0002436
  128. Finley JW, Gao S. A perspective on *Crocus sativus* L. (Saffron) constituent crocin: a potent water-soluble antioxidant and potential therapy for Alzheimer's disease. *J Agric Food Chem*. 2017;65(5):1005-20. doi: 10.1021/acs.jafc.6b04398
  129. Chen L, Qi Y, Yang X. Neuroprotective effects of crocin against oxidative stress induced by ischemia/reperfusion injury in rat retina. *Ophthalmic Res*. 2015;54(3):157-68. doi: 10.1159/000439026
  130. Rahiman N, Akaberi M, Sahebkar A, Emami SA, Tayarani-Najaran Z. Protective effects of saffron and its active components against oxidative stress and apoptosis in endothelial cells. *Microvasc Res*. 2018;118:82-9. doi: 10.1016/j.mvr.2018.03.003
  131. Ibach B, Haen E. Acetylcholinesterase inhibition in Alzheimer's Disease. *Curr Pharm Des*. 2004;10(3):231-51. doi: 10.2174/1381612043386509
  132. Mega MS. The cholinergic deficit in Alzheimer's disease: impact on cognition, behaviour and function. *Int J Neuropsychopharmacol*. 2000;3(Supplement\_2):S3-S12. doi: 10.1017/S1461145700001942
  133. Birks JS. Cholinesterase inhibitors for Alzheimer's disease. *Cochrane Database Syst Rev*. 2006;(1):CD005593. doi: 10.1002/14651858.CD005593.
  134. Howland RH. Drug therapies for cognitive impairment and dementia. *J Psychosoc Nurs Ment Health Serv*. Journal of psychosocial nursing and mental health services. 2010;48(4):11-4. doi: 10.3928/02793695-20100311-01
  135. Ademosun AO, Oboh G. Inhibition of acetylcholinesterase activity and Fe<sup>2+</sup>-induced lipid peroxidation in rat brain in vitro by some citrus fruit juices. *J Med Food*. 2012;15(5):428-34. doi: 10.1089/jmf.2011.0226
  136. Papandreou MA, Tsachaki M, Efthimiopoulos S, Cordopatis P, Lamari FN, Margariti M. Memory enhancing effects of saffron in aged mice are correlated with antioxidant protection. *Behav Brain Res*. 2011;219(2):197-204. doi: 10.1016/j.bbr.2011.01.007
  137. Rafieipour F, Hadipour E, Emami SA, Asili J, Tayarani-Najaran Z. Safranal protects against beta-amyloid peptide-induced cell toxicity in PC12 cells via MAPK and PI3 K pathways. *Metab Brain Dis*. 2019;34(1):165-72. doi: 10.1016/j.pbb.2015.10.011
  138. Rashedinia M, Lari P, Abnous K, Hosseinzadeh H. Protective effect of crocin on acrolein-induced tau phosphorylation in

- the rat brain. *Acta Neurobiol Exp (Wars)*. 2015;75(2):208-19.
139. Kirouac L, Rajic AJ, Cribbs DH, Padmanabhan J. Activation of Ras-ERK signaling and GSK-3 by amyloid precursor protein and amyloid beta facilitates neurodegeneration in Alzheimer's disease. *eNeuro*. 2017 Mar 27;4(2):ENEURO.0149-16.2017. doi: 10.1523/ENEURO.0149-16.2017.
  140. Ghahghaei A, Bathaie SZ, Bahraminejad E. Mechanisms of the effects of crocin on aggregation and deposition of A $\beta$ 1-40 fibrils in Alzheimer's disease. *Int J Pept Res The*. 2012;18(4):347-51. doi: 10.1007/s10989-012-9308-x
  141. Ghahghaei A, Bathaie SZ, Kheirkhah H, Bahraminejad E. The protective effect of crocin on the amyloid fibril formation of A $\beta$ 42 peptide in vitro. *Cell Mol Biol Lett*. 2013;18(3):328-39. doi: 10.2478/s11658-013-0092-1
  142. Morelli S, Salerno S, Piscioneri A, Tasselli F, Drioli E, De Bartolo L. Neuronal membrane bioreactor as a tool for testing crocin neuroprotective effect in Alzheimer's disease. *Chem Eng J*. 2016;305:69-78. doi:10.1016/j.cej.2016.01.035
  143. Asadi F, Jamshidi AH, Khodagholi F, Yans A, Azimi L, Faizi M, et al. Reversal effects of crocin on amyloid  $\beta$ -induced memory deficit: modification of autophagy or apoptosis markers. *Pharmacol Biochem Behav*. 2015;139(Pt A):47-58. doi: 10.1016/j.pbb.2015.10.011.
  144. Soeda S, Ochiai T, Paopong L, Tanaka H, Shoyama Y, Shimeno H. Crocin suppresses tumor necrosis factor- $\alpha$ -induced cell death of neuronally differentiated PC-12 cells. *Life Sci*. 2001;69(24):2887-98. doi: 10.1016/s0024-3205(01)01357-1
  145. Baluchnejadmojarad T, Mohamadi-Zarch S-M, Roghani M. Safranal, an active ingredient of saffron, attenuates cognitive deficits in amyloid  $\beta$ -induced rat model of Alzheimer's disease: underlying mechanisms. *Metab Brain Dis*. 2019;34(6):1747-59. doi: 10.1007/s11011-019-00481-6
  146. Holsinger RD, McLean CA, Beyreuther K, Masters CL, Evin G. Increased expression of the amyloid precursor  $\beta$ -secretase in Alzheimer's disease. *Ann Neurol*. 2002;51(6):783-6. doi: 10.1002/ana.10208
  147. Kaye R, Lasagna-Reeves CA. Molecular mechanisms of amyloid oligomers toxicity. *J Alzheimers Dis*. 2013;33(s1):S67-S78. doi: 10.3233/JAD-2012-129001
  148. Batarseh YS, Bharate SS, Kumar V, Kumar A, Vishwakarma RA, Bharate SB, et al. *Crocus sativus* extract tightens the blood-brain barrier, reduces amyloid  $\beta$  load and related toxicity in 5XFAD mice. *ACS Chem Neurosci*. 2017;8(8):1756-66. doi: 10.1021/acscchemneuro.7b00101
  149. Karakani AM, Riazi G, Mahmood Ghaffari S, Ahmadian S, Mokhtari F, Jalili Firuzi M, et al. Inhibitory effect of corcin on aggregation of 1N/4R human tau protein in vitro. *Iran J Basic Med Sci*. 2015;18(5):485-92.
  150. Mohammadzadeh L, Abnous K, Razavi BM, Hosseinzadeh H. Crocin-protected malathion-induced spatial memory deficits by inhibiting TAU protein hyperphosphorylation and antiapoptotic effects. *Nutr Neurosci*. 2020;23(3):221-236. doi: 10.1080/1028415X.2018.1492772
  151. Chawla A, Boisvert WA, Lee CH, Laffitte BA, Barak Y, Joseph SB, et al. A PPAR $\gamma$ -LXR-ABCA1 pathway in macrophages is involved in cholesterol efflux and atherogenesis. *Mol Cell*. 2001;7(1):161-71. doi: 10.1016/s1097-2765(01)00164-2
  152. Qosa H, Abuasal BS, Romero IA, Weksler B, Couraud PO, Keller JN, et al. Differences in amyloid- $\beta$  clearance across mouse and human blood-brain barrier models: kinetic analysis and mechanistic modeling. *Neuropharmacology*. 2014;79:668-78. doi: 10.1016/j.neuropharm.2014.01.023
  153. Wahrle SE, Jiang H, Parsadanian M, Han X, Fryer JD, Kowalewski T, et al. ABCA1 is required for normal central nervous system apoE levels and for lipidation of astrocyte-secreted apoE. *J Biol Chem*. 2004;279(39):40987-93. doi: 10.1074/jbc.M407963200
  154. Akhondzadeh S, Sabet MS, Harirchian MH, Gougol A, Yekehtaz H, Alimardani R, et al. A 22-week, multicenter, randomized, double-blind controlled trial of *Crocus sativus* in the treatment of mild-to-moderate Alzheimer's disease. *Hum Psychopharmacol*. 2010b;207(4):637-43. doi: 10.1002/hup.2412
  155. Farokhnia M, Shafiee Sabet M, Iranpour N, Gougol A, Yekehtaz H, Alimardani R, et al. Comparing the efficacy and safety of *Crocus sativus* L. with memantine in patients with moderate to severe Alzheimer's disease: a double-blind randomized clinical trial. *Hum Psychopharmacol*. 2014;29(4):351-59. doi: 10.1002/hup.2412
  156. Akhondzadeh S, Sabet MS, Harirchian M, Togha M, Cheraghmakani H, Razeghi S, et al. Saffron in the treatment of patients with mild to moderate Alzheimer's disease: a 16-week, randomized and placebo-controlled trial. *J Clin Pharm Ther*. 2010a;35(5):581-8. doi: 10.1111/j.1365-2710.2009.01133.x
  157. Steiner GZ, Yeung A, Liu JX, Camfield DA, Blasio FM, Pipingas A, et al. The effect of Sailuotong (SLT) on neurocognitive and cardiovascular function in healthy adults: a randomised, double-blind, placebo controlled crossover pilot trial. *BMC Complement Altern Med*. 2015;16(1):15. doi: 10.1186/s12906-016-0989-0
  158. Tsolaki M, Karathanasi E, Lazarou I, Dovas K, Verykoui E, Karacostas A, et al. Efficacy and safety of *Crocus sativus* L. in patients with mild cognitive impairment: one year single-blind randomized, with parallel groups, clinical trial. *J Alzheimers Dis*. 2016;54(1):129-33. doi: 10.3233/JAD-160304
  159. Dashti-r M, Zeinali F, Anvari M, Hosseini S. Saffron (*Crocus sativus* L.) extract prevents and improves D-galactose and NaNO<sub>2</sub> induced memory impairment in mice. *EXCLI J*. 2012;11:328.
  160. Ochiai T, Shimeno H, Mishima K-i, Iwasaki K, Fujiwara M, Tanaka H, et al. Protective effects of carotenoids from saffron on neuronal injury in vitro and in vivo. *Biochim Biophys Acta*. 2007;1770(4):578-84. doi: 10.1016/j.bbagen.2006.11.012
  161. Tashakori-Sabzevar F, Hosseinzadeh H, Sadat Motamedshariaty V, Reza Movassaghi A, Ahmad Mohajeri S. Crocetin attenuates spatial learning dysfunction and hippocampal injury in a model of vascular dementia. *Curr Neurovasc Res*. 2013;10(4):325-334. doi: 10.2174/15672026113109990032
  162. Hosseinzadeh H, Sadeghnia HR, Ghaeni FA, Motamedshariaty VS, Mohajeri SA. Effects of saffron (*Crocus sativus* L.) and its active constituent, crocin, on recognition and spatial memory after chronic cerebral hypoperfusion in rats. *Phytother Res*. 2012;26(3):381-386. doi: 10.1002/ptr.3566
  163. Zheng YQ, Liu JX, Wang JN, Xu L. Effects of crocin on reperfusion-induced oxidative/nitrative injury to cerebral microvessels after global cerebral ischemia. *Brain Res*. 2007;1138:86-94. doi: 10.1016/j.brainres.2006.12.064
  164. Khalili M, Hamzeh F. Effects of active constituents of *Crocus sativus* L., crocin on streptozocin-induced model of sporadic Alzheimer's disease in male rats. *Iran Biomed J*. 2010;14(1-2):59.
  165. Naghizadeh B, Mansouri M, Ghorbanzadeh B, Farbood Y, Sarkaki A. Protective effects of oral crocin against intracerebroventricular streptozotocin-induced spatial memory deficit and oxidative stress in rats. *Phytomedicine*. 2013;20(6):537-42. doi: 10.1016/j.phymed.2012.12.019
  166. amaddonfard E, Farshid AA, Asri-Rezaee S, Javadi S, Khosravi V, Rahman B, et al. Crocin improved learning and memory impairments in streptozotocin induced diabetic rats. *Iran J Basic Med Sci*. 2013;16(1):91-100.
  167. Moshahid Khan M, Ahmad A, Yusuf S, Islam F. Neuroprotective efficacy of *Nardostachys jatamansi* and crocetin in conjunction with selenium in cognitive impairment. *Neurol Sci*. 2012;33(5):1011-20. doi: 10.1007/s10072-011-0880-1.
  168. Bandegi AR, Rashidy-Pour A, Vafaei AA, Ghadrdoost B. Protective effects of *Crocus sativus* L. extract and crocin

- against chronic-stress induced oxidative damage of brain, liver and kidneys in rats. *Adv Pharm Bull.* 2014;4(Suppl 2):493-9. doi: 10.5681/apb.2014.073.
169. Ghadrdoost B, Vafaei AA, Rashidy-Pour A, Hajisoltani R, Bandegi AR, Motamedi F, et al. Protective effects of saffron extract and its active constituent crocin against oxidative stress and spatial learning and memory deficits induced by chronic stress in rats. *Eur J Pharmacol.* 2011;667(1-3):222-9. doi: 10.1016/j.ejphar.2011.05.012.
  170. El-Beshbishy HA, Hassan MH, Aly HA, Doghish AS, Alghaithy AA. Crocin "saffron" protects against beryllium chloride toxicity in rats through diminution of oxidative stress and enhancing gene expression of antioxidant enzymes. *Ecotoxicol Environ Saf.* 2012;83:47-54. doi: 10.1016/j.ecoenv.2012.06.003
  171. Pitsikas N, Zisopoulou S, Tarantilis PA, Kanakis CD, Polissiou MG, Sakellariadis N. Effects of the active constituents of *Crocus sativus* L., crocins on recognition and spatial rats' memory. *Behav Brain Res.* 2007;183(2):141-6. doi: 10.1016/j.bbr.2007.06.001
  172. Pitsikas N, Sakellariadis N. *Crocus sativus* L. extracts antagonize memory impairments in different behavioural tasks in the rat. *Behav Brain Res.* 2006;173(1):112-5. doi: 10.1016/j.bbr.2006.06.005
  173. Zhang Y, Shoyama Y, Sugiura M, Saito H. Effects of *Crocus sativus* L. on the ethanol-induced impairment of passive avoidance performances in mice. *Biol Pharm Bull.* 1994;17(2):217-21. doi: 10.1248/bpb.17.217
  174. Sugiura M, Shoyama Y, Saito H, Nishiyama N. Crocin improves the ethanol-induced impairment of learning behaviors of mice in passive avoidance tasks. *Proceedings of the Japan Academy, Series B.* 1995;71(10):319-24. doi:10.2183/pjab.71.319
  175. Abe K, Sugiura M, Yamaguchi S, Shoyama Y, Saito H. Saffron extract prevents acetaldehyde-induced inhibition of long-term potentiation in the rat dentate gyrus in vivo. *Brain Res.* 1999;851(1-2):287-9. doi: 10.1016/s0006-8993(99)02174-5
  176. Walton J, Wang MX. APP expression, distribution and accumulation are altered by aluminum in a rodent model for Alzheimer's disease. *J Inorg Biochem.* 2009;103(11):1548-54. doi: 10.1016/j.jinorgbio.2009.07.027
  177. Xu Y, Wang Z, You W, Zhang X, Li S, Barish PA, et al. Antidepressant-like effect of trans-resveratrol: involvement of serotonin and noradrenaline system. *Eur Neuropsychopharmacol.* 2010;20(6):405-13. doi: 10.1016/j.euroneuro.2010.02.013
  178. Yokel RA. Blood-brain barrier flux of aluminum, manganese, iron and other metals suspected to contribute to metal-induced neurodegeneration. *J Alzheimers Dis.* 2006;10(2-3):223-53. doi: 10.3233/jad-2006-102-309
  179. Zhang ZJ, Qian YH, Hu HT, Yang J, Yang GD. The herbal medicine *Dipsacus asper* Wall extract reduces the cognitive deficits and overexpression of  $\beta$ -amyloid protein induced by aluminum exposure. *Life Sci.* 2003;73(19):2443-54. doi: 10.1016/s0024-3205(03)00649-0
  180. Sethi P, Jyoti A, Singh R, Hussain E, Sharma D. Aluminium-induced electrophysiological, biochemical and cognitive modifications in the hippocampus of aging rats. *Neurotoxicology.* 2008;29(6):1069-79. doi: 10.1016/j.neuro.2008.08.005
  181. Sharma D, Sethi P, Hussain E, Singh R. Curcumin counteracts the aluminium-induced ageing-related alterations in oxidative stress, Na<sup>+</sup>, K<sup>+</sup> ATPase and protein kinase C in adult and old rat brain regions. *Biogerontology.* 2009;10(4):489-502. doi: 10.1007/s10522-008-9195-x
  182. Tripathi S, Mahdi AA, Nawab A, Chander R, Hasan M, Siddiqui MS, Mahdi F, et al. Influence of age on aluminum induced lipid peroxidation and neurolipofuscin in frontal cortex of rat brain: a behavioral, biochemical and ultrastructural study. *Brain Res.* 2009;1253:107-16. doi:10.1016/j.brainres.2008.11.060
  183. Butterfield DA, Abdul HM, Newman S, Reed T. Redox proteomics in some age-related neurodegenerative disorders or models thereof. *NeuroRx.* 2006;3(3):344-57. doi: 10.1016/j.nurx.2006.05.003
  184. Murali G, Panneerselvam C. Age-associated oxidative macromolecular damages in rat brain regions: role of glutathione monoester. *J Gerontol A Biol Sci Med Sci.* 2007;62(8):824-30. doi: 10.1093/gerona/62.8.824
  185. Forster MJ, Dubey A, Dawson KM, Stutts WA, Lal H, Sohal RS. Age-related losses of cognitive function and motor skills in mice are associated with oxidative protein damage in the brain. *Proc Natl Acad Sci U S A.* 1996;93(10):4765-9. doi: 10.1073/pnas.93.10.4765
  186. Gorini A, Ghigini B, Villa R. Acetylcholinesterase activity of synaptic plasma membranes during ageing: effect of L-acetylcarnitine. *Demantia.* 1996;7(3):147-54. doi: 10.1159/000106870
  187. Geromichalos GD, Lamari FN, Papandreou MA, Trafalis DT, Margarity M, Papageorgiou A, et al. Saffron as a source of novel acetylcholinesterase inhibitors: molecular docking and in vitro enzymatic studies. *J Agric Food Chem.* 2012;60(24):6131-8. doi: 10.1021/jf300589c
  188. Ochiai T, Ohno S, Soeda S, Tanaka H, Shoyama Y, Shimeno H. Crocin prevents the death of rat pheochromyctoma (PC-12) cells by its antioxidant effects stronger than those of  $\alpha$ -tocopherol. *Neurosci Lett.* 2004;362(1):61-4. doi: 10.1016/j.neulet.2004.02.067
  189. Mehri S, Abnous K, Mousavi SH, Shariaty VM, Hosseinzadeh H. Neuroprotective effect of crocin on acrylamide-induced cytotoxicity in PC12 cells. *Cell Mol Neurobiol.* 2012;32(2):227-35. doi: 10.1007/s10571-011-9752-8
  190. Mousavi SH, Tayarani N, Parsaee H. Protective effect of saffron extract and crocin on reactive oxygen species-mediated high glucose-induced toxicity in PC12 cells. *Cell Mol Neurobiol.* 2010;30(2):185-91. doi: 10.1007/s10571-009-9441-z
  191. Kannan K, Jain SK. Oxidative stress and apoptosis. *Pathophysiology.* 2000;7(3):153-63. doi: 10.1016/s0928-4680(00)00053-5
  192. Scorrano L, Korsmeyer SJ. Mechanisms of cytochrome c release by proapoptotic BCL-2 family members. *Biochem Biophys Res Commun.* 2003;304(3):437-44. doi: 10.1016/s0006-291x(03)00615-6
  193. Zhang G-F, Zhang Y, Zhao G. Crocin protects PC12 cells against MPP<sup>+</sup>-induced injury through inhibition of mitochondrial dysfunction and ER stress. *Neurochem Int.* 2015;89:101-10. doi: 10.1016/j.neuint.2015.07.011
  194. Nam KN, Park YM, Jung HJ, Lee JY, Min BD, Park SU, et al. Anti-inflammatory effects of crocin and crocetin in rat brain microglial cells. *Eur J Pharmacol.* 2010;648(1-3):110-16. doi: 10.1016/j.ejphar.2010.09.003
  195. Hosseinzadeh H, Shakib SS, Sameni AK, Taghiabadi E. Acute and subacute toxicities of safranal, a constituent of saffron, in mice and rats. *Iran J Pharm Res.* 2011;13(44):S122.
  196. Hosseinzadeh H, Shariaty VM, Sameni A. Acute and sub-acute toxicity of crocin, a constituent of *Crocus sativus* L. (Saffron), in mice and rats. *January 2010.*
  197. Mohajeri D, Mousavi G, Mesgari M, Doustar Y, Khayat Nouri M. Subacute toxicity of *Crocus sativus* L. (saffron) stigma ethanolic extract in rats. *Am J Pharmacol Toxicol.* 2007;2(4):189-93. doi: 10.3844/ajptsp.2007.189.193
  198. Khayatnouri M, Safavi SE, Safarmashaei S, Babazadeh D, Mikailpourardabili B. The effect of saffron orally administration on spermatogenesis index in rat. *Adv Environ Biol.* 2011;5:1514-21.
  199. Taheri F, Zahra Bathaie S, Ashrafi M, Ghasemi E. Assessment of Crocin Toxicity on the Rat Liver. *Modares Journal of Medical Sciences: Pathobiology.* 2014;17(3).
  200. Riahi-Zanjani B, Balali-Mood M, Mohammadi E, Badie-Bostan

- H, Memar B, Karimi G. Safranal as a safe compound to mice immune system. *Avicenna J Phytomed*. 2015;5(5):441.
201. Muosa F, AL-Rekabi K, Askar S, Yousif E. Evaluation of the toxic effect of ethanolic extract of saffron in male mice after subchronic exposure. *Donnish J Pharm Pharmacol*. 2015;1:1-7.
202. Bostan HB, Mehri S, Hosseinzadeh H. Toxicology effects of saffron and its constituents: a review. *Iran J Basic Med Sci*. 2017;20(2):110. doi: 10.22038/ijbms.2017.8230
203. Khan M, Hanif MA, Ayub MA, Jilani MI, Chatha SAS. Saffron. *Medicinal Plants of South Asia: Elsevier*; 2020:587-600.
204. Modagheh M-H, Shahabian M, Esmaili HA, Rajbai O, Hosseinzadeh H. Safety evaluation of saffron (*Crocus sativus*) tablets in healthy volunteers. *Phytomedicine*. 2008;15(12):1032-7. doi: 10.1016/j.phymed.2008.06.003
205. Ayatollahi H, Javan AO, Khajedaluee M, Shahroodan M, Hosseinzadeh H. Effect of *Crocus sativus* L.(saffron) on coagulation and anticoagulation systems in healthy volunteers. *Phytother Res*. 2014; 28(4):539-43. doi 10.1002/ptr.5021
206. Mohamadpour AH, Ayati Z, Parizadeh MR, Rajbai O, Hosseinzadeh H. Safety evaluation of crocin (a constituent of saffron) tablets in healthy volunteers. *Iran J Basic Med Sci*. 2013;16(1):39.
207. Ajam M, Reyhani T, Roshanravan V, Zare Z. Increased miscarriage rate in female farmers working in saffron fields: a possible effect of saffron toxicity. *Asia Pac J Med Toxicol*. 2014;3(2):73-5. doi: 10.22038/apjmt.2014.3047
208. Sadraei H, Ghannadi A, Takei-bavani M. Effects of *Zataria multiflora* and *Carum carvi* essential oils and hydroalcoholic extracts of *Passiflora incarnata*, *Berberis integerrima* and *Crocus sativus* on rat isolated uterus contractions. *International Journal of Aromatherapy*. 2003;13(2-3):121-7. doi: 10.1016/S0962-4562(03)00092-4
209. Inoue E, Shimizu Y, Shoji M, Tsuchida H, Sano Y, Ito C. Pharmacological properties of N-095, a drug containing red ginseng, polygala root, saffron, antelope horn and aloe wood. *Am J Chin Med*. 2005; 33(01):49-60. doi 10.1142/S0192415X05002655.
210. Ahmadi S, Azhari S, Jafarzadeh H, Rakhshandeh H, Mazlom R. The effect of oral capsules of saffron on anxiety and fatigue during the first stage of labor. *SSU\_Journals*. 2015;23(2):1915-26.
211. Ahmadi S, Azhari S, Rakhshandeh H, Jaafarzadeh H, Mazlum S. Evaluation of the effect of saffron oral capsules on duration of the active phase of labor first stage. *Iran J Reprod Med*. 2014;12(6):44.
212. Kamalipour M, Akhondzadeh S. Cardiovascular effects of saffron: an evidence-based review. *Journal of Tehran Heart Center*. 2011 6(2):59-61.