

pH

*

/ / : / / :

Study of pH gradient on liposomes trapping efficiency by spectrophotometry: using neutral red

Mohammadi-Bardbori A.*

Faculty of Pharmacy, Shiraz University of Medical Sciences

Received: 2006/7/24 , Accepted: 2007/1/1

Objectives: The purpose of this study was to establish a new experimental, rapid, simple and cost effective approach to study liposomes. NR (Neutral red) was selected as a model of lipophilic and hydrophilic drug. NR is an indicator of pH and a weak base with $pK_a=6.7$ which is red while ionized in acidic media but yellow while non-ionized in alkaline media and is used in vital staining. **Methods:** In this study the effects of pH internal (5) and external media (5- 9) on liposomes trapping efficiencies in wavelength of 533nm were examined. **Results:** %EE is (the entrapment efficiency of liposome) increased between (%24-%87) in pH (external media) range 5-9. CBC (CBC, contrast between liposomal compartment for visualized by LM) and EBC (EBC, exchanged between liposomal compartment) and EE were calculated. Liposomes with the internal (pH = 5) and external media (pH = 9) is the most suitable option for staining with NR. No significant correlations were observed among the two index with a 95% coefficient interval (EBC with CBC, $r^2 = 0.93$, $p<0.05$, and EE with CBC $r^2=0.93$, $p<0.05$). Significant correlations were also observed among the EBC index and EE index with a 95% coefficient interval (EBC with EE $r^2=0.66$, $p<0.05$). **Conclusion:** NR accumulates in liposomes in ionized form therefore, giving sharp contrast with the surrounding media of liposomes. This makes them to be visualized by LM (light microscope).

Keywords: Liposome; Molecular NR; Ionic NR; Trapping efficiency; spectrophotometry.

() EE () pH () pH = () CBC () EBC () EBC () EE EBC () CBC EE () CBC pH () pH = () CBC EE () CBC

*Corresponding Author: Dr. Afshin Mohammadi-Bardbori, Associate Professor, Faculty of Pharmacy, Shiraz University of Medical Sciences, Tel: 0711-2424127; Fax: 0711-2424128; E-mail: toxicology@sums.ac.ir

mmol/L ()

(pH= pH= /) pH

mmol mmol/L NaCl

°C .() pH

pH .() pH

/M () .()

(pH =) (/ mg/ml) .()

:

pH () ()

() .()

:

Leitz .()

× / × pH ()

:

(EE) .()

/ / ml pH

/ rpm

EE=Qt-Qs/Qt

Qt EE

Qs

:

(Alabastar, AL) Avanti Polar Lipid (PC)

(Milwaukee) (CHOL) %

HPLC

:

(SEM) ±

ANOVA

Student - Newman Kules .()

SPSS /



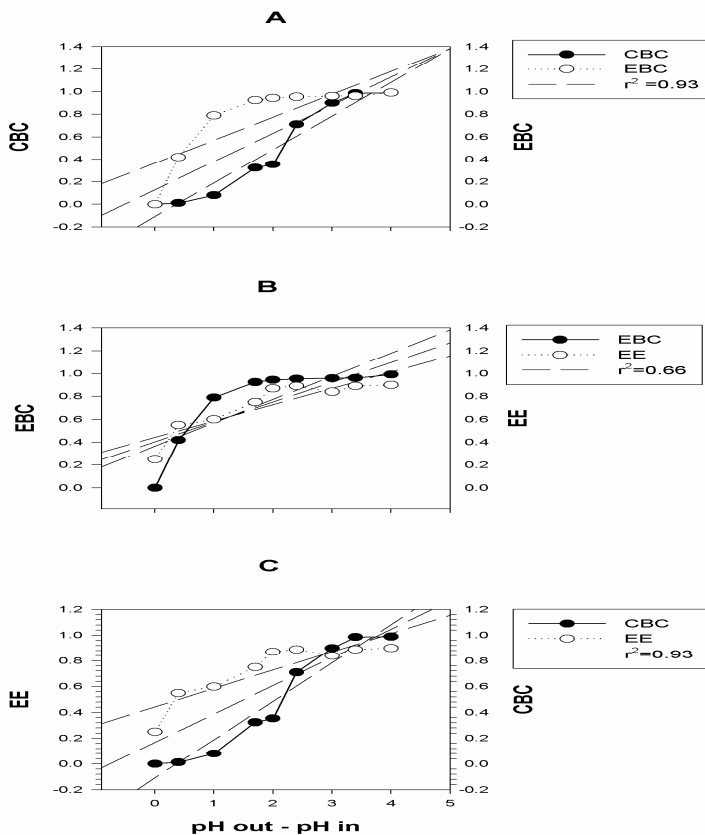
EBC = %up (%UP, percentage of unprotonated NR) out - %up in / %up total

EBC / pH
 EE EBC CBC
 %

$Pka-PH = \text{Log } \frac{\text{protonated NR}}{\text{Unprotonated NR}}$
 () ()
 ()
 (CBC)

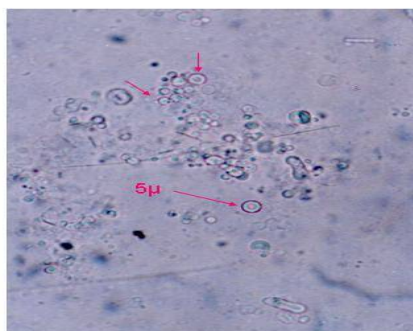
CBC EBC
 (r = / , p < /) CBC EE (r = / , p < /)
 EE EBC
 .C, B, A (r = / , p < /)

CBC=%p (%p, percentage of protonated NR) in-%Pout / %P total
 pH pH
 .()
 pH
 pH



(CBC, EBC and EE) (C B,A) :

pH in	pH out	%p in	%p out	%up in	%up out	CBC	EBC	EE
5	5	98.04	98.04	1.95	1.95	0.000	0.000	0.25
5	5.4	98.04	95.23	1.95	4.77	.0114	0.419	0.55
5	6	98.04	83.36	1.95	16.6	0.080	0.789	0.6
5	6.7	98.04	50	1.95	50	0.324	0.924	0.75
5	7	98.04	33.44	1.95	66.5	0.354	0.943	0.87
5	7.4	98.04	16.6	1.95	83.36	0.710	0.954	0.75
5	8	98.04	4.78	1.95	95.2	0.900	0.959	0.84
5	8.4	98.04	1.56	1.95	98.43	0.986	0.961	0.89
5	9	98.04	.49	1.95	99.5	0.990	0.990	0.90



Leitz () × / × pH pH :

pH pH Pka = /
% / % % /
% /

(.) pH pH

(μ) μ / μ

.() pH

 μ

.()

6- References:

1. budavari S. The Merck Index, 12th, Whitehouse station, NJ: Merck & Co. U.S.A., 1996.
2. Allen T.M. Liposomal drug delivery curr. Opin. Colloid Interface. Sci., 1996, 1, 645-651.
3. Mastrobattista E., Koning G.A., Storm G. Immunoliposomes for the targeted delivery of antitumor drugs. Adv. Drug Delive.Rev., 1999,40, 103-127.
4. Maruyama K. In vivo targering by liposome. Biol. Pharm. Bull., 2000, 23, 791-799.
5. Reddy J.A., Low P.S. Folate-mediated targeting of therapeutic and imaging agents to cancers.Crit. Rev. Ther. Drug, 1998, 15, 587-627.
6. Jain R.K. Transport of molecules in the tumor interstitium: a review, cancers, 1987, 47, 3039-3051.
7. Jain N. Yang., G. Tabibi., E.S. Yalkowsky, S.H. Solubilization of NSC-639829. Int. J. Pharm., 2001, 225, 41-47.
8. Harrigan P.R., Wong K.F., Redelmeier T.E.J., Wheeler J., Cullis P.R. Accumulation of doxorubicin and other lipophilic amines into large unilamellar vesicles in response to transmembrane pH gradients. Biochim. Biophys. Acta., 1993, 149, 329 – 338.
9. Choi M.J., Han H.S., Kim H. pH-sensitive liposomes containing polymerized phosphatidylethanolamine and fatty acid. J. Biochem., 1992,112, 694-699.
10. Bangham A.D., Standish M.M., Watkin S.J.C. J. Diffusion of univalent ions across the lamellae of swollen phospholipids.Mol. Biol., 1965, 13, 238-252.
11. Mozafari M.R., Reed C.J., Rostron C., Kocum C., Piskin E. Construction of stable anionic liposome-plasmid particles using the heating method: A preliminary investigation. Cell. Mol. Biol. Lett., 2002, 7, 923-927.
12. Aurora T.S., Cummins L.W., Haines H.Z. Preparation and characterization of monodisperse unilamellar phospholipid vesicles with selected diameters of from 300 to 600 nm.. Biochim.Biophys. Acta., 1985, 820,250-258.
13. Brand M.: a technological approach. Biotechnology Annual Review, 2001, 759-85.
14. L'opez-Pinto J.M., Gonz'alez-Rodr'iguez M.L., Rabasco A.M. Effect of cholesterol and ethanol on dermal delivery from DPPC liposomes. Int J Pharm., 2005, 298, 1-12.
15. Moscho A., Orwar O., Chiu D.T., Modi B.P., Zare R.N. Rapid preparation of giant unilamellar vesicles. Proc Natl Acad Sci U S A., 1996, 93, 1443-7.
16. Coderch L., Oliva M., Pons M., Maza Ade. la. Manich., A.M. Parra., J.L. The Effect of Liposomes on Skin Barrier Structure. Int. J. Pharm., 1996, 139, 197–203.