



TENT: A Rapid DNA Extraction Method of *Staphylococcus aureus*

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Dear Editor-in-Chief

Recently different methods are introduced for DNA extraction from bacteria. Bacterial DNA can be extracted using a commercial kit and conventional (e.g., boiling, phenol-chloroform and detergent methods). In the manual methods, chemically solution such as EDTA, SDS, NaCl, Tris hydrochloride, phenol-chloroform, proteinase K and lysozyme can be proposed (1).

The phenol-chloroform DNA extraction method is time-consuming and manipulation of toxic solvents may be hazardous to the environment and the technician. Furthermore, several washing and centrifugation steps increases the risk of sample contamination (2). Therefore, several methods have been introduced as alternative techniques to the phenol-chloroform method including boiling and detergent methods. These approaches are convenient and low-cost, but due to remaining of some residual protein with the DNA, the purity of the extracted DNA is poor. Commercial DNA extraction kits offer a low risk of contamination and they are faster than conventional protocols, but the amount of DNA recovered is highly variable (3). We examined 'TENT' (Tris-EDTA-NaCl-TritonX100) buffer for DNA extraction from *Staphylococcus aureus* and DNA purity compared to detergent and kit methods.

In this study, *S. aureus* ATCC 29247 was cultured on tryptic soy agar (TSA) at 37 °C for 24 h and used in different DNA extraction methods. Pure colonies were suspended in 300 µL of TENT buffer (10 mM Tris-HCl, 0.1 M NaCl, 1 mM EDTA, 5% [v/v] Triton X100, pH 8.0). The cell suspension was boiled at 100 °C and then centrifuged. Supernatant fluid was transferred into a new sterile tube. Subsequently, cold 95% ethanol was added to the supernatant and kept at -20 °C for 20 min. After this stage, the solution was centrifuged. DNA template was dissolved in 50 µL sterile distilled water and stored at -20 °C until PCR amplification. This step was repeated three times and the result were compared.

Manual genomic DNA extraction of *S. aureus* was performed using detergent method (4). Genomic DNA extract by a kit, the product of the Viogene Company (UK) according to the manufacturer's instructions. Like before, this step was repeated three times. Purity of extracted DNA was determined based on measurement of OD $\frac{260}{280}$ using Nano Drop instrument (Thermo Scientific). Additionally, the integrity of extracted DNA was evaluated by electrophoresis in 0.8% agarose gel. To determine any inhibitors remained from the extraction procedure, PCR amplification of *mecA*

was conducted. The amplified PCR products were electrophoresed in 1% agarose gel at 120 V for 1 h and lastly stained with KBC (0.5 µg/ml) (Kowsar, Iran) and photographed under UV light. Concentration and purity of the extracted DNAs was evaluated using Nanodrop (NanoDrop Technologies and the results are shown in Table

1. Electrophoresis of extracted DNAs on 1% agarose gel (Fig. 1) showed that all of the extracted DNAs by three methods have high integrity and perfection. The results of PCR products amplified from extracted DNA using three methods showed that the amplification of *mecA* gene was successful (Fig. 2).

Table 1: Performance results of different methods tested based on purity factor and concentration of extracted DNA

Method	Purity factor1 (A260/280)	Concentration (mg/µL)
TENT	18.35 ± 8.55	919.0
Detergent	5.78 ± 2.89	286.3
Kit	1.40 ± 0.68	70.1

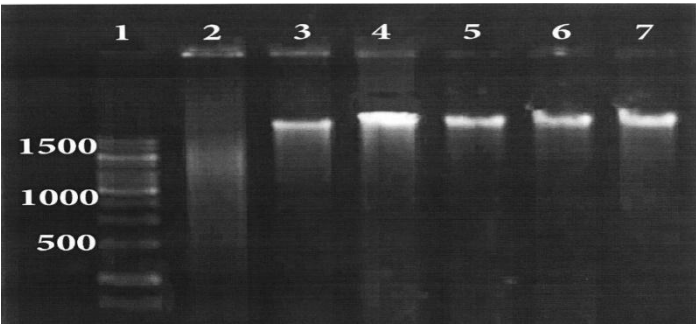


Fig. 1: Analysis of extracted genomic DNA from *S.aureus* on 0.1% agarose gel. Lane 1 is DNA size marker; lane 2, control negative; lane 3, extracted DNA using kit; lane 4 and 5, extracted DNA using TENT; lane 6, 7 extracted DNA using detergent

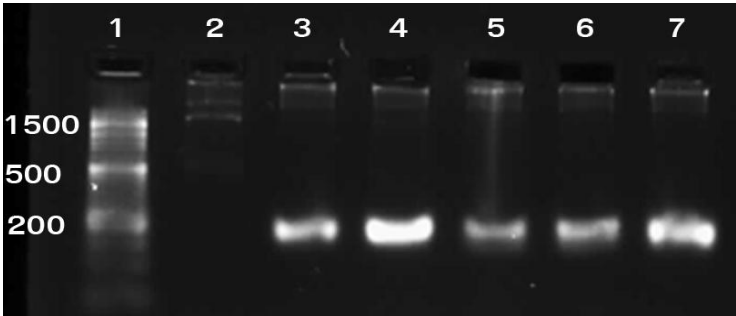


Fig. 2: Agarose gel electrophoresis of the PCR-amplified *mecA* gene. Lanes: 1: 50-bp ladder; 2: Negative control (*S. aureus* ATCC 8325-4); 3: Positive control (*S. aureus* strain COL); 4-7: *S. aureus* isolates showing 162 bp *mecA* amplicon.

In many studies, various methods including phenol-chloroform, detergent and commercial kits have been conducted for extraction DNA. Each of these methods has advantages and disadvantages. The advantages of phenol-chloroform approach can be pointed to the high purity extracted DNA but it is time-consuming and unsafe to user. Detergent and kit methods are more convenient and faster, but in these methods the purity of the extracted DNA may be low (5). In current study, the highest concentration of the extracted DNA was associated to TENT method (919µg/µl). This difference in concentration of DNA may be attributed using Triton x100 along

boiling that cause complete lysis of cell wall. Recently, similar buffer solution has been applied for other bacteria, including *Clostridium perfringens*, *Listeria monocytogenes* and *Nocardia* species (6-7). In our study, TENT buffer showed acceptable and satisfactory results for molecular assay such as PCR. The described method is simple, fast, cost-effective, sensitive, and highly reproducible for DNA extraction from *S. aureus*, and there is no need for a skillful specialist to perform this method as well as DNA concentration is higher than commercial kits or detergent methods.

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